

Supporting Information

Enantioselective Copper(I)/Chiral Phosphoric Acid Catalyzed Intramolecular Amination of Allylic and Benzylic C–H Bonds

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Supporting Information

Table of contents

Tables S1-S4, Figures S1-S3, Schemes S1-S2	S2
General Information	S13
Experimental Section and Characterization Data of Substrates	S13
Procedure for Enantioselective Radical Allylic C–H Amination under Conditions A and Conditions B	S35
Characterization Data of Products 2	S36
Procedure for Enantioselective Radical Benzylic C–H Amination	S41
Characterization Data of Products 4	S43
Preparative Scale Reaction and Transformation	S53
Mechanistic Studies	S55
References	S61
NMR Spectra of new compounds	S62
HPLC spectra of products	S115

Table S1 Optimization of reaction conditions of enantioselective allylic C–H bond amination^a

<p>(<i>R</i>)-C3: Ar = 4-CF₃-Ph</p>		<p>Oxidant:</p> <p>R = cyclopentyl, O2</p> <p>R = Me, O3 R = Et, O4</p>		<p>PG =</p> <p>PG =</p> <p>4-OMe-NHPI</p>			
entry	[Cu]	CPA	R-NHPI	oxidant	additive	yield(%) ^b	er ^c
1	CuTc	(<i>R</i>)- C1	---	O2 , O1 etc	---	0	N.A.
2	CuTc	(<i>R</i>)- C1	NHPI	O2	---	64	60:40
3	CuTc	(<i>R</i>)- C1	4-Me-NHPI	O2	---	50	71:29
4	CuTc	(<i>R</i>)- C1	4-OMe-NHPI	O2	---	72	75:25
5	CuTc	(<i>R</i>)- C2	4-OMe-NHPI	O2	---	78	69:31
6	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O2	---	72	87:13
7	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O1	---	79	88:12
8	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O3	---	82	86:14
9	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O4	---	60(75)	86:14
10	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O3	ZnO(1.5)	77	87:13
11 ^d	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O3	ZnO(1.5)	65	90:10
12 ^d	CuOAc	(<i>R</i>)- C3	4-OMe-NHPI	O3	ZnO(1.5)	76 ^h	93:7
13 ^{d,e}	CuOAc	(<i>R</i>)- C3	4-OMe-NHPI	O3	ZnO(1.5)	70	93:7
14 ^f	CuOAc	(<i>R</i>)- C3	4-OMe-NHPI	O3	ZnO(1.5)	60(90)	93:7
15 ^f	CuOAc	(<i>R</i>)- C3	4-OMe-NHPI	O3	---	45(70)	83:17
16 ^d	---	(<i>R</i>)- C3	4-OMe-NHPI	O3	ZnO(1.5)	0	N.A.
17 ^d	CuOAc	(<i>R</i>)- C3 ^g	4-OMe-NHPI	O3	---	76 ^h	95:5

^a Reaction was performed on a 0.025 mmol scale. ^b Ratio was determined by crude ¹H NMR using 1,3,5-trimethoxybenzene as internal standard. ^cer value was determined by HPLC. ^d Reaction temperature was 10 °C, 72 h. ^e 30% mol (*R*)-**C3** were added. ^f Reaction temperature was 0 °C. ^g 15% mol of [(*R*)-C₃]₂Zn was used. ^h Isolated yield. Reaction conversion was shown in parentheses. CuTc = Copper thiophene-2-carboxylate. N.A. = not available.

Table S2 Detailed screening of reaction conditions of enantioselective allylic C–H bond amination.

H₈-BINOL-CPAs:

(*R*)-**C1**: Ar = Ph
 (*R*)-**C2**: Ar = 4-OMe-Ph
 (*R*)-**C3**: Ar = 4-CF₃-Ph

Oxidant:

R = Bn, **O1**
 R = cyclopentyl, **O2**

R = Me, **O3**
 R = Et, **O4**

PG =

R-NHPI

entry	[Cu]	CPA	R-NHPI	oxidant	additive	yield(%) ^b	er ^c
1	CuTc	(<i>R</i>)- C1	---	O2 , O1 , LPO, DTBP, PIDA, <i>etc.</i>	---	0	N.A.
2	CuTc	(<i>R</i>)- C1	NHPI	O2	---	64	60:40
3	CuTc	(<i>R</i>)- C1	4-Me-NHPI	O2	---	50	71:29
4	CuTc	(<i>R</i>)- C1	4-OMe-NHPI	O2	---	72	75:25
5	CuTc	(<i>R</i>)- C2	4-OMe-NHPI	O2	---	78	69:31
6	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O2	---	72	87:13
7	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O1	---	79	88:12
8	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O3	---	82	86:14
9	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O4	---	60(75)	86:14
10	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O3	CaO(1.5)	69	84:16
11	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O3	MgO(1.5)	63	90:10
12	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O3	ZnO(1.5)	77	87:13
13 ^d	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O3	MgO(1.5)	65	83:17
14 ^d	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O3	ZnO(1.5)	75	90:10
15 ^d	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O3	ZnO(1.0)	70	91:9
16 ^d	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	O3	ZnO(0.5)	62(85)	92:8
17 ^d	CuOAc	(<i>R</i>)- C3	4-OMe-NHPI(10%)	O3	ZnO(1.5)	40(60)	91:9
18 ^d	CuOAc	(<i>R</i>)- C3	4-OMe-NHPI(30%)	O3	ZnO(1.5)	63(90)	91:9
19 ^d	CuOAc	---	4-OMe-NHPI	O3	ZnO(1.5)	25(50)	N.A.
20 ^d	CuOAc	---	4-OMe-NHPI	O3	---	18(40)	N.A.
21 ^d	CuOAc	(<i>R</i>)- C3	4-OMe-NHPI	O3	ZnO(1.5)	76	93:7
22 ^e	CuOAc	(<i>R</i>)- C3	4-OMe-NHPI	O3	ZnO(1.5)	60(90)	93:7
23 ^d	CuOAc	(<i>R</i>)- C3	4-OMe-NHPI	O3	ZnO(1.5)	70	93:7

		(30%)					
24 ^d	CuOAc	(<i>R</i>)- C3	4-OMe-NHPI	O3	ZnO(1.5)	60(90)	93:7
25 ^d	---	(<i>R</i>)- C3	4-OMe-NHPI	O3	ZnO(1.5)	0	N.A.
26 ^d	CuOAc	(<i>R</i>)- C3 ^f	4-OMe-NHPI	O3	---	76	95:5
		(15%)					
27 ^d	CuOAc	(<i>R</i>)- C3 ^f	4-OMe-NHPI	O3	---	74	95:5
		(20%)					
28 ^d	CuOAc	(<i>R</i>)- C3 ^f	4-OMe-NHPI	O3	---	74	93:7
		(10%)					

^a Reaction was performed on a 0.025 mmol scale. ^b Ratio was determined by crude ¹H NMR using 1,3,5-trimethoxybenzene as internal standard. ^cer value was determined by HPLC. ^d Reaction temperature was 10 °C, 72 h. ^e Reaction temperature was 0 °C. ^f (*R*)-**C3** was pre-treated with 0.5 equiv. of ZnEt₂. ^g Isolated yield. Reaction conversion was shown in parentheses. N.A. = not available.

Table S3 Reaction conditions screening of enantioselective allylic C–H bond amination

H₈-BINOL-CPAs:

(*R*)-**C1**: Ar = Ph
 (*R*)-**C2**: Ar = 4-OMe-Ph
 (*R*)-**C3**: Ar = 4-CF₃-Ph

Oxidant:

R = Bn, **O1**
 R = cyclopentyl, **O2**

R = Me, **O3**
 R = Et, **O4**

PG =

R-NHPI

entry	[Cu]	CPA	R-NHPI	additive	yield(%) ^b	er ^c
1	CuTc	(<i>R</i>)- C3	4-OMe-NHPI	ZnO(1.5)	65	91:9
2	CuCN	(<i>R</i>)- C3	4-OMe-NHPI	ZnO(1.5)	30(50)	80:20
3	CuBr	(<i>R</i>)- C3	4-OMe-NHPI	ZnO(1.5)	78	89:11
4	CuCl	(<i>R</i>)- C3	4-OMe-NHPI	ZnO(1.5)	72	90:10
5	Cu ₂ S	(<i>R</i>)- C3	4-OMe-NHPI	ZnO(1.5)	28(40)	72:28
6	CuOAc	(<i>R</i>)- C3	4-OMe-NHPI	ZnO(1.5)	76	93:7
7 ^c	CuOAc(15%)	(<i>R</i>)- C3 (30%)	4-OMe-NHPI	ZnO(1.5)	68(90)	90:10
8 ^d	CuOAc(5%)	(<i>R</i>)- C3 (10%)	4-OMe-NHPI	ZnO(1.5)	62(80)	92:8
9 ^d	CuOAc(5%)	(<i>R</i>)- C3 (15%)	4-OMe-NHPI	ZnO(1.5)	60(78)	85:15
10 ^d	CuOAc(10%)	(<i>R</i>)- C3 (10%)	4-OMe-NHPI	ZnO(1.5)	60(78)	90:10
11 ^d	CuOAc(10%)	(<i>R</i>)- C3 (15%)	4-OMe-NHPI	ZnO(1.5)	60(80)	87:13
12 ^d	CuOAc(15%)	(<i>R</i>)- C3 (10%)	4-OMe-NHPI	ZnO(1.5)	61(81)	82:18
13 ^d	CuOAc(10%)	(<i>R</i>)- C3 (25%)	4-OMe-NHPI	ZnO(1.5)	60(80)	92:8

^a Reaction was performed on a 0.025 mmol scale. ^b Ratio was determined by crude ¹H NMR using 1,3,5-trimethoxybenzene as internal standard. ^c-5 °C. ^d 0 °C. Reaction conversion was shown in parentheses. N.A. = not available.

Table S4 Optimization of reaction conditions of enantioselective benzylic C–H bond amination^a

<div>CPA:</div> <div> </div> <div> <div>(<i>R</i>)-C1: Ar = Ph</div> <div>(<i>R</i>)-C2: Ar = 4-OMe-Ph</div> <div>(<i>R</i>)-C3: Ar = 4-CF₃-Ph</div> </div> <div> <div>Oxidant:</div> <div> </div> <div>R = Bn, O1</div> <div>R = cyclopentyl, O2</div> <div> </div> <div>R = Me, O3</div> <div>R = Et, O4</div> </div> <div> <div>PG =</div> <div> </div> </div> <div> <div>R-NHPI</div> <div> </div> </div>				
entry	Variation from standard conditions	Conv. (%)	Yield of 2a /(5A / 5B) ^b	ee (%) ^c
1	without 4-OMe-NHPI	<5%	trace(-/-)	N.A.
2	NHPI instead of 4-OMe-NHPI	40	1/2.8	87
3	4-Me-NHPI instead of 4-OMe-NHPI	42	1/2.0	92
4	3-F-NHPI instead of 4-OMe-NHPI	50	1/1.5	70
5	O1 instead of O4	40	30(6/4)	70
6	O2 instead of O4	42	33(4/2)	85
7	O3 instead of O4	43	34(6/4)	81
8	without (<i>R</i>)- C3	20	10(4/5)	N.A.
9	(<i>R</i>)- C1 instead of (<i>R</i>)- C3	30	24(2/3)	84
10	(<i>R</i>)- C2 instead of (<i>R</i>)- C3	34	27(4/-)	81
11	none	35	30(2/-)	94
12	without ZnO	20	15(3/-)	86
13 ^d	(CPA) ₂ Zn instead of (<i>R</i>)- C3 and ZnO	30	22(2/-)	98
14	without CuTc	<5%	trace(-/-)	N.A.
15 ^e	4-OMe-NHPI (1.0 equiv), O4 (3.0 equiv)	55	40(-/-)(67 ^f)	90
16 ^{d,g}	4-OMe-NHPI (1.0 equiv), O4 (3.0 equiv)	50	38(3/4)(76 ^f)	88

^a Reaction was performed on a 0.025 mmol scale. ^b Ratio was determined by crude ¹H NMR using 1,3,5-trimethoxybenzene as internal standard. ^c ee value on HPLC. ^d Zinc phosphate (20 mol%) was used. ^e (*R*)-**C3** (5 mol%) was added at 12 h and 36 h respectively and NaSbF₆ (1.0 equiv) was added.

Reaction was performed on a 0.1 mmol scale and reaction time was 7 days. ^fIsolated yield in parentheses was based on recovered starting material. ^g (*R*)-**C3** (5 mol%) was added at 18 h and 40 h respectively. Reaction was performed on a 0.1 mmol scale and reaction time was 7 days. N.A. = not available.

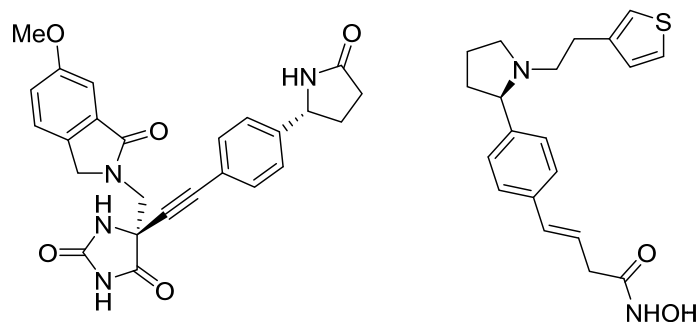
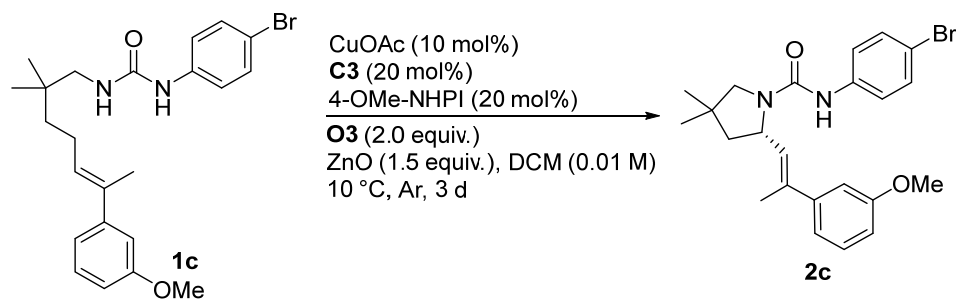


Figure S1 Bioactive molecules containing chiral α -aryl pyrrolidines with olefin or alkyne functional groups at para position



ee of C3 (%)	1 st run ee 2c (%)	2 nd run ee 2c (%)	3 rd ee 2c (%)	average (%)
0	-3.1	-1.0	4.3	0.07
20(<i>R</i>)	25.9	26.6	27.0	26.5
40(<i>R</i>)	55.3	51.9	53.0	53.4
60(<i>R</i>)	66.5	66.4	63.0	65.3
80(<i>R</i>)	77.0	75.7	76.0	76.2
99(<i>R</i>)	86.0	86.0	86.0	86.0

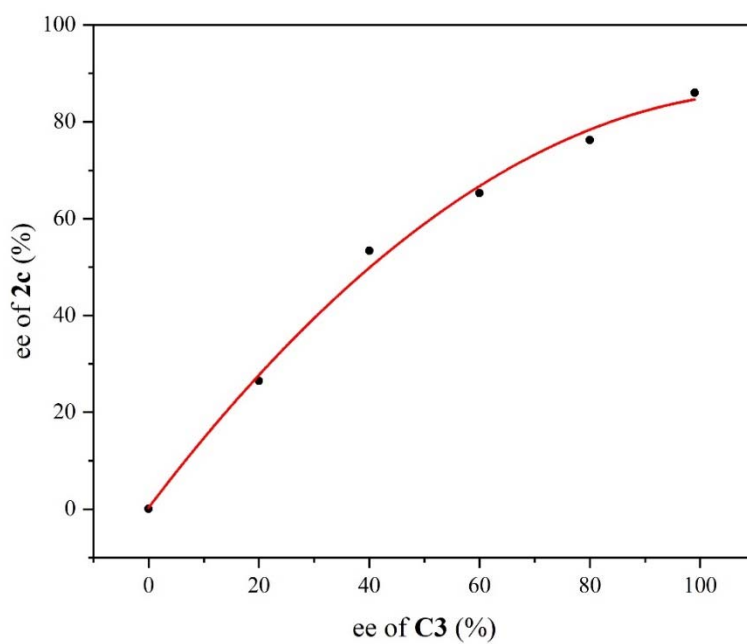
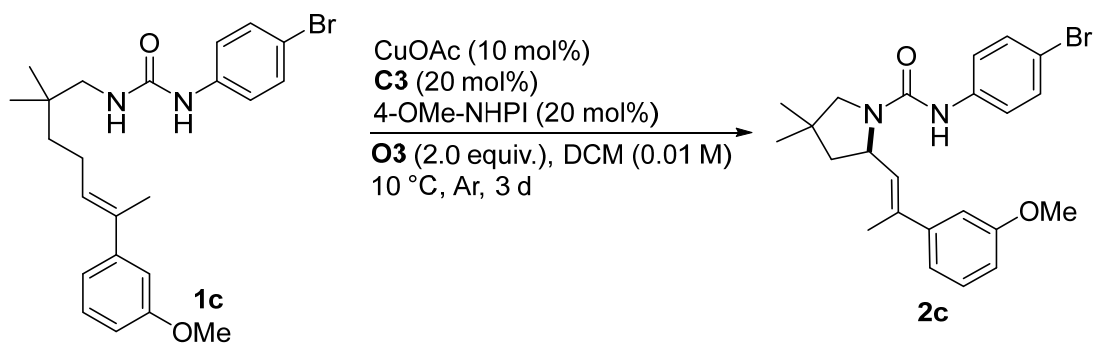


Figure S2 Non-linear effect experiments in the presence of ZnO



ee of C3 (%)	1 st run ee 2c (%)	2 nd run ee 2c (%)	average (%)
0	6	8	7
20(<i>R</i>)	17	20	18.5
40(<i>R</i>)	33	30	31.5
60(<i>R</i>)	37	39	38
80(<i>R</i>)	55	53	54
99(<i>R</i>)	66	68	67

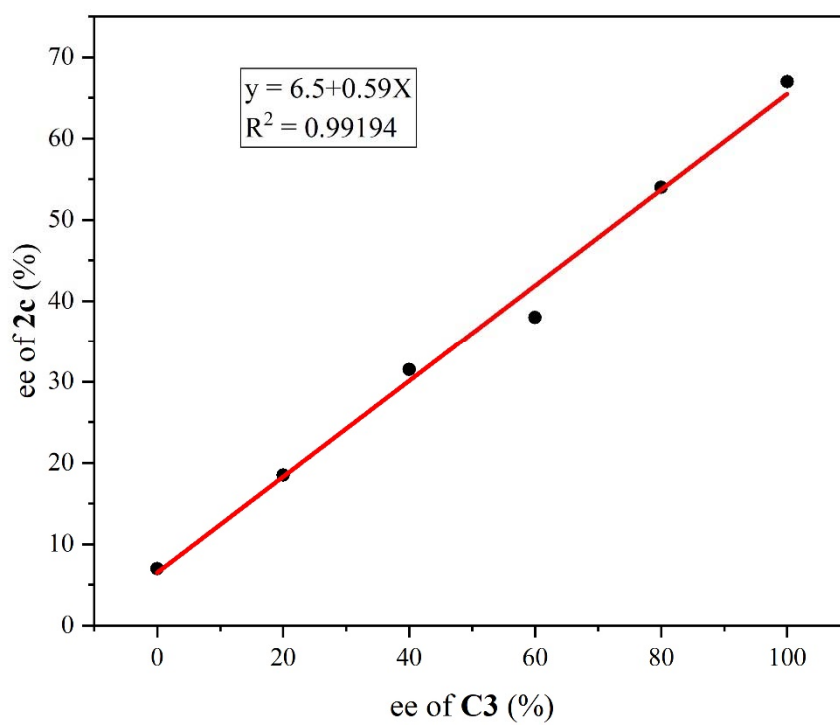
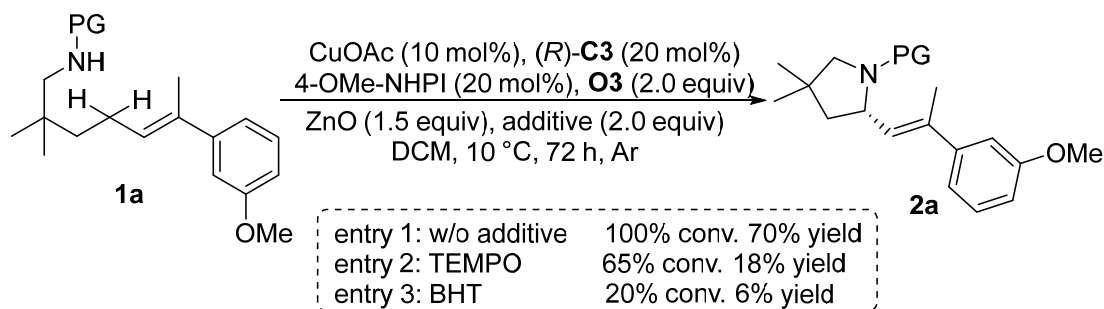
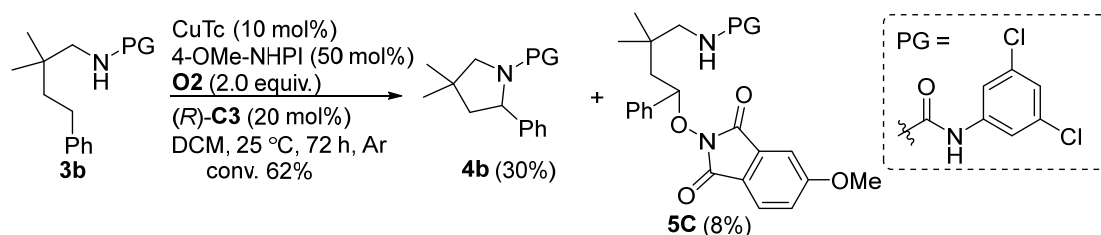


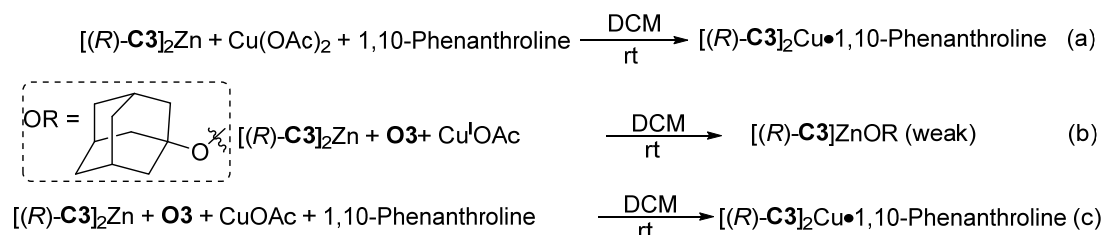
Figure S3 Linear effect experiments without ZnO



Scheme S1 Control experiments with radical inhibitors

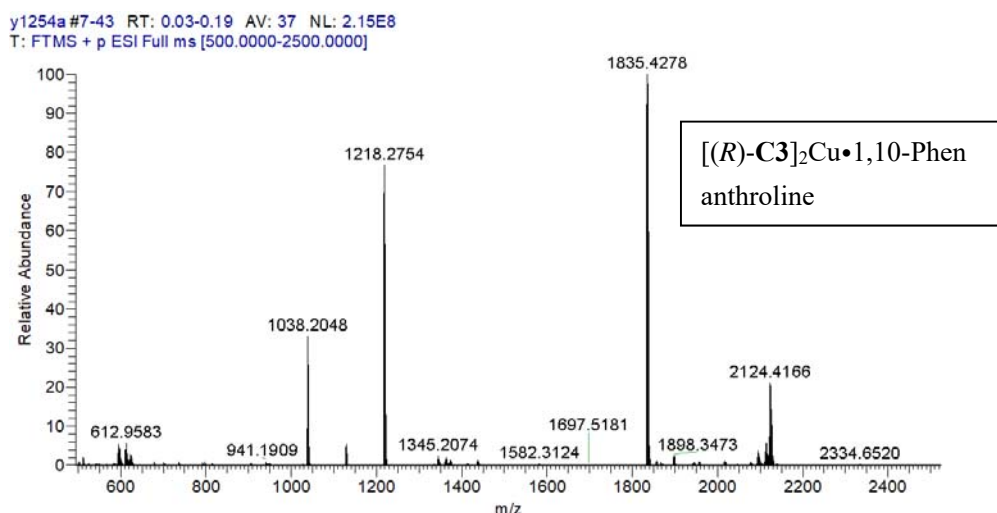


Scheme S2 Benzylic C–H amination of 3b

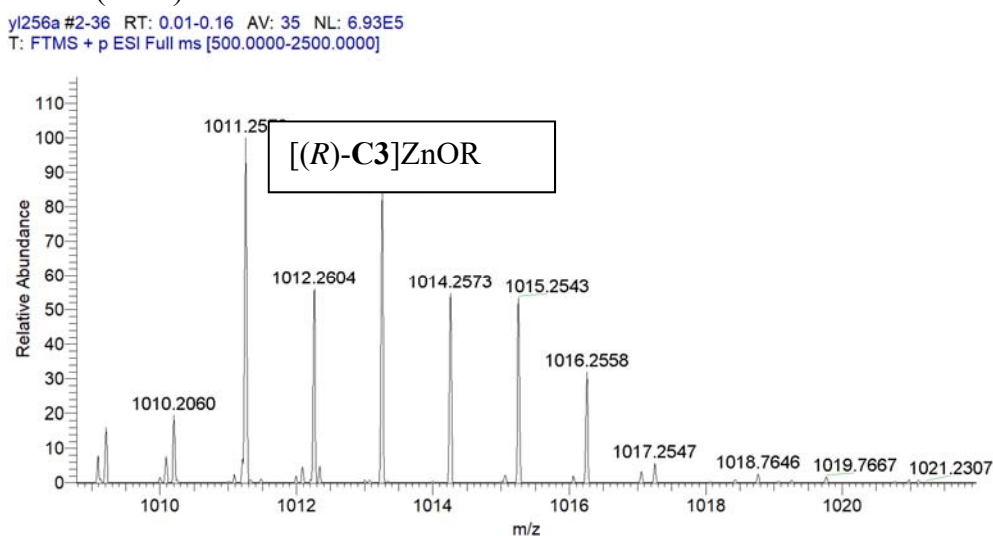


Scheme S3 Transmetalation between Cu(II) and (CPA)₂Zn

Scheme S3-a: To a flame-dried Schlenk tube equipped with a magnetic stir bar were added $[(R)\text{-C3}]_2\text{Zn}$ (6.23 mg, 1.5 equiv.), Cu(OAc)_2 (0.46 mg 1.0 equiv.) and 1,10-Phenanthroline (0.54 mg, 1.2 equiv.). The tube was evacuated and backfilled with argon for three times, then dry DCM (0.5 mL) was added *via* syringe. The tube was stirred at rt overnight. The reaction mixture was evaporated and the sample was analyzed by ESI-MS. The following peaks were observed: **HRMS** (ESI) m/z calcd. for $\text{C}_{104}\text{H}_{77}\text{F}_{12}\text{N}_2\text{O}_8\text{P}_2\text{Cu}$ $[\text{M}+\text{H}]^+$ 1834.4254, found 1834.4246.

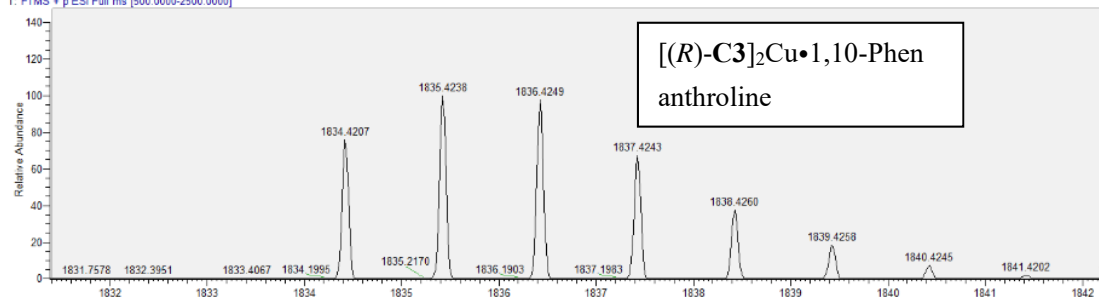


Scheme S3-b: To a flame-dried Schlenk tube equipped with a magnetic stir bar were added [(R)-C3]₂Zn (8.3 mg, 1.5 equiv.), CuOAc (0.62 mg 1.5 equiv.) and **O3** (0.9 mg, 1.0 equiv.). The tube was evacuated and backfilled with argon for three times, then dry DCM (0.5 mL) was added *via* syringe. The tube was stirred at rt for 3 h. The reaction mixture was evaporated and the sample was analyzed by ESI-MS. The following peaks were observed: **HRMS** (ESI) m/z calcd. for C₅₆H₅₀F₆O₅PZn [M+H]⁺ 1011.2586, found 1011.2570 (weak).



Scheme S3-c: To a flame-dried Schlenk tube equipped with a magnetic stir bar were added [(R)-C3]₂Zn (8.3 mg, 1.5 equiv.), CuOAc (0.62 mg 1.5 equiv.), **O3** (0.9 mg, 1.0 equiv.) and 1,10-Phenanthroline (1.5 mg, 2.5 equiv.). The tube was evacuated and backfilled with argon for three times, then dry DCM (0.5 mL) was added *via* syringe. The tube was stirred at rt for 3 h. The reaction mixture was evaporated and the sample was analyzed by ESI-MS. The following peaks were observed: **HRMS** (ESI) m/z calcd. for C₁₀₄H₇₇F₁₂N₂O₈P₂Cu [M+H]⁺ 1834.4254, found 1834.4214.

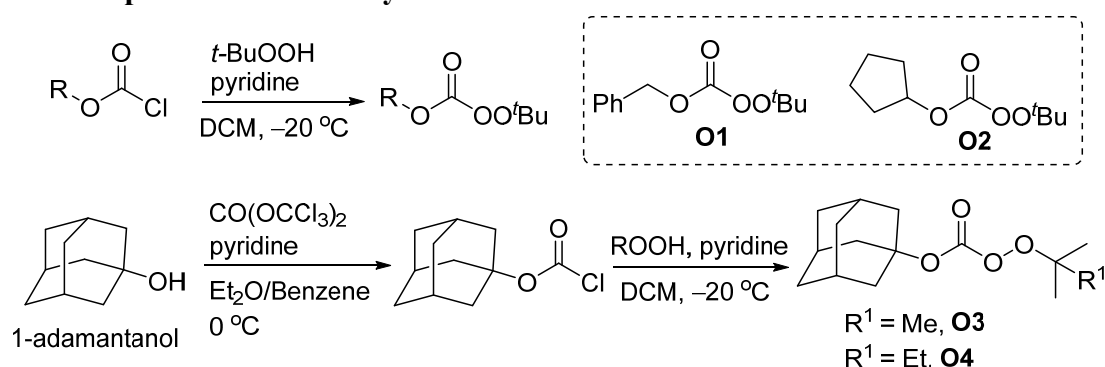
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T: FTMS • p ESI Full ms [500.0000-2500.0000]



General information

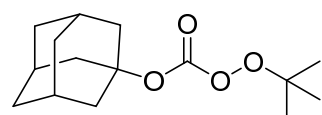
All reactions were carried out under argon (Ar) atmosphere using Schlenk techniques with magnetic stirring. Reagents were purchased at the highest commercial quality and used without further purification, unless otherwise stated. CuTc and CuOAc were purchased from TCL. Chiral phosphoric acids (CPAs) were purchased from Daicel Chiral Technologies (China). Dichloromethane was purchased anhydrous from JK and transferred under an argon atmosphere. Analytical thin layer chromatography (TLC) was performed on precoated silica gel 60 F254 plates. Flash column chromatography was performed using Tsingdao silica gel (60, particle size 0.040-0.063 mm). Visualization on TLC was achieved by use of UV light (254 nm) or iodine. NMR spectra were recorded on a Bruker DPX 400 spectrometer at 400 MHz for ^1H NMR, 100 MHz for ^{13}C NMR and 376 MHz for ^{19}F NMR in CDCl_3 with tetramethylsilane (TMS) as internal standard. Chemical shifts are reported in ppm and coupling constants are given in Hz. Data for ^1H NMR are recorded as follows: chemical shift (ppm), multiplicity (s, singlet; d, doublet; t, triplet; q, quarter; m, multiplet), coupling constant (Hz), integration. Data for ^{13}C NMR are reported in terms of chemical shift (δ , ppm). ^{19}F NMR spectra were recorded on a Bruker DPX 400 MHz spectrometer [CFCl_3 as an external reference (0 ppm)]. HMRS were obtained on a Bruker Apex IV RTMS. The blue LEDs were directly got from the supermarket.

General procedure for the synthesis of oxidant:

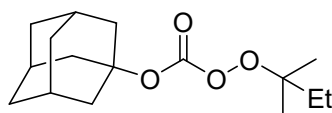


O1 and **O2** were prepared directly from the corresponding benzyl carbonochloridate and cyclopentyl carbonochloridate in 80% and 85% yields, respectively according to reference¹ and the data are consistent with those reported in the literature.

O3 and **O4** were prepared from 1-adamantanol via two steps according to reference².

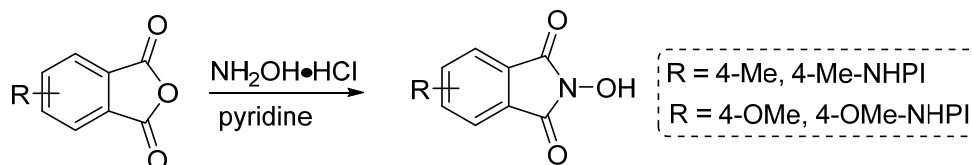


O3, 70% yield from 1-adamantanol, ^1H NMR (400 MHz, **Chloroform-*d***) δ 2.25 – 2.17 (m, 3H), 2.14 (d, J = 3.2 Hz, 6H), 1.67 (t, J = 3.1 Hz, 6H), 1.32 (s, 9H). ^{13}C NMR (101 MHz, **CDCl₃**) δ 152.4, 84.3, 83.5, 41.0, 35.9, 31.0, 25.9. **HRMS** (ESI) m/z calcd. for $\text{C}_{15}\text{H}_{24}\text{NaO}_4$ $[\text{M}+\text{H}]^+$ 291.1563, found 291.1567.



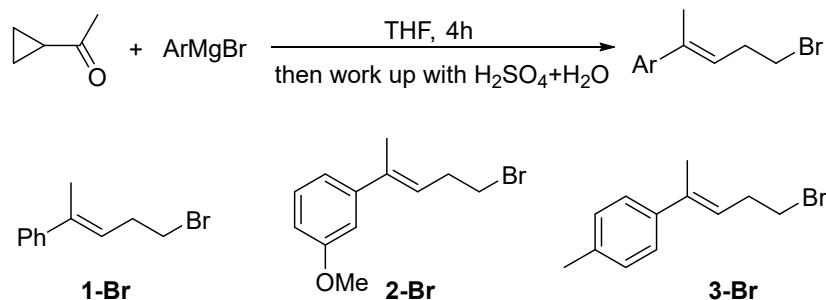
O4, 60% yield from 1-adamantanol, ^1H NMR (400 MHz, **Chloroform-*d***) δ 2.21 (s, 3H), 2.16 – 2.09 (m, 6H), 1.70 – 1.58 (m, 8H), 1.26 (s, 6H), 0.96 – 0.89 (m, 3H). ^{13}C NMR (101 MHz, **CDCl₃**) δ 152.4, 85.9, 84.3, 47.7, 41.0, 35.9, 31.1, 23.5, 8.2. **HRMS** (ESI) m/z calcd. for $\text{C}_{16}\text{H}_{26}\text{NaO}_4$ $[\text{M}+\text{H}]^+$ 305.1723, found 305.1721.

General procedure for the synthesis of R-NHPI.



4-Me-NHPI and 4-OMe-NHPI were prepared directly from the corresponding 5-methylisobenzofuran-1,3-dione and 5-methoxyisobenzofuran-1,3-dione in 85% and 82% yields, respectively according to reference³. Data are consistent with those reported in the literature.

General procedure for the synthesis of homoallylic bromide.



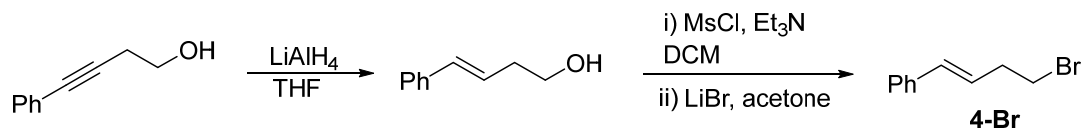
Tri-substituted homoallylic bromides (**1-Br** to **3-Br**) were prepared according to reference⁴.

A flame-dried 50-mL Schlenk flask was evacuated and flushed three times with nitrogen, and then charged with (3-methoxyphenyl)magnesium bromide (60 mmol, 1.0 M in THF, generated in situ from 1-bromo-3-methoxybenzene and magnesium). The solution was cooled to 0 °C, after which a solution of 1-cyclopropylethanone (4.26 mg, 50 mmol, 1 equiv) was added. The resulting solution was warmed up to room temperature and stirred for 4 h. The mixture was cooled to 0 °C, and then a mixture of H_2SO_4 and H_2O ($\text{H}_2\text{SO}_4:\text{H}_2\text{O} = 1:1$, v/v, 16 mL) was added. The resulting mixture was stirred for 30 min at room temperature and extracted with EtOAc (50 mL x 3). The combined organic layers were washed with brine, dried over Na_2SO_4 , and concentrated

in vacuo. The residue was purified by flash column chromatography on silica gel (eluent: PE) to afford the final product **2-Br** (10.4 g, 68% yield).

1-Br and **3-Br** were prepared according to the similar procedure in 62% and 71% yields, respectively.

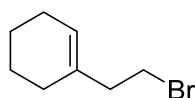
Di-substituted homoallylic bromides **4-Br** was prepared according to procedure reported in the reference⁵.



To a three necked flask, THF (60 mL) and LiAlH_4 (3.8 g, 100 mmol) were added. This solution was cooled at 0 °C, and 4-phenylbut-3-yn-1-ol (2.92 g, 20 mmol) in THF (10 mL) was dropped to the mixture over 15 min. After refluxing for 18 h, the reaction mixture was quenched with 1N NaOH (10 mL) at 0 °C. This mixture was filtered, and the filtrate was dried (MgSO_4). The solvent was evaporated, and the residue was purified by column chromatography (eluent: hexane/ethyl acetate = 80:20) to give the alcohol product (2.84 g, 96%).

To an ice cooled solution of above alcohol (9.1 mmol) and Et_3N (1.9 mL, 13.7 mmol) in dry DCM (20 mL) was added methanesulfonyl chloride (0.8 mL, 10 mmol) *via* syringe at 0 °C under argon atmosphere. The mixture was stirred for 45 min at 0 °C before the reaction was quenched with H_2O . The organic materials were washed with saturated NaHCO_3 aqueous solution, then extracted twice with DCM. The combined organic extracts were washed with brine, dried over Na_2SO_4 , and concentrated *in vacuo* after filtration.

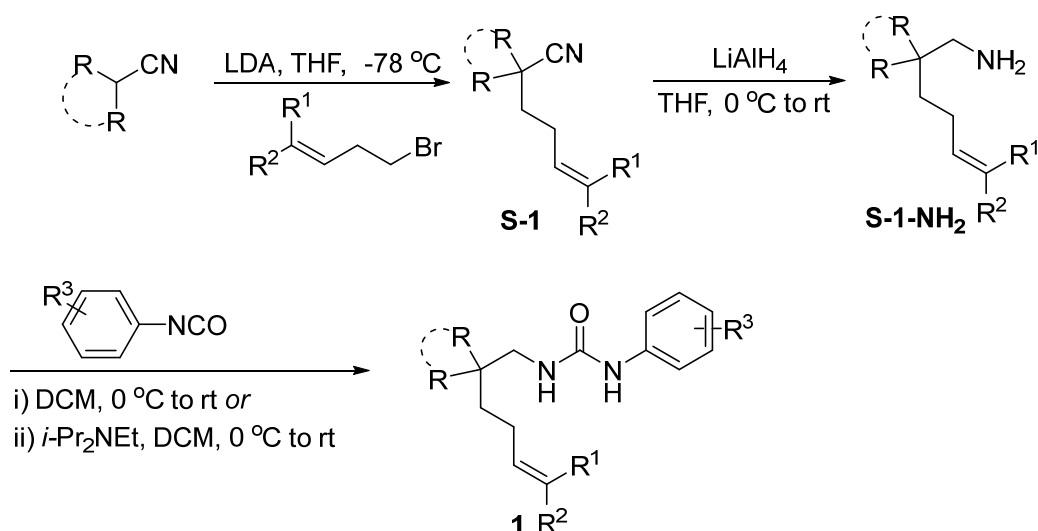
The resulting crude material was dissolved in acetone (40 mL) before addition of lithium bromide (3.6 g, 41 mmol) at 0 °C. The mixture was stirred at 50 °C for 5 h. The reaction was then cooled to room temperature and acetone was removed under reduced pressure. The resulting residue was diluted with H_2O and the organic materials were extracted thrice with ethyl acetate. The combined organic extracts were dried over Na_2SO_4 and concentrated *in vacuo* after filtration. The resulting residue was purified by flash column chromatography (petroleum ether/ ethyl acetate = 100/1-30/1) to give the **4-Br** (1.63 g, 7.7 mmol) in 80% overall isolated yield in 2 steps.



5-Br

5-Br was prepared according to reference and the data are consistent with those reported in the literature⁶.

General procedure for the synthesis of alkenyl urea substrates 1.



To a two-neck round bottom flask containing diisopropylamine (2.0 mL, 13.3 mmol) in dry THF (15 mL) cooled to $-78\text{ }^{\circ}\text{C}$ was added dropwise *n*-BuLi (5.6 mL, 2.4 M, 13.3 mmol). The solution was stirred at $-78\text{ }^{\circ}\text{C}$ for 30 min. A solution of isobutyronitrile (1.0 mL, 11.1 mmol) was added dropwise and the solution was stirred at $-78\text{ }^{\circ}\text{C}$ for further 30 min. To the reaction mixture was added dropwise a solution of homoallylic bromide (12.2 mmol) in dry THF (5.0 mL). Upon completion of the addition, the reaction was allowed to warm to room temperature and stirred overnight. The reaction was quenched by adding saturated aqueous NH_4Cl (20 mL), and the mixture was extracted three times with Et_2O . The combined organic extracts were washed with water and brine. After drying with MgSO_4 and removal of the solvents under reduced pressure, the resulting crude material was purified by flash column chromatography to yield the corresponding products **S-1**⁷.

The following reduction of **S-1** by LiAlH_4 and reaction with aryl isocyanate were performed according to procedures reported in the reference⁸.

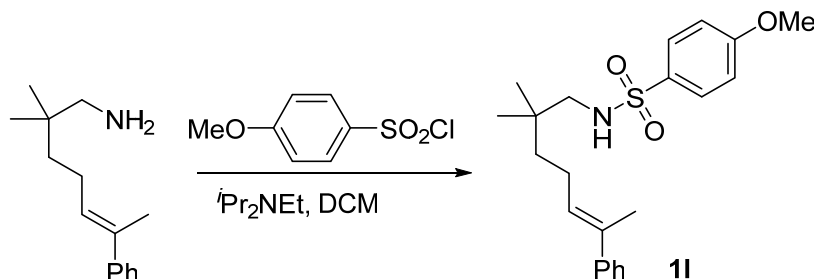
Reduction of S-1: To a suspension solution of LiAlH_4 (10.0 mmol) in anhydrous THF (20.0 mL) was slowly added **S-1** (5.0 mmol) in dry THF (5.0 mL) in ice-bath under Argon. Then the mixture was warmed to room temperature, and stirred for an additional 2 hours. The reaction mixture was quenched by slow, sequential addition of water (0.5 mL) in Na_2SO_4 (4.0 g) at $0\text{ }^{\circ}\text{C}$. The reaction mixture was warmed to room temperature, stirred for an additional 30 minutes, filtered and concentrated *in vacuo* to afford corresponding products **S-1-NH₂**, which was directly used in the next reaction without further purification.

Synthesis of urea substrate 1a. To a stirred solution of **S-1a-NH₂** (2.0 mmol) in dry DCM (5.0 mL) was slowly added 1-isocyanato-3,5-bis(trifluoromethyl)-benzene (2.0 mmol) at $0\text{ }^{\circ}\text{C}$. Then, the reaction mixture was stirred for an additional 15 min at $0\text{ }^{\circ}\text{C}$. After complete conversion (monitored by TLC), the crude mixture was directly purified by silica gel column chromatography and dried under reduced pressure to afford **1a** (0.60 g, 60%) as a white powder.

Synthesis of urea substrates 1b. 1,2-dichloro-4-isocyanatobenzene (2.0 mmol) was slowly added to a stirred solution of **S-2a-NH₂** (2.0 mmol) (2.0 mmol) and ethyldiisopropylamine (*i*- Pr_2NEt , 2.0 mmol) in dry DCM (5.0 mL) at $0\text{ }^{\circ}\text{C}$. The reaction

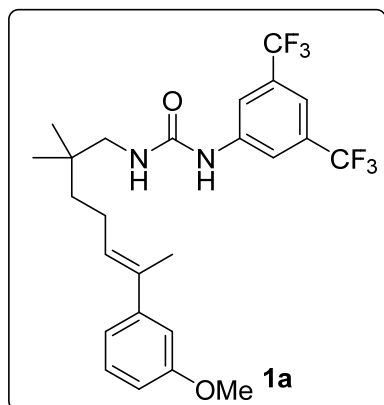
mixture was stirred for an additional 15 min at 0 °C. After complete conversion (monitored by TLC), the crude mixture was directly purified by silica gel column chromatography to give urea substrate **1b** (0.48 g, 55%).

Procedure for the synthesis of substrate **1l**.



A flame-dried 25-mL round-bottom flask was charged with (*E*)-2,2-dimethyl-6-phenylhept-5-en-1-amine (392 mg, 1.8 mmol) and *i*Pr₂NEt (0.62 mL, 3.6 mmol) in DCM (8.0 mL), 4-methoxybenzenesulfonyl chloride (447 mg, 2.16 mmol) was added. The resulting mixture was stirred at rt overnight, and the reaction mixture was quenched with saturated NaHCO₃ aqueous solution, then extracted twice with DCM. The combined organic extracts was washed with brine, dried over Na₂SO₄, and concentrated in vacuo after filtration. The crude mixture was directly purified by silica gel column chromatography to give substrate **1l** (419 mg, 60% yield).

(*E*)-1-(3,5-bis(trifluoromethyl)phenyl)-3-(6-(3-methoxyphenyl)-2,2-dimethylhept-5-en-1-yl)urea



1a, 60% yield, ¹H NMR (400 MHz, Chloroform-*d*) δ 7.78 (s, 1H), 7.73 (d, *J* = 1.5 Hz, 2H), 7.41 (t, *J* = 1.4 Hz, 1H), 7.20 (t, *J* = 7.9 Hz, 1H), 6.92 (ddd, *J* = 7.7, 1.8, 0.9 Hz, 1H), 6.87 (dd, *J* = 2.6, 1.7 Hz, 1H), 6.76 (ddd, *J* = 8.2, 2.6, 0.9 Hz, 1H), 5.68 (tq, *J* = 7.2, 1.4 Hz, 1H), 5.58 (t, *J* = 6.1 Hz, 1H), 3.79 (s, 3H), 3.12 (d, *J* = 6.0 Hz, 2H), 2.11 (dd, *J* = 10.7, 6.1 Hz, 2H), 1.96 (d, *J* = 1.3 Hz, 3H), 1.34 – 1.27 (m, 2H), 0.90 (s, 6H).

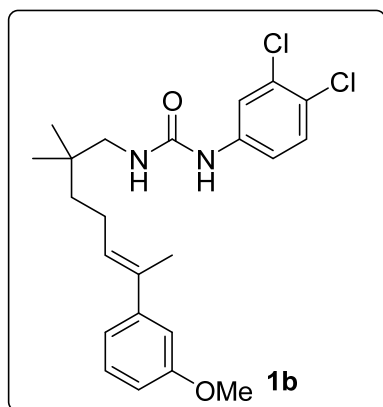
¹³C NMR (101 MHz, CDCl₃) δ 159.4, 155.7, 145.3, 140.5, 134.6, 132.2 (q, *J*_{C-F} = 33.1 Hz), 129.2, 128.4, 123.1 (q, *J*_{C-F} = 271.0 Hz), 118.5, 118.2, 115.9 – 115.7

(m, 1C), 111.7, 111.6, 55.2, 50.1, 39.4, 34.4, 24.7, 23.3, 15.7.

¹⁹F NMR (376 MHz, CDCl₃) δ -63.27.

HRMS (ESI) *m/z* calcd. for C₂₅H₂₉F₆N₂O₂ [M+H]⁺ 503.2133, found 503.2137.

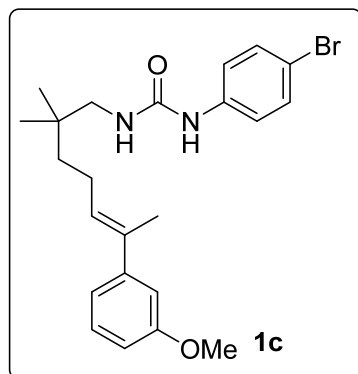
(*E*)-1-(3,4-dichlorophenyl)-3-(6-(3-methoxyphenyl)-2,2-dimethylhept-5-en-1-yl)urea



1b, 55% yield, ^1H NMR (400 MHz, Chloroform-*d*) δ 8.19 (s, 1H), 7.43 (d, $J = 2.4$ Hz, 1H), 7.18 (t, $J = 8.0$ Hz, 1H), 7.13 (d, $J = 8.8$ Hz, 1H), 6.94 (dd, $J = 8.8, 2.4$ Hz, 1H), 6.91 (dt, $J = 8.0, 1.2$ Hz, 1H), 6.86 (t, $J = 2.0$ Hz, 1H), 6.74 (ddd, $J = 8.2, 2.4, 0.8$ Hz, 1H), 6.05 (t, $J = 6.0$ Hz, 1H), 5.66 (td, $J = 7.2, 1.6$ Hz, 1H), 3.77 (s, 3H), 3.06 (d, $J = 6.0$ Hz, 2H), 2.11 – 2.05 (m, 2H), 1.95 (d, $J = 1.2$ Hz, 3H), 1.31 – 1.26 (m, 2H), 0.86 (s, 6H). ^{13}C NMR (101 MHz, CDCl_3) δ 159.5, 156.7, 145.3, 138.6, 134.6, 132.6, 130.3, 129.2, 128.5, 125.9, 121.2, 118.9, 118.2, 111.7, 111.7, 55.2, 50.1, 39.5, 34.5, 24.8, 23.4, 15.8.

HRMS (ESI) m/z calcd. for $\text{C}_{23}\text{H}_{29}\text{Cl}_2\text{N}_2\text{O}_2$ $[\text{M}+\text{H}]^+$ 435.1601, found 435.1599.

(E)-1-(4-bromophenyl)-3-(6-(3-methoxyphenyl)-2,2-dimethylhept-5-en-1-yl)urea

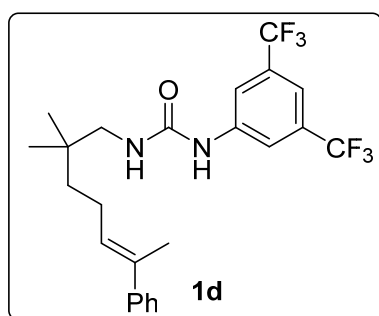


1c, 65% yield, ^1H NMR (400 MHz, Chloroform-*d*) δ 7.85 (s, 1H), 7.27 – 7.22 (m, 2H), 7.19 (t, $J = 8.0$ Hz, 1H), 7.11 – 7.09 (m, 2H), 6.91 (ddd, $J = 7.8, 1.8, 1.0$ Hz, 1H), 6.87 (dd, $J = 2.8, 1.6$ Hz, 1H), 6.75 (ddd, $J = 8.0, 2.4, 0.8$ Hz, 1H), 5.82 (t, $J = 6.0$ Hz, 1H), 5.69 – 5.65 (m, 1H), 3.78 (s, 3H), 3.04 (d, $J = 6.0$ Hz, 2H), 2.11 – 2.04 (m, 2H), 1.94 (d, $J = 1.6$ Hz, 3H), 1.31 – 1.21 (m, 2H), 0.84 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3) δ 159.5, 156.7, 145.4, 138.2, 134.5, 131.9, 129.2, 128.6, 121.4, 118.2, 115.5, 111.7, 111.6, 55.3, 50.0, 39.5, 34.5, 24.8, 23.4, 15.8.

HRMS (ESI) m/z calcd. for $\text{C}_{23}\text{H}_{30}\text{BrN}_2\text{O}_2$ $[\text{M}+\text{H}]^+$ 445.1485, found 445.1483.

(E)-1-(3,5-bis(trifluoromethyl)phenyl)-3-(2,2-dimethyl-6-phenylhept-5-en-1-yl)urea



1d, 62% yield, ^1H NMR (400 MHz, Chloroform-*d*) δ 8.26 (s, 1H), 7.73 (s, 2H), 7.41 (s, 1H), 7.36 – 7.26 (m, 4H), 7.25 – 7.19 (m, 1H), 5.95 (t, $J = 6.0$ Hz, 1H), 5.71 – 5.67 (m, 1H), 3.15 (d, $J = 6.0$ Hz, 2H), 2.22 – 2.06 (m, 2H), 2.00 (d, $J = 1.3$ Hz, 3H), 1.42 – 1.29 (m, 2H), 0.92 (s, 6H).

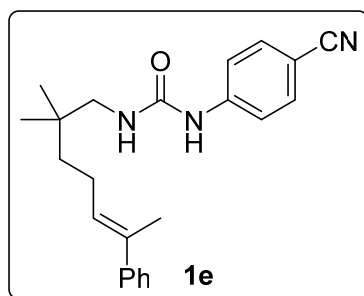
^{13}C NMR (101 MHz, CDCl_3) δ 156.2, 143.7, 140.4, 134.8, 132.2 (q, $J_{\text{C-F}} = 33.1$ Hz), 128.2, 128.0, 126.6, 125.5, 123.1 (q, $J_{\text{C-F}} = 271.0$ Hz), 118.5, 115.8, 50.2,

39.6, 34.4, 24.6, 23.3, 15.6.

^{19}F NMR (376 MHz, CDCl_3) δ -63.31.

HRMS (ESI) m/z calcd. for $\text{C}_{24}\text{H}_{27}\text{F}_6\text{N}_2\text{O}$ $[\text{M}+\text{H}]^+$ 473.2028, found 473.2030.

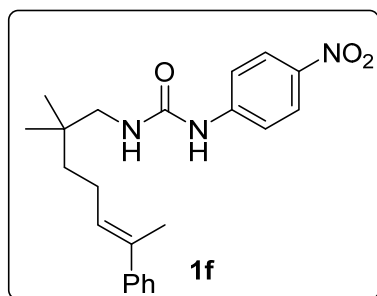
(E)-1-(4-cyanophenyl)-3-(2,2-dimethyl-6-phenylhept-5-en-1-yl)urea



1e, 55% yield, ^1H NMR (400 MHz, Chloroform-*d*) δ 8.05 (s, 1H), 7.43 (s, 4H), 7.33 – 7.22 (m, 4H), 7.21 – 7.14 (m, 1H), 5.77 (t, J = 6.0 Hz, 1H), 5.67 – 5.64 (m, 1H), 3.12 (d, J = 6.0 Hz, 2H), 2.13 – 2.11 (m, 2H), 1.97 (d, J = 1.3 Hz, 3H), 1.38 – 1.27 (m, 2H), 0.91 (s, 6H). ^{13}C NMR (101 MHz, CDCl_3) δ 155.7, 144.1, 143.7, 134.8, 133.3, 128.2, 128.1, 126.7, 125.5, 119.7, 118.3, 104.1, 49.9, 39.5, 34.5, 24.9, 23.4, 15.7.

HRMS (ESI) m/z calcd. for $\text{C}_{23}\text{H}_{28}\text{N}_3\text{O}$ $[\text{M}+\text{H}]^+$ 362.2232, found 362.2228.

(E)-1-(2,2-dimethyl-6-phenylhept-5-en-1-yl)-3-(4-nitrophenyl)urea

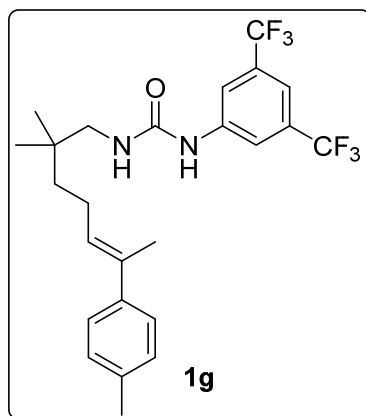


1f, 50% yield, ^1H NMR (400 MHz, Chloroform-*d*) δ 8.11 (d, J = 9.2 Hz, 2H), 8.05 (s, 1H), 7.53 (d, J = 9.2 Hz, 2H), 7.36 – 7.27 (m, 4H), 7.24 – 7.17 (m, 1H), 5.77 (t, J = 6.1 Hz, 1H), 5.73 – 5.68 (m, 1H), 3.19 (d, J = 6.1 Hz, 2H), 2.20 – 2.14 (m, 2H), 2.01 (d, J = 1.2 Hz, 3H), 1.42 – 1.34 (m, 2H), 0.96 (s, 6H). ^{13}C NMR (101 MHz, CDCl_3) δ 155.3, 146.1, 143.6, 141.7, 134.8, 128.2, 128.1, 126.6, 125.5, 125.4, 117.7,

50.1, 39.4, 34.5, 24.9, 23.4, 15.7.

HRMS (ESI) m/z calcd. for $\text{C}_{22}\text{H}_{28}\text{N}_3\text{O}_2$ $[\text{M}+\text{H}]^+$ 382.2131, found 382.2127.

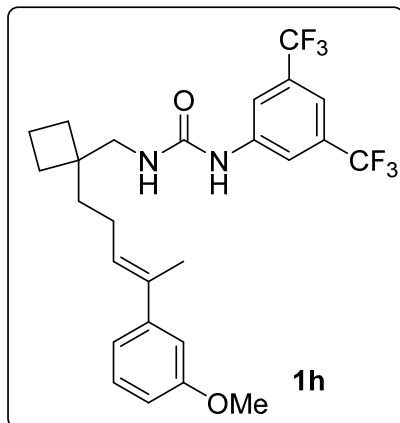
(E)-1-(3,5-bis(trifluoromethyl)phenyl)-3-(2,2-dimethyl-6-(p-tolyl)hept-5-en-1-yl)urea



1g, 58% yield, ^1H NMR (400 MHz, Chloroform-*d*) δ 7.86 (s, 2H), 7.50 (s, 1H), 7.28 – 7.26 (m, 3H), 7.14 – 7.11 (m, 2H), 6.82 (s, 1H), 5.74 – 5.70 (m, 1H), 4.90 (t, J = 6.1 Hz, 1H), 3.18 (d, J = 6.2 Hz, 2H), 2.35 (s, 3H), 2.22 – 2.17 (m, 2H), 2.03 (d, J = 1.2 Hz, 3H), 1.43 – 1.34 (m, 2H), 0.98 (s, 6H). ^{13}C NMR (101 MHz, CDCl_3) δ 155.7, 140.8, 140.5, 136.3, 134.6, 132.2 (q, $J_{\text{C-F}}$ = 33.1 Hz), 128.9, 127.3, 125.3, 123.2 (q, $J_{\text{C-F}}$ = 271.0 Hz), 118.5, 115.8, 50.1, 39.6, 34.4, 24.7, 23.3, 21.0, 15.6. ^{19}F NMR (376 MHz, CDCl_3) δ -63.13.

HRMS (ESI) m/z calcd. for $\text{C}_{25}\text{H}_{29}\text{F}_6\text{N}_2\text{O}$ $[\text{M}+\text{H}]^+$ 487.2184, found 487.2186.

(E)-1-(3,5-bis(trifluoromethyl)phenyl)-3-((1-(4-(3-methoxyphenyl)pent-3-en-1-yl)cyclobutyl)methyl)urea



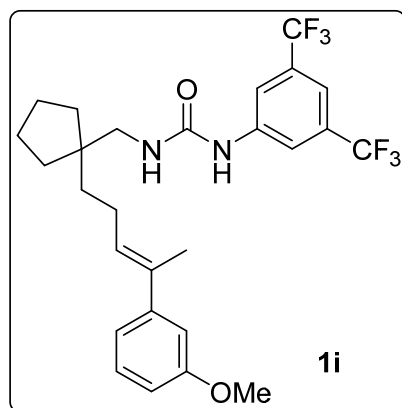
1h, 65% yield, ^1H NMR (400 MHz, Chloroform-*d*) δ 8.00 (s, 1H), 7.74 (s, 2H), 7.42 (s, 1H), 7.22 (t, J = 8.0 Hz, 1H), 6.95 – 6.92 (m, 1H), 6.90 – 6.89 (m, 1H), 6.80 – 6.77 (m, 1H), 5.82 – 5.61 (m, 2H), 3.81 (s, 3H), 3.38 (d, J = 5.6 Hz, 2H), 2.18 – 2.08 (m, 2H), 1.99 (s, 3H), 1.94 – 1.74 (m, 6H), 1.62 – 1.53 (m, 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 159.4, 156.0, 145.3, 140.5, 134.8, 132.1 (q, $J_{\text{C-F}}$ = 33.1 Hz), 129.2, 128.1, 123.1 (q, $J_{\text{C-F}}$ = 271.0 Hz), 118.4, 118.2, 115.7, 111.7, 111.6, 55.2, 46.3, 41.7, 37.2, 29.1, 23.4, 15.7, 15.1.

^{19}F NMR (376 MHz, CDCl_3) δ -63.25.

HRMS (ESI) m/z calcd. for $\text{C}_{26}\text{H}_{29}\text{F}_6\text{N}_2\text{O}_2$ $[\text{M}+\text{H}]^+$ 515.2133, found 515.2134.

(*E*)-1-(3,5-bis(trifluoromethyl)phenyl)-3-((1-(4-(3-methoxyphenyl)pent-3-en-1-yl)cyclopentyl)methyl)urea



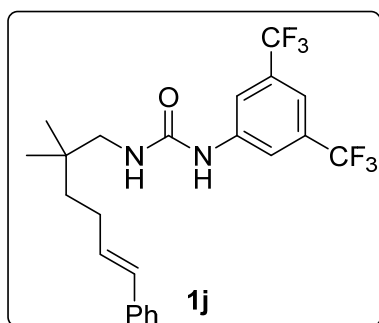
1i, 62% yield, ^1H NMR (400 MHz, Chloroform-*d*) δ 8.41 (s, 1H), 7.66 (s, 2H), 7.35 (s, 1H), 7.17 (t, J = 7.9 Hz, 1H), 6.90 – 6.87 (m, 1H), 6.85 (t, J = 2.1 Hz, 1H), 6.75 – 6.73 (m, 1H), 6.06 (t, J = 5.9 Hz, 1H), 5.68 – 5.64 (m, 1H), 3.77 (s, 3H), 3.22 (d, J = 5.5 Hz, 2H), 2.15 – 2.07 (m, 2H), 1.93 (s, 3H), 1.58 – 1.55 (m, 4H), 1.44 – 1.36 (m, 6H).

^{13}C NMR (101 MHz, CDCl_3) δ 159.4, 156.5, 145.3, 140.4, 134.7, 132.1 (q, $J_{\text{C-F}}$ = 33.1 Hz), 129.1, 128.2, 123.1 (q, $J_{\text{C-F}}$ = 271.0 Hz), 118.5, 118.2, 115.8, 111.7, 111.6, 55.2, 46.7, 45.8, 37.4, 35.5, 25.0, 24.1, 15.6.

^{19}F NMR (376 MHz, CDCl_3) δ -63.32.

HRMS (ESI) m/z calcd. for $\text{C}_{27}\text{H}_{31}\text{F}_6\text{N}_2\text{O}_2$ $[\text{M}+\text{H}]^+$ 529.2290, found 529.2295.

(*E*)-1-(3,5-bis(trifluoromethyl)phenyl)-3-(2,2-dimethyl-6-phenylhex-5-en-1-yl)urea



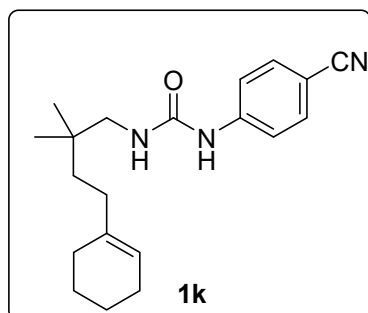
1j, 50% yield, ^1H NMR (400 MHz, Chloroform-*d*) δ 7.83 (s, 2H), 7.46 (s, 1H), 7.33 – 7.26 (m, 4H), 7.21 – 7.19 (m, 1H), 7.13 (s, 1H), 6.42 – 6.33 (dt, J = 16.0, 1.6 Hz, 1H), 6.18 (dt, J = 16.0, 6.7 Hz, 1H), 5.08 (t, J = 6.2 Hz, 1H), 3.15 (d, J = 6.1 Hz, 2H), 2.24 – 2.13 (m, 2H), 1.42 – 1.36 (m, 2H), 0.94 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3) δ 154.9, 140.6, 137.6, 132.2 (q, $J_{\text{C-F}}$ = 33.1 Hz), 130.8, 129.8, 128.6, 127.0, 125.9, 123.2 (q, $J_{\text{C-F}}$ = 271.0 Hz), 118.4, 115.8, 50.0,

39.3, 34.4, 27.6, 24.9. ^{19}F NMR (376 MHz, CDCl_3) δ -63.08.

HRMS (ESI) m/z calcd. for $C_{23}H_{25}F_6N_2O$ $[M+H]^+$ 459.1866, found 459.1863.

1-(4-cyanophenyl)-3-(4-(cyclohex-1-en-1-yl)-2,2-dimethylbutyl)urea

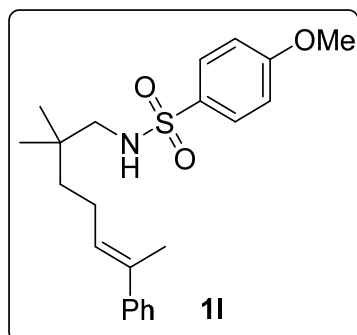


1k, 45% yield, 1H NMR (400 MHz, Chloroform-*d*) δ 7.75 (s, 1H), 7.58 – 7.37 (m, 4H), 5.51 (t, J = 6.0 Hz, 1H), 5.36 – 5.33 (m, 1H), 3.09 (d, J = 6.0 Hz, 2H), 1.99 – 1.82 (m, 6H), 1.64 – 1.43 (m, 4H), 1.36 – 1.23 (m, 2H), 0.88 (s, 6H).

^{13}C NMR (101 MHz, $CDCl_3$) δ 155.4, 143.9, 137.9, 133.3, 120.6, 119.5, 118.4, 104.4, 49.9, 37.9, 34.2, 32.2, 28.6, 25.2, 24.9, 23.1, 22.5.

HRMS (ESI) m/z calcd. for $C_{20}H_{28}N_3O$ $[M+H]^+$ 326.2232, found 326.2230.

(*E*)-N-(2,2-dimethyl-6-phenylhept-5-en-1-yl)-4-methoxybenzenesulfonamide

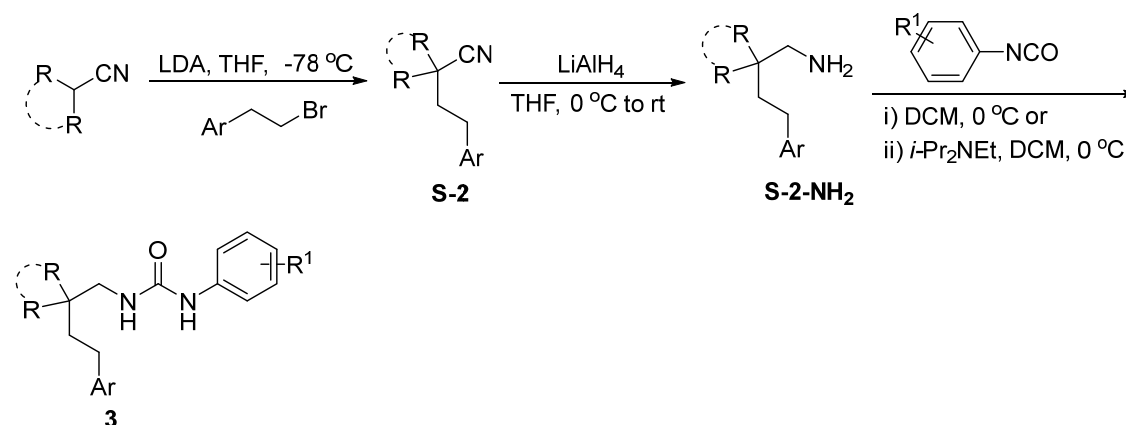


1l, 75% yield, 1H NMR (400 MHz, Chloroform-*d*) δ 7.81 (d, J = 8.9 Hz, 2H), 7.36 – 7.25 (m, 5H), 7.22 – 7.16 (m, 1H), 6.93 (d, J = 8.9 Hz, 2H), 5.66 (td, J = 7.1, 1.4 Hz, 1H), 5.13 – 5.06 (m, 1H), 3.79 (s, 3H), 2.70 (d, J = 6.8 Hz, 2H), 2.12 – 2.02 (m, 2H), 1.98 (d, J = 1.3 Hz, 3H), 1.36 – 1.27 (m, 2H), 0.89 (s, 6H).

^{13}C NMR (101 MHz, $CDCl_3$) δ 162.8, 143.8, 134.6, 131.6, 129.2, 128.4, 128.2, 126.5, 125.6, 114.3, 55.6, 52.9, 39.1, 33.9, 25.0, 23.3, 15.8.

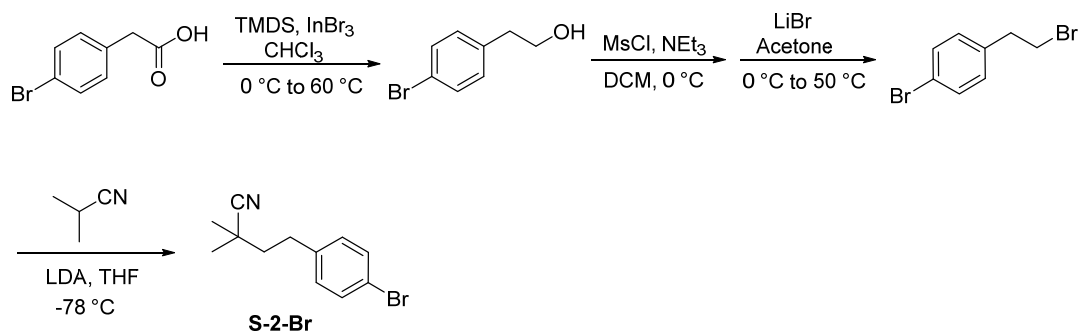
HRMS (ESI) m/z calcd. for $C_{22}H_{30}NO_3S$ $[M+H]^+$ 388.1946, found 388.1944.

General procedure for the synthesis of benzyl urea substrates 3.



S-2 were synthesized from homobenzylic bromides according to the similar procedures reported in the reference^{7,8}. Benzyl urea substrates **3** were prepared from **S-2** through $LiAlH_4$ reduction and reaction with different aryl isocyanates with the similar procedures for the synthesis of substrates **1**.

General procedure for the synthesis of homobenzylic bromide S-2-Br.⁹



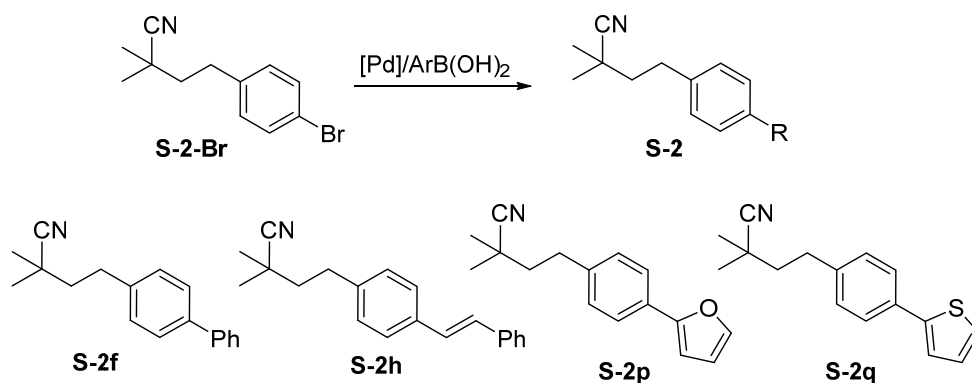
To a two-neck round bottom flask equipped with a magnetic stir bar was added 2-(4-bromophenyl)acetic acid (10.8 g, 50 mmol) and InBr_3 (886.7 mg, 5 mmol%), CHCl_3 (50 mL) was successively added under argon atmosphere. Then 1,1,3,3-Tetramethyldisiloxane (TMDS) (17.7 mL, 100 mmol) was dropwise added at 0 °C under argon atmosphere for 5 min. The mixture was stirred at 60 °C ca. 2h. Then, the reaction was quenched with H_2O at 0 °C and the organic layer was dried with anhydrous Na_2SO_4 and evaporated under reduced pressure. The crude product was purified by flash column chromatography (petroleum ether/ ethyl acetate =10/1-2/1) to afford the 2-(4-bromophenyl)ethan-1-ol (5.5 g, 27.5 mmol) in 55% isolated yields.

To an ice cooled solution of above alcohol and Et_3N (5.7 mL, 41.3 mmol) in dry DCM (80 mL) was added methanesulfonyl chloride (2.3 mL, 41.3 mmol) *via* syringe at 0 °C under argon atmosphere. The mixture was stirred for 45 min at 0 °C before the reaction was quenched with H_2O . The organic materials were washed with saturated NaHCO_3 aqueous solution, then extracted twice with DCM. The combined organic extracts were washed with brine, dried over Na_2SO_4 , and concentrated in vacuo after filtration.

The resulting crude material was dissolved in acetone (80 mL) before addition of lithium bromide (10.8 g, 123.8 mmol) at 0 °C. The mixture was stirred at 50 °C overnight. The reaction was then cooled to room temperature and acetone was removed under reduced pressure. The resulting residue was diluted with H_2O and the organic materials were extracted thrice with ethyl acetate. The combined organic extracts were dried over Na_2SO_4 and concentrated in vacuo after filtration. The resulting residue was purified by flash column chromatography (petroleum ether/ ethyl acetate =100/1-30/1) to give the 1-bromo-4-(2-bromoethyl)benzene (7.0 g, 26.7 mmol) in 97% overall isolated yield in 2 steps.

S-2-Br (3.52g, 70% yield) was synthesized from isobutyronitrile (20 mmol) and 1-bromo-4-(2-bromoethyl)benzene (21 mmol) under LDA conditions with the similar procedure mentioned above.

General procedure for the synthesis of S-2f, S-2h, S-2p and S-2q.



S-2f, **S-2h**, **S-2p** and **S-2q** were prepared from **S-2-Br** with different ArB(OH)_2 under palladium catalysis.

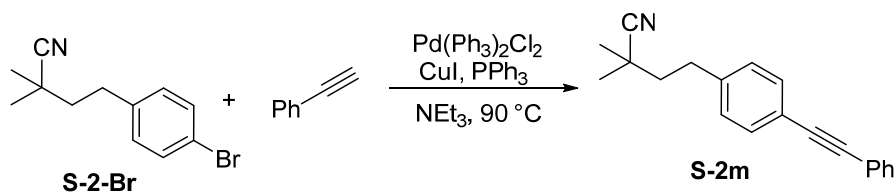
To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **S-2-Br** (1.52 g, 6.0 mmol), phenylboronic acid (0.88 g, 7.2 mmol), $\text{Pd(PPh}_3)_4$ (350 mg, 5 mol%) and Na_2CO_3 (2.6 g, 24 mmol). The tube was evacuated and backfilled with argon for three times, then toluene (15.0 mL) and water (15.0 mL) were added *via* syringe. The tube was stirred at 80 °C overnight. After completion, the reaction mixture was diluted with ethyl acetate (30 mL) and filtered through celite. The filtrate was washed with water and brine. After drying with MgSO_4 and removal of the solvents under reduced pressure, the resulting crude material was purified by flash column chromatography to yield 1.27 g of 4-([1,1'-biphenyl]-4-yl)-2,2-dimethylbutanenitrile **S-2f** in 85% isolated yield.

S-2h (90% yield) was prepared from **S-2-Br** and correspond (*E*)-styrylboronic acid with the similar procedure.

To a flame-dried Schlenk tube equipped with a magnetic stir bar was added **S-2-Br** (1.3 g, 5 mmol), and furan-2-ylboronic acid (1.4 g, 12.5 mmol) or thiophen-2-ylboronic acid (1.6 g, 12.5 mmol) were dissolved in dimethoxyethane (DME) (20 mL) under argon atmosphere. Then 2 M aq K_2CO_3 (1.25 mL, 12.5 mmol) was successively added and the mixture was degassed for 10 min. Finally, $\text{Pd(PPh}_3)_4$ (578.0 mg, 10 mmol%) was added under argon atmosphere and the mixture was stirred at 85 °C overnight. The resulting mixture was cooled to ambient temperature and diluted with H_2O and the organic materials were extracted thrice with ethyl acetate. The combined organic extracts were dried over Na_2SO_4 and concentrated in vacuo after filtration. The resulting residue was purified by flash chromatography (petroleum ether/ ethyl acetate =100/1–50/1) to give the 4-(4-(furan-2-yl)phenyl)-2,2-dimethylbutanenitrile **S-2p** (1.0 g, 4.2 mmol) or 2,2-dimethyl-4-(4-(thiophen-2-yl)phenyl)butanenitrile **S-2q** (1.23 g, 4.8 mmol) in 84% and 96% isolated yields respectively.

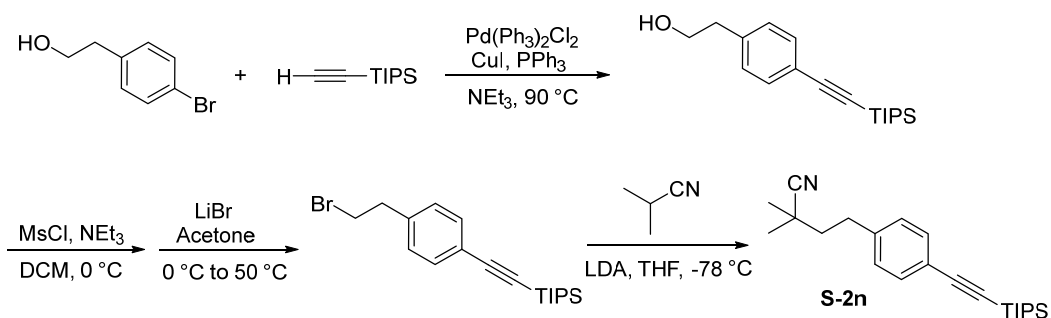
General procedure for the synthesis of alkynylated starting materials:

Procedure for the synthesis of **S-2m**



To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **S-2-Br** (1.26 g, 5.0 mmol), CuI (53 mg 5.5 mol%), Pd(PPh₃)₂Cl₂ (176 mg, 5 mol%) and PPh₃ (132 mg, 10 mol%). The tube was evacuated and backfilled with argon for three times, then ethynylbenzene (0.7 mL, 6.0 mmol) and dry Et₃N (12.0 mL) were added *via* syringe. The tube was stirred at 90 °C overnight. After completion, the reaction mixture was diluted with ethyl acetate (30 mL) and filtered through celite. The filtrate was washed with water and brine. After drying with MgSO₄ and removal of the solvents under reduced pressure, the resulting crude material was purified by flash column chromatography to yield 1.35 g of 2,2-dimethyl-4-(4-(phenylethynyl)phenyl)butanenitrile **S-2m** in 99% isolated yield.

Procedure for the synthesis of S-2n.

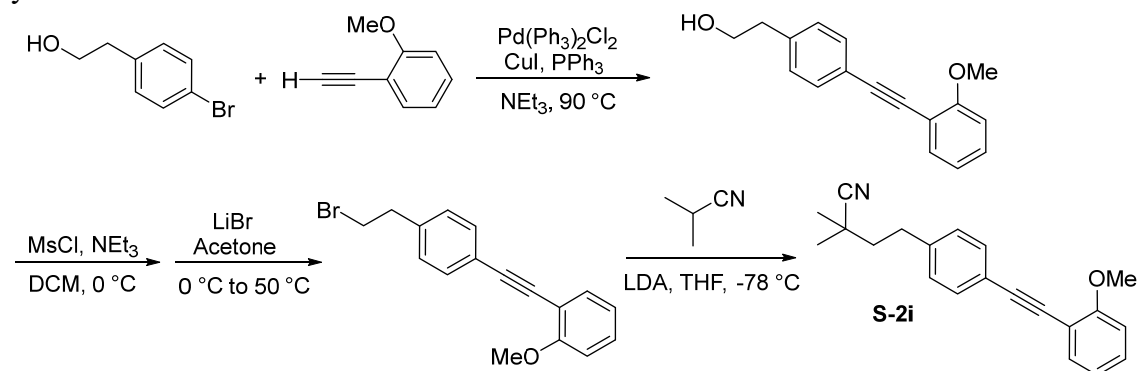


To a round bottom flask equipped with a magnetic stir bar was added 2-(4-bromophenyl)ethan-1-ol (3.0 g, 15 mmol), Pd(PPh₃)₂Cl₂ (526.4 mg, 5 mmol%), CuI (157.1 mg, 5.5 mmol%) and PPh₃ (393.4 mg, 10 mmol%). The flask was evacuated and backfilled with argon for three times. Then anhydrous Et₃N (30 mL) was added *via* syringe under argon atmosphere. Finally, ethynyltriisopropylsilane was dropwise added under argon atmosphere and the mixture was stirred for 60 h at 90 °C. The crude reaction mixture was then filtered through a short celite/SiO₂ pad, and neutralized by using saturated NH₄Cl aqueous solution (50 mL x 2). The aqueous phase was extracted with ethyl acetate (30 mL x 3), and the combined organic phases were washed with brine (20 mL), dried over Na₂SO₄, filtered, and concentrated under reduced pressure to give the desired crude products. Purification by flash column chromatography (petroleum ether/ ethyl acetate =30/1–10/1) to give the 2-(4-((triisopropylsilyl)ethynyl)phenyl)ethan-1-ol (2.5 g, 8.4 mmol) in 56% isolated yields. To an ice cooled solution of above propargylic alcohol and Et₃N (1.8 mL, 12.6 mmol) in dry DCM (25 mL) was added methanesulfonyl chloride (0.7 mL, 12.6 mmol) *via* syringe at 0 °C under argon atmosphere. The mixture was stirred for 45 min at 0 °C before the reaction was quenched with H₂O. The organic materials were washed with saturated NaHCO₃ aqueous solution, then extracted twice with DCM. The combined organic extracts were washed with brine, dried over Na₂SO₄, and concentrated in vacuo

after filtration.

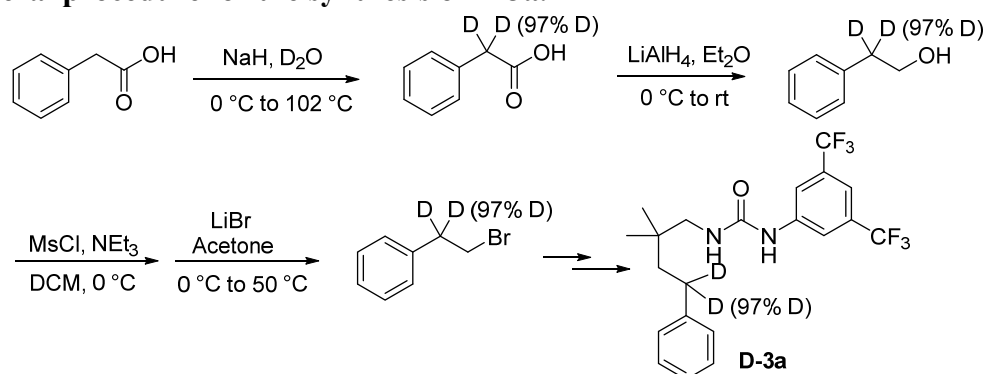
The resulting crude material including mesylate was then, without purification, dissolved in acetone (25 mL) before addition of lithium bromide (3.3 g, 37.8 mmol) at 0 °C. The mixture was stirred at 50 °C overnight. The reaction was then cooled to room temperature and acetone was removed under reduced pressure. The resulting residue was diluted with H₂O and the organic materials were extracted thrice with ethyl acetate. The combined organic extracts were dried over Na₂SO₄ and concentrated in vacuo after filtration. The resulting residue was purified by flash column chromatography (petroleum ether/ ethyl acetate =100/1–30/1) to give the ((4-(2-bromoethyl)phenyl)ethynyl)triisopropylsilane (3.0 g, 8.2 mmol) in 98% overall isolated yield in 2 steps.

S-2n was prepared from ((4-(2-bromoethyl)phenyl)ethynyl)triisopropylsilane and isobutyronitrile under LDA conditions with the similar procedure shown above in 70% yield.



S-2i was prepared from 2-(4-bromophenyl)ethan-1-ol and corresponding 1-ethynyl-2-methoxybenzene in 45% yield over three steps.

General procedure for the synthesis of **D-3a**.



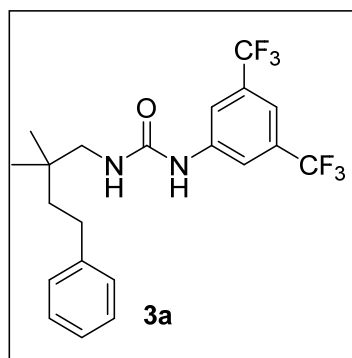
In a three-neck 100 mL round bottom flask equipped with a magnetic stir bar, reflux condenser, and rubber septum, NaH (1.7 g, 42 mmol) was added slowly to D₂O (12 mL) at 0 °C under argon atmosphere. Phenylacetic acid (4.1 g, 30 mmol) was added, and the mixture was heated at reflux at 100 °C for 4 days under argon. The reaction mixture was acidified with 4 N HCl to a pH of 4 and extracted thrice with DCM. The combined organic extracts were dried with Na₂SO₄, filtered, and concentrated to give a quantitative yield of phenylacetic acid with 93% deuterium incorporation in the benzylic position. This procedure was repeated to give 97% deuterated product.

To a stirred ice-cooled suspension of lithium aluminum hydride (1.1 g, 36 mmol) in 80 mL of anhydrous diethyl ether was dropwise added of the preceding acid (4.1 g, 30 mmol) in 20 mL of anhydrous diethyl ether. The hydride mixture was stirred at room temperature overnight. Excess hydride was quenched by adding 8 mL of 10% aqueous NaOH. The mixture was filtered and organic layer was washed with brine, extracted with EtOAc. The combined organic extracts were dried with Na₂SO₄, filtered, and concentrated to give the 2-phenylethan-2,2-*d*₂-1-ol (3.4 g, 27.5 mmol, 97% deuterium incorporation) in 92% yield.

(2-bromoethyl-1,1-*d*₂)benzene was prepared from 2-phenylethan-2,2-*d*₂-1-ol with the similar procedures for the synthesis of 1-bromo-4-(2-bromoethyl)benzene.

D-3a was prepared from (2-bromoethyl-1,1-*d*₂)benzene with the similar procedure mentioned above.

1-(3,5-bis(trifluoromethyl)phenyl)-3-(2,2-dimethyl-4-phenylbutyl)urea



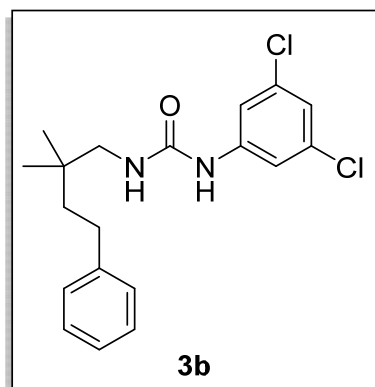
3a, 72% yield, ¹H NMR (400 MHz, Chloroform-*d*): δ 8.50 (s, 1H), 7.62 (s, 2H), 7.36 (s, 1H), 7.19 (t, *J* = 7.3 Hz, 2H), 7.13 – 7.09 (m, 1H), 7.06 (d, *J* = 6.8 Hz, 2H), 6.16 (t, *J* = 6.1 Hz, 1H), 3.08 (d, *J* = 6.0 Hz, 2H), 2.51 – 2.46 (m, 2H), 1.46 – 1.42 (m, 2H), 0.88 (s, 6H).

¹³C NMR (101 MHz, CDCl₃): δ 156.6, 142.4, 140.3, 132.2 (q, *J*_{C-F} = 33.1 Hz), 128.4, 128.1, 125.8, 123.0 (q, *J*_{C-F} = 271.1 Hz), 118.7 – 118.6 (m, 1C), 115.9 – 115.8 (m, 1C), 50.1, 42.0, 34.5, 30.4, 24.5.

¹⁹F NMR (376 MHz, CDCl₃): δ -63.06.

HRMS (ESI) *m/z* calcd. for C₂₁H₂₃F₆N₂O [M+H]⁺ 433.1709, found 433.1703.

1-(3,5-dichlorophenyl)-3-(2,2-dimethyl-4-phenylbutyl)urea

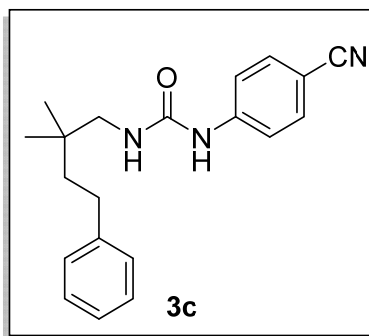


3b, 56% yield, ¹H NMR (400 MHz, Chloroform-*d*): δ 7.61 (s, 1H), 7.25 – 7.21 (m, 2H), 7.16 – 7.14 (m, 3H), 7.12 – 7.10 (m, 2H), 6.92 (t, *J* = 1.8 Hz, 1H), 5.62 (t, *J* = 6.2 Hz, 1H), 3.08 (d, *J* = 6.1 Hz, 2H), 2.54 – 2.50 (m, 2H), 1.47 – 1.43 (m, 2H), 0.90 (s, 6H).

¹³C NMR (101 MHz, CDCl₃): δ 155.9, 142.7, 140.9, 135.1, 128.4, 128.2, 125.8, 122.7, 117.6, 50.0, 42.0, 34.6, 30.4, 24.8.

HRMS (ESI) *m/z* calcd. for C₁₉H₂₃Cl₂N₂O [M+H]⁺ 365.1182, found 365.1177.

11-(4-cyanophenyl)-3-(2,2-dimethyl-4-phenylbutyl)urea



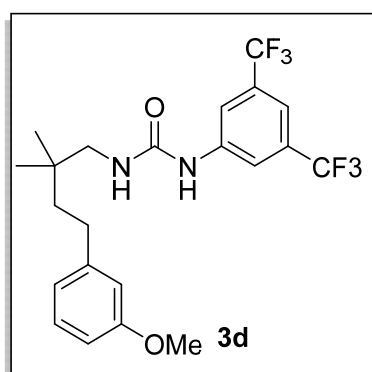
3c, 52% yield, ^1H NMR (400 MHz, Chloroform-*d*): δ 7.66 (s, 1H), 7.44 (s, 4H), 7.25 – 7.21 (m, 2H), 7.17 – 7.11 (m, 3H), 5.48 (t, J = 6.0 Hz, 1H), 3.13 (d, J = 6.0 Hz, 2H), 2.56 – 2.52 (m, 2H), 1.51 – 1.46 (m, 2H), 0.93 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3): δ 155.9, 144.1, 142.6, 133.3, 128.5, 128.2, 125.9, 119.7, 118.4, 104.1, 49.9, 41.9, 34.6, 30.5, 24.9.

HRMS (ESI) m/z calcd. for $\text{C}_{20}\text{H}_{24}\text{N}_3\text{O}$ $[\text{M}+\text{H}]^+$

322.1914, found 322.1909.

1-(3,5-bis(trifluoromethyl)phenyl)-3-(4-(3-methoxyphenyl)-2,2-dimethylbutyl)urea



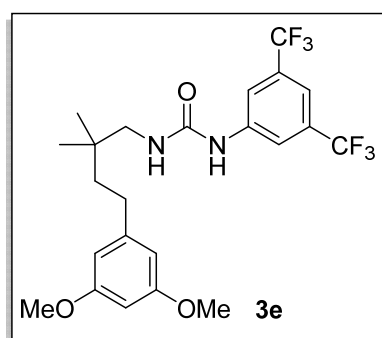
3d, 66% yield, ^1H NMR (400 MHz, Chloroform-*d*): δ 7.72 – 7.65 (m, 3H), 7.42 – 7.41 (m, 1H), 7.16 (t, J = 7.8 Hz, 1H), 6.73 – 6.68 (m, 3H), 5.47 (t, J = 6.1 Hz, 1H), 3.78 (s, 3H), 3.10 (d, J = 6.0 Hz, 2H), 2.53 – 2.48 (m, 2H), 1.46 – 1.41 (m, 2H), 0.91 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3): δ 159.6, 156.3, 144.3, 140.4, 132.1 (q, $J_{\text{C-F}}$ = 33.1 Hz), 129.4, 122.8 (q, $J_{\text{C-F}}$ = 271.1 Hz), 120.7, 118.6 – 118.5 (m, 1C), 115.7(8) – 115.7(4) (m, 1C), 114.3, 110.8, 55.1, 50.1, 41.9, 34.5, 30.4, 24.5.

^{19}F NMR (376 MHz, CDCl_3): δ -63.30.

HRMS (ESI) m/z calcd. for $\text{C}_{22}\text{H}_{25}\text{F}_6\text{N}_2\text{O}_2$ $[\text{M}+\text{H}]^+$ 463.1815, found 463.1808

1-(3,5-bis(trifluoromethyl)phenyl)-3-(4-(3,5-dimethoxyphenyl)-2,2-dimethylbutyl)urea



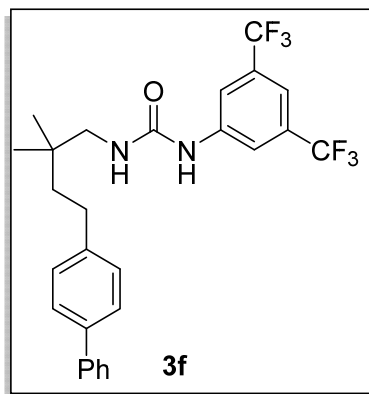
3e, 70% yield, ^1H NMR (400 MHz, Chloroform-*d*): δ 8.05 (s, 1H), 7.71 (s, 2H), 7.39 (s, 1H), 6.30 – 6.27 (m, 3H), 5.72 (t, J = 6.1 Hz, 1H), 3.72 (s, 6H), 3.09 (d, J = 6.0 Hz, 2H), 2.48 – 2.44 (m, 2H), 1.45 – 1.40 (m, 2H), 0.89 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3): δ 160.8, 156.1, 145.2, 140.5, 132.1 (q, $J_{\text{C-F}}$ = 33.1 Hz), 123.1 (q, $J_{\text{C-F}}$ = 271.1 Hz), 118.5 – 118.4 (m, 1C), 115.8 – 115.7 (m, 1C), 106.5, 97.5, 55.2, 50.0, 41.7, 34.5, 30.7, 24.5.

^{19}F NMR (376 MHz, CDCl_3): δ -63.28.

HRMS (ESI) m/z calcd. for $\text{C}_{23}\text{H}_{27}\text{F}_6\text{N}_2\text{O}_3$ $[\text{M}+\text{H}]^+$ 493.1920, found 493.1915.

1-(4-([1,1'-biphenyl]-4-yl)-2,2-dimethylbutyl)-3-(3,5-bis(trifluoromethyl)phenyl)urea

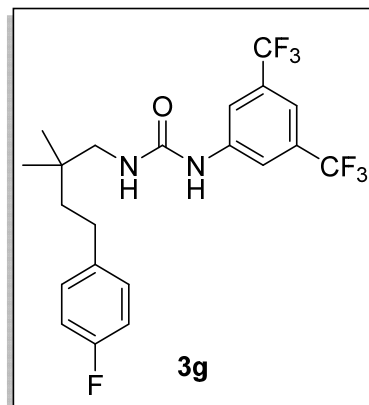


3f, 70% yield, ^1H NMR (400 MHz, Chloroform-*d*): δ 7.80 (s, 2H), 7.56 – 7.53 (m, 2H), 7.50 – 7.40 (m, 5H), 7.34 – 7.30 (m, 1H), 7.22 (d, J = 8.0 Hz, 2H), 7.17 (s, 1H), 5.11 (t, J = 6.2 Hz, 1H), 3.17 (d, J = 6.1 Hz, 2H), 2.63 – 2.59 (m, 2H), 1.56 – 1.52 (m, 2H), 0.97 (s, 6H). ^{13}C NMR (101 MHz, Acetone-*d*₆): δ 155.1, 142.8, 142.6, 140.9, 138.3, 131.5 (q, $J_{\text{C-F}}$ = 33.1 Hz), 128.9, 128.8, 127.0, 126.8, 126.6, 123.7 (q, $J_{\text{C-F}}$ = 271.1 Hz), 117.5 – 117.4 (m, 1C), 113.9 – 113.7 (m, 1C), 48.9, 41.9, 34.6, 29.9, 24.5.

^{19}F NMR (376 MHz, Acetone-*d*₆) δ -63.58.

HRMS (ESI) m/z calcd. for $\text{C}_{27}\text{H}_{27}\text{F}_6\text{N}_2\text{O}$ $[\text{M}+\text{H}]^+$ 509.2022, found 509.2018.

1-(3,5-bis(trifluoromethyl)phenyl)-3-(4-(4-fluorophenyl)-2,2-dimethylbutyl)urea



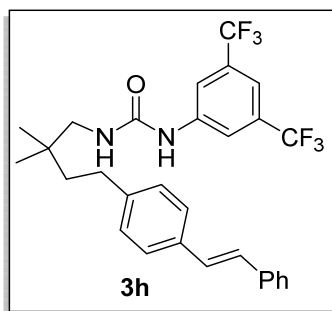
3g, 62% yield, ^1H NMR (400 MHz, Chloroform-*d*): δ 7.84 (s, 2H), 7.49 (s, 1H), 7.13 – 7.09 (m, 2H), 6.96 – 6.91 (m, 2H), 6.86 (s, 1H), 4.90 (t, J = 6.2 Hz, 1H), 3.17 (d, J = 6.2 Hz, 2H), 2.59 – 2.55 (m, 2H), 1.51 – 1.47 (m, 2H), 0.97 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3): δ 161.2 (d, $J_{\text{C-F}}$ = 241.9 Hz), 156.6, 140.3, 138.0 (d, $J_{\text{C-F}}$ = 3.5 Hz), 132.2 (q, $J_{\text{C-F}}$ = 33.1 Hz), 129.4 (d, $J_{\text{C-F}}$ = 7.8 Hz), 123.0 (q, $J_{\text{C-F}}$ = 271.1 Hz), 118.6 – 118.5 (m, 1C), 115.9 – 115.8 (m, 1C), 115.1 (d, $J_{\text{C-F}}$ = 21.0 Hz), 50.1, 42.1, 34.4, 29.5, 24.5.

^{19}F NMR (376 MHz, CDCl_3): δ -63.10, -117.75.

HRMS (ESI) m/z calcd. for $\text{C}_{21}\text{H}_{22}\text{F}_7\text{N}_2\text{O}$ $[\text{M}+\text{H}]^+$ 451.1615, found 154.1609.

(*E*)-1-(3,5-bis(trifluoromethyl)phenyl)-3-(2,2-dimethyl-4-(4-styrylphenyl)butyl)urea



3h, 70% yield, ^1H NMR (400 MHz, Acetone-*d*₆): δ 8.58 (s, 1H), 8.17 (m, 2H), 7.59 – 7.57 (m, 2H), 7.53 – 7.49 (m, 3H), 7.38 – 7.34 (m, 2H), 7.26 – 7.22 (m, 3H), 7.20 (d, J = 4.6 Hz, 2H), 6.18 (t, J = 6.6 Hz, 1H), 3.21 (d, J = 6.3 Hz, 2H), 2.69 – 2.64 (m, 2H), 1.58 – 1.53 (m, 2H), 1.00 (s, 6H).

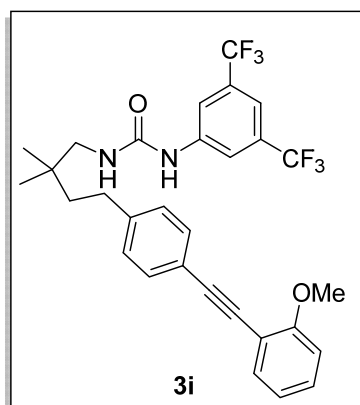
^{13}C NMR (101 MHz, Acetone-*d*₆): δ 155.1, 143.0, 142.8, 137.6, 134.9, 131.5 (q, $J_{\text{C-F}}$ = 33.3 Hz), 128.7, 128.6, 128.5, 127.6, 127.3, 126.5, 126.4, 123.6 (q, $J_{\text{C-F}}$ = 270.3 Hz), 117.5 – 117.4 (m, 1C), 113.9 – 113.7 (m, 1C), 48.9, 41.8, 34.6,

30.1, 24.5.

^{19}F NMR (376 MHz, Acetone-*d*₆): δ -63.55.

HRMS (ESI) m/z calcd. for $\text{C}_{29}\text{H}_{29}\text{F}_6\text{N}_2\text{O}$ $[\text{M}+\text{H}]^+$ 535.2179, found 535.2179.

1-(3,5-bis(trifluoromethyl)phenyl)-3-(4-(4-((2-methoxyphenyl)ethynyl)phenyl)-2,2-dimethylbutyl)urea



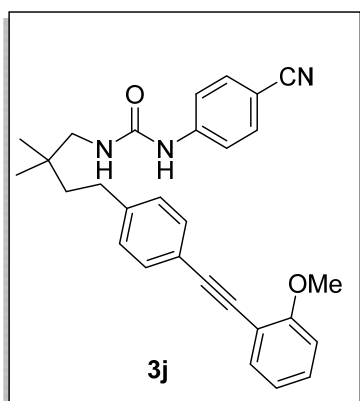
3i, 52% yield, ^1H NMR (400 MHz, Chloroform-*d*): δ 7.77 – 7.71 (m, 3H), 7.49 (dd, J = 7.6, 1.6 Hz, 1H), 7.41 – 7.38 (m, 3H), 7.34 – 7.29 (m, 1H), 7.05 (d, J = 8.0 Hz, 2H), 6.97 – 6.90 (m, 2H), 5.53 – 5.48 (m, 1H), 3.89 (s, 3H), 3.08 (d, J = 6.0 Hz, 2H), 2.53 – 2.49 (m, 2H), 1.42 – 1.38 (m, 2H), 0.90 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3): δ 159.7, 156.2, 143.0, 140.4, 133.7, 132.2 (q, $J_{\text{C-F}}$ = 33.1 Hz), 131.7, 129.9, 128.2, 123.1 (q, $J_{\text{C-F}}$ = 271.1 Hz), 120.8, 120.7, 118.4, 115.8 – 115.6 (m, 1C), 112.4, 110.8, 93.7, 85.3, 55.8, 50.1, 41.5, 34.5, 30.3, 24.5.

^{19}F NMR (376 MHz, CDCl_3): δ -63.14.

HRMS (ESI) m/z calcd. for $\text{C}_{30}\text{H}_{29}\text{F}_6\text{N}_2\text{O}_2$ $[\text{M}+\text{H}]^+$ 563.2128, found 563.2123.

1-(4-cyanophenyl)-3-(4-(4-((2-methoxyphenyl)ethynyl)phenyl)-2,2-dimethylbutyl)urea

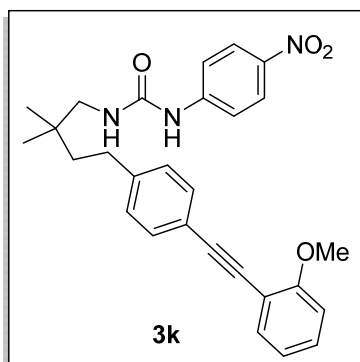


3j, 54% yield, ^1H NMR (400 MHz, Chloroform-*d*): δ 8.00 (s, 1H), 7.48 – 7.37 (m, 7H), 7.31 – 7.27 (m, 1H), 7.05 (d, J = 8.0 Hz, 2H), 6.95 – 6.87 (m, 2H), 5.65 (t, J = 6.4 Hz, 1H), 3.87 (s, 3H), 3.08 (d, J = 6.0 Hz, 2H), 2.53 – 2.48 (m, 2H), 1.44 – 1.40 (m, 2H), 0.90 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3): δ 159.7, 155.5, 144.1, 143.1, 133.6, 133.3, 131.7, 129.9, 128.3, 120.8, 119.7, 118.3, 112.4, 110.9, 104.1, 93.7, 85.4, 55.9, 49.8, 41.5, 34.6, 30.4, 24.9.

HRMS (ESI) m/z calcd. for $\text{C}_{29}\text{H}_{30}\text{N}_3\text{O}_2$ $[\text{M}+\text{H}]^+$ 452.2333, found 452.2327.

1-(4-(4-((2-methoxyphenyl)ethynyl)phenyl)-2,2-dimethylbutyl)-3-(4-nitrophenyl)urea



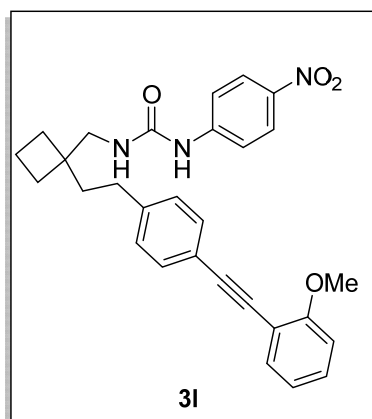
3k, 50% yield, ^1H NMR (400 MHz, Chloroform-*d*): δ 8.07 (d, J = 9.2 Hz, 2H), 7.59 (s, 1H), 7.52 – 7.48 (m, 3H), 7.39 (d, J = 8.2 Hz, 2H), 7.33 – 7.29 (m, 1H), 7.08 (d, J = 8.1 Hz, 2H), 6.97 – 6.90 (m, 2H), 5.27 (t, J = 6.1 Hz, 1H), 3.91 (s, 3H), 3.13 (d, J = 6.1 Hz, 2H), 2.57 – 2.53 (m, 2H), 1.46 – 1.42 (m, 2H), 0.94 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3): δ 159.6, 154.9, 145.9, 143.2, 141.7, 133.7, 131.7, 129.9, 128.3, 125.3, 120.9, 120.7, 117.6, 112.4, 111.0, 93.7, 85.3, 56.0, 50.0, 41.4, 34.6, 30.4, 24.8.

HRMS (ESI) m/z calcd. for $\text{C}_{28}\text{H}_{30}\text{N}_3\text{O}_4$ $[\text{M}+\text{H}]^+$ 472.2231, found 472.2227.

1-(4-(4-((2-methoxyphenyl)ethynyl)phenyl)cyclobutyl)methyl)-3-(4-nitrophenyl)urea

urea



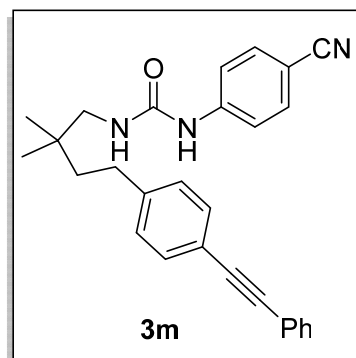
3l, 62% yield, ^1H NMR (400 MHz, Chloroform-*d*): δ 8.15 (s, 1H), 8.07 (d, J = 9.2 Hz, 2H), 7.54 (d, J = 9.2 Hz, 2H), 7.47 (dd, J = 7.6, 1.7 Hz, 1H), 7.38 (d, J = 8.1 Hz, 2H), 7.29 (ddd, J = 8.4, 7.5, 1.7 Hz, 1H), 7.07 (d, J = 8.2 Hz, 2H), 6.93 (td, J = 7.5, 1.0 Hz, 1H), 6.89 (dd, J = 8.4, 1.0 Hz, 1H), 5.72 (t, J = 5.8 Hz, 1H), 3.88 (s, 3H), 3.38 (d, J = 5.7 Hz, 2H), 2.57 – 2.44 (m, 2H), 1.94 – 1.82 (m, 2H), 1.76 (t, J = 7.5 Hz, 4H), 1.72 – 1.59 (m, 2H).

^{13}C NMR (101 MHz, CDCl_3): δ 159.6, 155.0, 145.9, 142.9, 141.8, 133.7, 131.7, 129.9, 128.3, 125.3, 120.8,

120.7, 117.5, 112.4, 110.9, 93.7, 85.3, 55.9, 46.2, 41.8, 39.1, 30.3, 29.2, 15.1.

HRMS (ESI) m/z calcd. for $\text{C}_{29}\text{H}_{30}\text{N}_3\text{O}_4$ $[\text{M}+\text{H}]^+$ 484.2231, found 484.2228.

1-(4-cyanophenyl)-3-(2,2-dimethyl-4-(4-(phenylethynyl)phenyl)butyl)urea



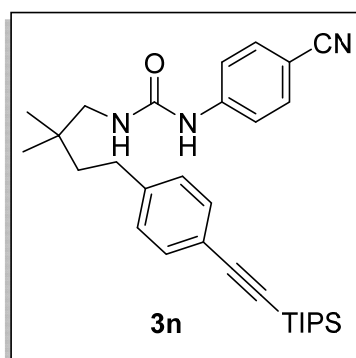
3m, 57% yield, ^1H NMR (400 MHz, Chloroform-*d*): δ 7.69 (s, 1H), 7.51 – 7.49 (m, 2H), 7.44 (s, 4H), 7.39 (d, J = 7.8 Hz, 2H), 7.34 – 7.31 (m, 3H), 7.09 (d, J = 7.8 Hz, 2H), 5.49 (t, J = 6.0 Hz, 1H), 3.11 (d, J = 6.1 Hz, 2H), 2.56 – 2.52 (m, 2H), 1.48 – 1.44 (m, 2H), 0.92 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3): δ 155.3, 143.9, 143.1, 133.3, 131.7, 131.6, 128.4, 128.3, 128.2, 123.3, 120.7, 119.6, 118.4, 104.3, 89.4, 89.1, 49.8, 41.6, 34.6, 30.5, 24.9.

HRMS (ESI) m/z calcd. for $\text{C}_{28}\text{H}_{28}\text{N}_3\text{O}$ $[\text{M}+\text{H}]^+$

422.2227, found 422.2220.

1-(4-cyanophenyl)-3-(2,2-dimethyl-4-(4-((triisopropylsilyl)ethynyl)phenyl)butyl)urea

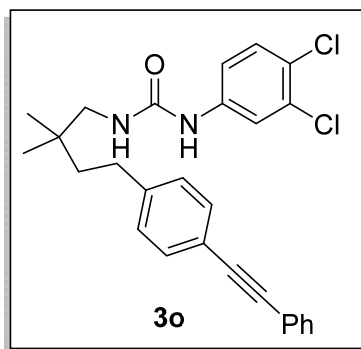


3n, 58% yield, ^1H NMR (400 MHz, Chloroform-*d*): δ 7.48 (q, J = 8.8 Hz, 4H), 7.36 (d, J = 8.4 Hz, 2H), 7.33 (s, 1H), 7.07 (d, J = 8.0 Hz, 2H), 5.24 (t, J = 6.0 Hz, 1H), 3.13 (d, J = 6.0 Hz, 2H), 2.57 – 2.53 (m, 2H), 1.48 – 1.44 (m, 2H), 1.12 (s, 21H), 0.94 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3): δ 155.0, 143.7, 143.2, 133.3, 132.1, 128.2, 121.0, 119.4, 118.4, 107.1, 104.6, 90.0, 49.8, 41.7, 34.6, 30.5, 24.9, 18.7, 11.3.

HRMS (ESI) m/z calcd. for $\text{C}_{31}\text{H}_{44}\text{N}_3\text{OSi}$ $[\text{M}+\text{H}]^+$ 502.3254, found 502.3251.

1-(3,4-dichlorophenyl)-3-(2,2-dimethyl-4-(4-(phenylethynyl)phenyl)butyl)urea



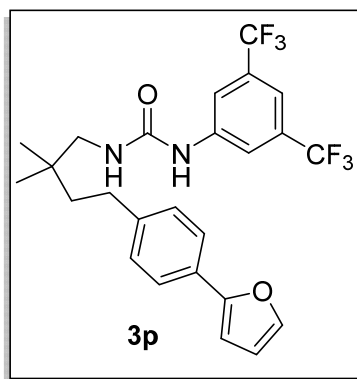
3o, 60% yield, ^1H NMR (400 MHz, Acetone- d_6) δ 8.20 (s, 1H), 8.00 (d, J = 2.4 Hz, 1H), 7.62 – 7.53 (m, 2H), 7.50 – 7.39 (m, 6H), 7.35 (dd, J = 8.8, 2.5 Hz, 1H), 7.34 – 7.29 (m, 2H), 6.04 (t, J = 6.3 Hz, 1H), 3.21 (d, J = 6.4 Hz, 2H), 2.77 – 2.67 (m, 2H), 1.63 – 1.51 (m, 2H), 1.02 (s, 6H).

^{13}C NMR (101 MHz, Acetone) δ 155.1, 144.3, 141.0, 131.7, 131.4, 131.3, 130.3, 128.7, 128.6, 128.3, 123.4, 123.2, 120.3, 119.2, 117.7, 89.4, 88.5, 48.7, 41.6, 34.6,

30.3, 24.5.

HRMS (ESI) m/z calcd. for $\text{C}_{27}\text{H}_{27}\text{Cl}_2\text{N}_2\text{O}$ $[\text{M}+\text{H}]^+$ 465.1495, found 465.1496.

1-(3,5-bis(trifluoromethyl)phenyl)-3-(4-(4-(furan-2-yl)phenyl)-2,2-dimethylbutyl) urea



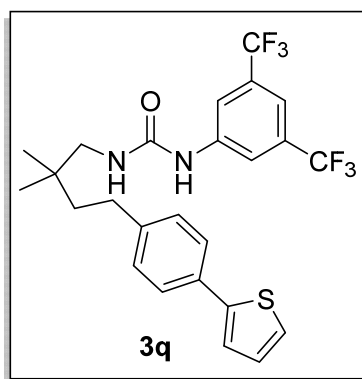
3p, 65% yield, ^1H NMR (400 MHz, Acetone- d_6): δ 8.56 (s, 1H), 8.17 (s, 2H), 7.62 (s, 1H), 7.60 – 7.59 (m, 2H), 7.53 (s, 1H), 7.28 (d, J = 7.9 Hz, 2H), 6.76 (d, J = 3.4 Hz, 1H), 6.53 – 6.52 (m, 1H), 6.17 (s, 1H), 3.21 (d, J = 6.3 Hz, 2H), 2.70 – 2.65 (m, 2H), 1.58 – 1.54 (m, 2H), 1.00 (s, 6H).

^{13}C NMR (101 MHz, Acetone): δ 155.1, 154.0, 142.8, 142.7, 142.0, 131.5 (q, $J_{\text{C-F}}$ = 32.6 Hz), 128.7, 128.5, 123.6 (q, $J_{\text{C-F}}$ = 271.0 Hz), 123.5, 117.5, 113.8, 111.6, 104.5, 48.9, 41.8, 34.6, 30.1, 24.5.

^{19}F NMR (376 MHz, Acetone): δ -63.59.

HRMS (ESI) m/z calcd. for $\text{C}_{25}\text{H}_{25}\text{F}_6\text{N}_2\text{O}_2$ $[\text{M}+\text{H}]^+$ 499.1820, found 499.1821.

1-(3,5-bis(trifluoromethyl)phenyl)-3-(2,2-dimethyl-4-(4-(thiophen-2-yl)phenyl) butyl)urea



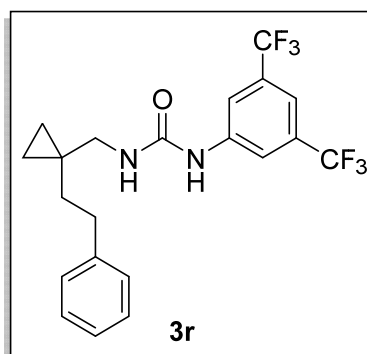
3q, 66% yield, ^1H NMR (400 MHz, CDCl_3): δ 8.26 (s, 1H), 7.65 (s, 2H), 7.43 (d, J = 7.8 Hz, 2H), 7.37 (s, 1H), 7.22 - 7.19 (m, 2H), 7.07 – 7.01 (m, 3H), 5.95 (t, J = 6.1 Hz, 1H), 3.09 (d, J = 5.9 Hz, 2H), 2.51 – 2.46 (m, 2H), 1.47 – 1.42 (m, 2H), 0.88 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3): δ 156.3, 144.2, 141.9, 140.3, 132.2 (q, $J_{\text{C-F}}$ = 33.0 Hz), 132.1, 128.7, 128.0, 125.9, 124.4, 123.0 (q, $J_{\text{C-F}}$ = 270.9 Hz), 122.7, 118.6, 115.9, 50.1, 41.9, 34.5, 30.1, 24.6.

^{19}F NMR (376 MHz, CDCl_3): δ -63.27.

HRMS (ESI) m/z calcd. for $\text{C}_{25}\text{H}_{25}\text{F}_6\text{N}_2\text{OS}$ $[\text{M}+\text{H}]^+$ 515.1592, found 515.1592.

1-(3,5-bis(trifluoromethyl)phenyl)-3-((1-phenethylcyclopropyl)methyl)urea



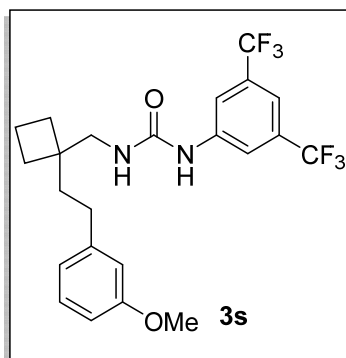
3r, 54% yield, ^1H NMR (400 MHz, Chloroform-*d*): δ 7.76 (s, 1H), 7.72 (s, 2H), 7.40 (s, 1H), 7.22 – 7.18 (m, 2H), 7.14 – 7.13 (m, 1H), 7.10– 7.08 (m, 2H), 5.57 (t, J = 5.5 Hz, 1H), 3.20 (d, J = 5.4 Hz, 2H), 2.67 – 2.63 (m, 2H), 1.58 – 1.54 (m, 2H), 0.38 – 0.31 (m, 4H).

^{13}C NMR (101 MHz, CDCl_3): δ 156.5, 141.8, 140.3, 132.2 (q, $J_{\text{C-F}}$ = 33.1 Hz), 128.4, 128.1, 125.9, 123.1 (q, $J_{\text{C-F}}$ = 271.1 Hz), 118.7, 115.9, 46.0, 36.7, 33.0, 20.0, 10.6.

^{19}F NMR (376 MHz, CDCl_3): δ -63.36.

HRMS (ESI) m/z calcd. for $\text{C}_{21}\text{H}_{21}\text{F}_6\text{N}_2\text{O}$ $[\text{M}+\text{H}]^+$ 431.1553, found 431.1549.

1-(3,5-bis(trifluoromethyl)phenyl)-3-((1-(3-methoxyphenethyl)cyclobutyl)methyl)urea



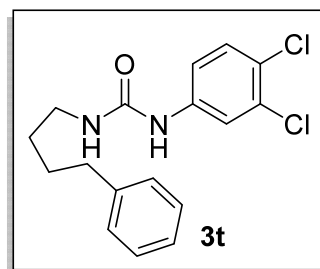
3s, 63% yield, ^1H NMR (400 MHz, CDCl_3): δ 8.09 (s, 1H), 7.67 (s, 2H), 7.38 (s, 1H), 7.14 – 7.10 (m, 1H), 6.70 – 6.66 (m, 3H), 5.80 (t, J = 5.5 Hz, 1H), 3.73 (s, 3H), 3.33 (d, J = 5.7 Hz, 2H), 2.48 – 2.44 (m, 2H), 1.86 – 1.78 (m, 2H), 1.75 – 1.70 (m, 4H), 1.69 – 1.65 (m, 2H).

^{13}C NMR (101 MHz, CDCl_3): δ 159.6, 156.8, 143.9, 140.4, 132.2 (q, $J_{\text{C-F}}$ = 33.1 Hz), 129.4, 123.1 (q, $J_{\text{C-F}}$ = 271.1 Hz), 120.6, 118.7, 115.8, 114.3, 110.8, 55.0, 46.3, 41.8, 39.3, 30.4, 29.0, 15.0.

^{19}F NMR (376 MHz, CDCl_3): δ -63.31.

HRMS (ESI) m/z calcd. for $\text{C}_{23}\text{H}_{25}\text{F}_6\text{N}_2\text{O}_2$ $[\text{M}+\text{H}]^+$ 475.1815, found 475.1813.

1-(3,4-dichlorophenyl)-3-(4-phenylbutyl)urea

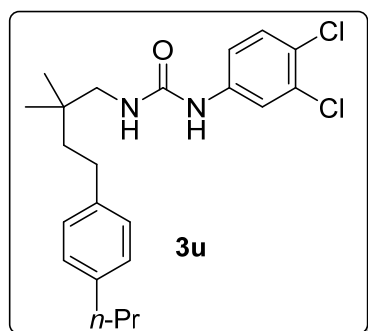


3t, 52% yield, ^1H NMR (400 MHz, Chloroform-*d*) δ 7.44 (d, J = 2.4 Hz, 1H), 7.27 – 7.24 (m, 3H), 7.20 – 7.04 (m, 5H), 5.20 (t, J = 5.7 Hz, 1H), 3.21 – 3.16 (m, 2H), 2.58 (t, J = 7.6 Hz, 2H), 1.63 – 1.56 (m, 2H), 1.52 – 1.46 (m, 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 155.5, 142.0, 138.4, 132.7, 130.5, 128.4, 126.2, 125.9, 121.4, 119.0, 40.2, 35.5, 29.6, 28.6.

HRMS (ESI) m/z calcd. for $\text{C}_{17}\text{H}_{19}\text{Cl}_2\text{N}_2\text{O}$ $[\text{M}+\text{H}]^+$ 337.0869, found 337.0867.

1-(3,4-dichlorophenyl)-3-(2,2-dimethyl-4-(4-propylphenyl)butyl)urea



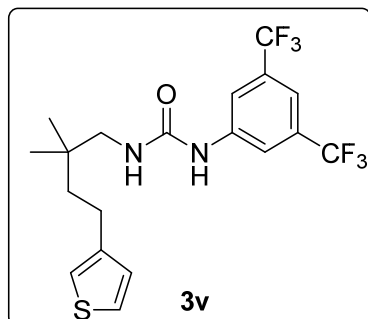
3u, 64% yield, ^1H NMR (400 MHz, Chloroform-*d*): δ 8.28 (s, 1H), 7.39 (d, J = 2.5 Hz, 1H), 7.09 (d, J = 8.6 Hz, 1H), 7.00 (q, J = 8.1 Hz, 4H), 6.88 (dd, J = 8.7, 2.5 Hz, 1H), 6.14 (t, J = 6.2 Hz, 1H), 3.04 (d, J = 5.9 Hz, 2H), 2.56 – 2.38 (m, 4H), 1.57 (dt, J = 14.7, 7.4 Hz, 2H), 1.47 – 1.37 (m, 2H), 0.91 (t, J = 7.3 Hz, 3H), 0.85 (s, 6H).

^{13}C NMR (101 MHz, Chloroform-*d*): δ 157.1, 140.2,

139.8, 138.6, 132.6, 130.4, 128.6, 128.1, 126.1, 121.5, 119.1, 50.1, 42.2, 37.8, 34.7, 30.1, 24.9, 24.8, 14.1.

HRMS (ESI) m/z calcd. for $\text{C}_{22}\text{H}_{29}\text{Cl}_2\text{N}_2\text{O}$ $[\text{M}+\text{H}]^+$ 407.1651, found 407.1650.

1-(3,5-bis(trifluoromethyl)phenyl)-3-(2,2-dimethyl-4-(thiophen-3-yl)butyl)urea



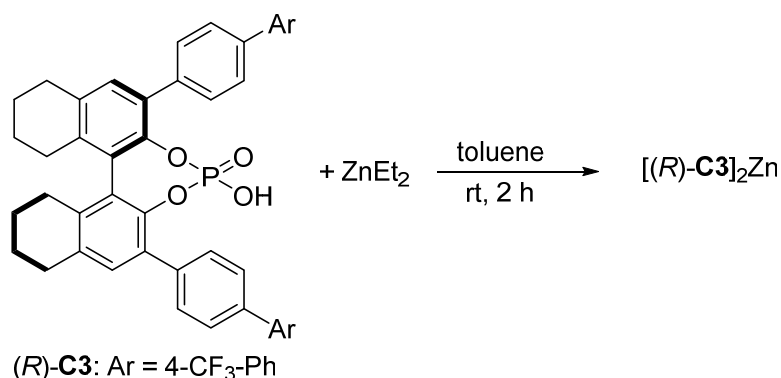
3v, 62% yield, ^1H NMR (400 MHz, Chloroform-*d*) δ 8.30 (s, 1H), 7.64 (s, 2H), 7.37 (s, 1H), 7.16 (dd, J = 4.7, 3.2 Hz, 1H), 6.88 – 6.78 (m, 2H), 6.01 (t, J = 6.1 Hz, 1H), 3.09 (d, J = 6.0 Hz, 2H), 2.61 – 2.46 (m, 2H), 1.57 – 1.42 (m, 2H), 0.89 (s, 6H).

^{13}C NMR (101 MHz, CDCl_3) δ 156.4, 142.6, 140.2, 132.2 (q, $J_{\text{C-F}}$ = 33.1 Hz), 128.0, 125.4, 123.0 (q, $J_{\text{C-F}}$ = 271.0 Hz), 119.6, 118.7, 116.0, 50.1, 40.5, 34.4, 24.7, 24.6.

^{19}F NMR (376 MHz, CDCl_3) δ -63.33.

HRMS (ESI) m/z calcd. for $\text{C}_{19}\text{H}_{21}\text{F}_6\text{N}_2\text{OS}$ $[\text{M}+\text{H}]^+$ 439.1279, found 439.1276.

Procedure for the synthesis of $[(R)\text{-C3}]_2\text{Zn}$.



To a flame-dried round bottom flask equipped with a magnetic stir bar was added (R)-C3 (160 mg, 0.2 mmol) and dry toluene (2 mL) was then added *via* syringe. ZnEt_2 (100 μL , 1.0 M in hexane solution) was added slowly *via* syringe under Ar. The resulting mixture was stirred at room temperature for 2 h. The solvent was removed under reduced pressure to afford 140 mg $[(R)\text{-C3}]_2\text{Zn}$ as brown crystal.

$[(R)\text{-C3}]_2\text{Zn}$

¹H NMR (400 MHz, DMSO-*d*₆) δ 7.95 (dd, *J* = 8.4, 3.0 Hz, 16H), 7.80 (d, *J* = 8.0 Hz, 8H), 7.75 (d, *J* = 8.0 Hz, 8H), 7.18 (s, 4H), 2.93 – 2.76 (m, 8H), 2.70 – 2.59 (m, 4H), 2.21 – 2.14 (m, 4H), 1.81 – 1.73 (m, 12H), 1.58 – 1.53 (m, 4H).

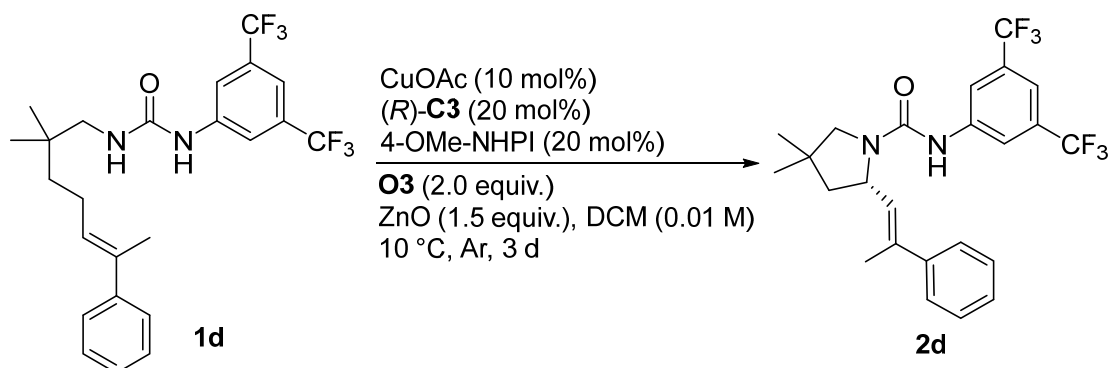
¹³C NMR (101 MHz, DMSO) δ 145.3, 145.2, 144.4, 138.5, 137.2, 136.9, 133.7, 130.9, 130.8, 130.7, 128.5, 128.1 (q, *J* = 31.6 Hz), 127.74, 127.01, 126.2–126.1 (m, 1C), 124.7 (q, *J*_{C-F} = 271.0 Hz), 29.0, 27.9, 22.7, 22.6.

¹⁹F NMR (376 MHz, DMSO) δ -60.85.

³¹P NMR (162 MHz, DMSO) δ 0.02.

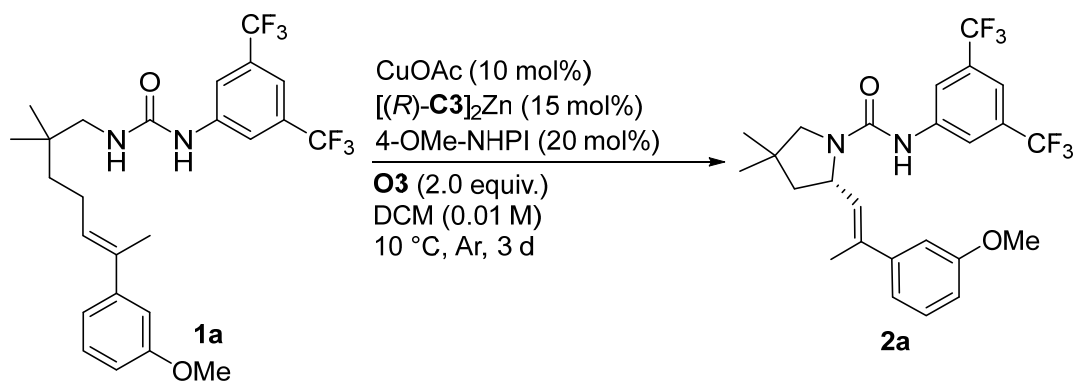
HRMS (ESI) *m/z* calcd. for C₉₂H₆₉F₁₂O₈P₂Zn [M+H]⁺ 1655.3562, found 1655.3566.

General procedure for enantioselective radical allylic C–H amination under conditions A:



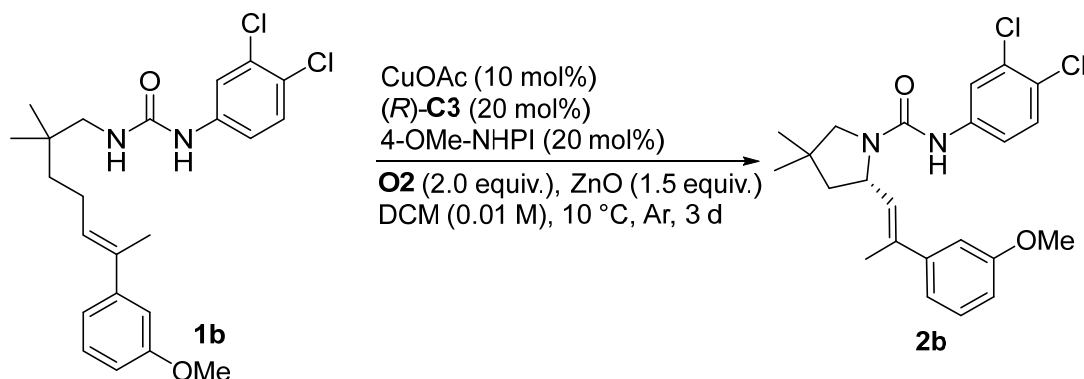
To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **1d** (47.5 0.1 mmol), CuOAc (1.24 mg 10 mol%), (*R*)-**C3** (16 mg, 20 mol%), 4-OMe-NHPI (4.0 mg, 20 mol%), and ZnO (12 mg, 0.15 mmol). The tube was evacuated and backfilled with argon for three times, the **O3** (48 μ L, 0.2 mmol) and dry DCM (10.0 mL) was added *via* syringe. The tube was stirred at 10 °C for 3 days. After completion, the solvent was removed under reduced pressure, and the residue was purified by flash column chromatography (petroleum ether/ ethyl acetate =20/1–10/1) to give the corresponding product **2d** (35.8 mg) in 76% isolated yield.

General procedure for enantioselective radical allylic C–H amination under conditions B:



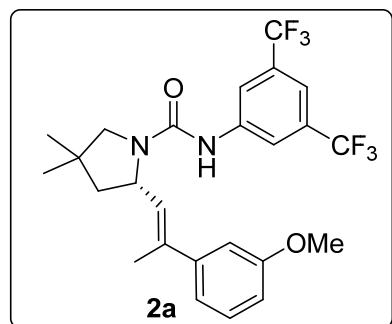
To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **1a** (50.5 mg, 0.1 mmol), CuOAc (1.24 mg 10 mol%), [(*R*)-**C3**]₂Zn (25 mg, 15 mol%) and 4-OMe-NHPI (4.0 mg, 20 mol%). The tube was evacuated and backfilled with argon for three times, the **O3** (48 μ L, 0.2 mmol) and dry DCM (10.0 mL) was added *via* syringe. The tube was stirred at 10 °C for 3 days. After completion, the solvent was removed under reduced pressure, and the residue was purified by flash chromatography (petroleum ether/ ethyl acetate =20/1–10/1) to give the corresponding product **2a** (38 mg) in 76% isolated yield.

Procedure for enantioselective radical allylic C–H amination of **1b**.



To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **1d** (47.5 0.1 mmol), CuOAc (1.24 mg 10 mol%), (*R*)-**C3** (16 mg, 20 mol%), 4-OMe-NHPI (4.0 mg, 20 mol%), and ZnO (12 mg, 0.15 mmol). The tube was evacuated and backfilled with argon for three times, the **O2** (36 μ L, 0.2 mmol) and dry DCM (10.0 mL) was added *via* syringe. The tube was stirred at 10 °C for 3 days. After completion, the solvent was removed under reduced pressure, and the residue was purified by flash column chromatography (petroleum ether/ ethyl acetate =20/1–10/1) to give the corresponding product **2b** (38 mg) in 88% isolated yield.

(*S*)(*E*)-N-(3,5-bis(trifluoromethyl)phenyl)-2-(2-(3-methoxyphenyl)prop-1-en-1-yl)-4,4-dimethylpyrrolidine-1-carboxamide



2a, 76% yield, $[\alpha]_D^{27} = -75.8$ (*c* 1.04, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*) δ 7.79 (s, 2H), 7.47 (s, 1H), 7.35 – 7.28 (m, 1H), 7.07 – 6.99 (m, 2H), 6.95 (t, *J* = 2.1 Hz, 1H), 6.89 (ddd, *J* = 8.2, 2.6, 1.2 Hz, 1H), 5.89 (dd, *J* = 9.0, 1.2 Hz, 1H), 4.75 (td, *J* = 9.0, 7.1 Hz, 1H), 3.84 (s, 3H), 3.67 (d, *J* = 10.6 Hz, 1H), 3.26 (d, *J* = 10.6 Hz, 1H), 2.28 (d, *J* = 1.2 Hz, 3H), 2.11 (ddd, *J* = 12.7, 7.1, 1.6 Hz, 1H), 1.75 (dd, *J* = 12.7,

9.1 Hz, 1H), 1.21 (s, 3H), 1.17 (s, 3H).

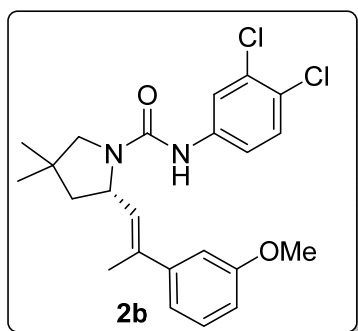
¹³C NMR (101 MHz, CDCl₃) δ (one C signal was overlapped) 159.8, 153.8, 143.4, 140.8, 138.4, 132.1 (q, *J*_{C-F} = 33.1 Hz), 129.7, 123.2 (q, *J*_{C-F} = 271.0 Hz), 118.6 – 118.5 (m, 1C), 118.1, 115.8 – 115.6 (m, 1C), 113.2, 111.7, 59.1, 55.8, 55.3, 47.6, 36.7, 26.3(0), 26.2(7), 16.3.

¹⁹F NMR (376 MHz, CDCl₃) δ -63.07.

HRMS (ESI) *m/z* calcd. for C₂₅H₂₇F₆N₂O₂ [*M*+*H*]⁺ 501.1977, found 501.1976.

HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 0.5 mL/min. λ = 254 nm, *t*(minor) = 10.1 min, *t*(major) = 14.5 min, 95:5 er.

(*S*)(*E*)-N-(3,4-dichlorophenyl)-2-(2-(3-methoxyphenyl)prop-1-en-1-yl)-4,4-dimethylpyrrolidine-1-carboxamide



2b, 88% yield, $[\alpha]_D^{27} = -106.6$ (*c* 0.62, CHCl_3).

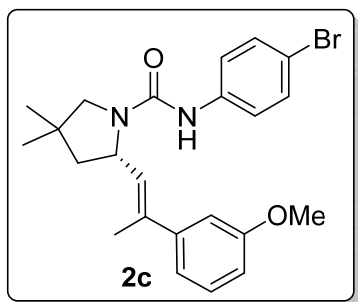
^1H NMR (400 MHz, CHloroform-d) δ 7.59 (d, *J* = 2.5 Hz, 1H), 7.27 (d, *J* = 8.4 Hz, 1H), 7.24 (d, *J* = 8.4 Hz, 1H), 7.04 – 6.97 (m, 2H), 6.92 (t, *J* = 2.0 Hz, 1H), 6.86 (ddd, *J* = 8.2, 2.4, 0.8 Hz, 1H), 6.67 (s, 1H), 5.85 (dq, *J* = 8.8, 1.3 Hz, 1H), 4.68 (td, *J* = 9.0, 7.2 Hz, 1H), 3.82 (s, 3H), 3.66 – 3.63 (m, 1H), 3.21 (d, *J* = 10.6 Hz, 1H), 2.23 (d, *J* = 1.2 Hz, 3H), 2.06 (ddd, *J* = 12.6, 7.1, 1.5 Hz, 1H), 1.69 (dd, *J* = 12.7, 9.0 Hz, 1H), 1.17 (s, 3H), 1.13 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 159.8, 154.0, 143.5, 138.9, 137.9, 132.6, 130.3, 129.9, 129.6, 125.4, 120.5, 118.2, 118.1, 113.1, 111.8, 59.1, 55.6, 55.3, 47.6, 36.7, 26.3, 26.2, 16.2.

HRMS (ESI) *m/z* calcd. for $\text{C}_{23}\text{H}_{27}\text{Cl}_2\text{N}_2\text{O}_2$ $[\text{M}+\text{H}]^+$ 433.1444, found 433.1441.

HPLC condition: Chiralcel AD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 0.5 mL/min. λ = 254 nm, *t*(minor) = 24.0 min, *t*(major) = 28.6 min, 94:6 er.

(S)(E)-N-(4-bromophenyl)-2-(2-(3-methoxyphenyl)prop-1-en-1-yl)-4,4-dimethylpyrrolidine-1-carboxamide



2c, 80% yield, $[\alpha]_D^{27} = -78.05$ (*c* 0.54, CHCl_3).

^1H NMR (400 MHz, CHloroform-d) δ 7.34 – 7.29 (m, 2H), 7.28 – 7.25 (m, 1H), 7.23 – 7.13 (m, 2H), 7.00 (ddd, *J* = 7.6, 2.0, 0.8 Hz, 1H), 6.92 (t, *J* = 2.1 Hz, 1H), 6.85 (ddd, *J* = 8.0, 2.4, 0.8 Hz, 1H), 6.65 (s, 1H), 5.86 (dq, *J* = 8.8, 1.2 Hz, 1H), 4.68 (td, *J* = 9.0, 7.2 Hz, 1H), 3.81 (s, 3H), 3.65 (dd, *J* = 10.4, 1.6 Hz, 1H), 3.21 (d, *J* = 10.4 Hz, 1H), 2.23 (d, *J* = 1.3 Hz, 3H), 2.05 (ddd, *J* = 12.7, 7.2,

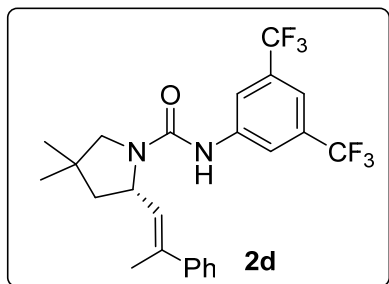
1.6 Hz, 1H), 1.70 – 1.64 (m, 1H), 1.17 (s, 3H), 1.13 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ (one C signal was overlapped) 159.8, 154.3, 143.6, 138.5, 137.7, 131.8, 130.1, 129.6, 120.4, 118.2, 114.8, 113.0, 111.9, 59.1, 55.5, 55.3, 47.7, 36.6, 26.3, 16.2.

HRMS (ESI) *m/z* calcd. for $\text{C}_{23}\text{H}_{28}\text{BrN}_2\text{O}_2$ $[\text{M}+\text{H}]^+$ 443.1329, found 443.1325.

HPLC condition: Chiralcel AD-3, *i*-PrOH/*n*-hexane = 85/15, flow rate 0.8 mL/min. λ = 260 nm, *t*(minor) = 16.6 min, *t*(major) = 22.0 min, 94:6 er.

(S)(E)-N-(3,5-bis(trifluoromethyl)phenyl)-4,4-dimethyl-2-(2-phenylprop-1-en-1-yl)pyrrolidine-1-carboxamide



2d, 76% yield, $[\alpha]_D^{27} = -23.88$ (*c* 0.1, CHCl_3).

^1H NMR (400 MHz, CHloroform-d) δ 7.76 (s, 2H), 7.48 – 7.29 (m, 6H), 7.01 (s, 1H), 5.86 (dd, *J* = 9.0, 1.4 Hz, 1H), 4.73 (td, *J* = 9.0, 7.1 Hz, 1H), 3.66 (dd, *J* = 10.6, 1.6 Hz, 1H), 3.23 (d, *J* = 10.7 Hz, 1H), 2.28 (d, *J* = 1.4 Hz, 3H), 2.09 (ddd, *J* = 12.6, 7.1, 1.6 Hz, 1H), 1.73 (dd, *J* = 12.7, 9.1 Hz, 1H), 1.19 (s, 3H), 1.15 (s,

3H).

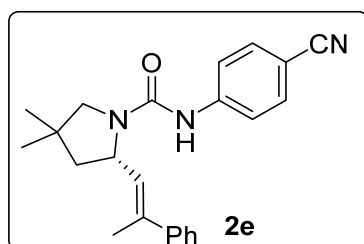
¹³C NMR (151 MHz, CDCl₃) δ 153.8, 141.9, 140.8, 132.1 (q, J_{C-F} = 33.1 Hz), 129.6, 128.7, 128.1, 127.3, 125.7, 123.2 (q, J_{C-F} = 271.0 Hz), 118.5 – 118.4 (m, 1C), 115.8 – 115.6 (m, 1C), 59.2, 55.8, 47.6, 36.7, 26.3(2), 26.2(9), 16.3.

¹⁹F NMR (376 MHz, CDCl₃) δ -63.09.

HRMS (ESI) m/z calcd. for C₂₄H₂₅F₆N₂O [M+H]⁺ 471.1871, found 471.1875.

HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.4 mL/min. λ = 254 nm, t(minor) = 11.9 min, t(major) = 14.8 min, 93:7 er.

(S)(E)-N-(4-cyanophenyl)-4,4-dimethyl-2-(2-phenylprop-1-en-1-yl)pyrrolidine-1-carboxamide



2e, 78% yield, $[\alpha]_D^{27} = -137.1$ (c 0.5, CHCl₃).

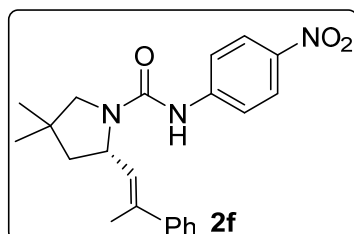
¹H NMR (400 MHz, Chloroform-*d*) δ 7.53 – 7.45 (m, 2H), 7.44 – 7.31 (m, 7H), 6.97 (s, 1H), 5.86 (dd, J = 9.1, 1.4 Hz, 1H), 4.72 (td, J = 9.0, 7.1 Hz, 1H), 3.67 (dd, J = 10.8, 1.6 Hz, 1H), 3.22 (d, J = 10.7 Hz, 1H), 2.27 (d, J = 1.6 Hz, 3H), 2.07 (ddd, J = 12.7, 7.1, 1.6 Hz, 1H), 1.71 (dd, J = 12.7, 9.1 Hz, 1H), 1.17 (s, 3H), 1.14 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 153.7, 143.6, 141.8, 138.4, 133.3, 129.5, 128.7, 128.1, 125.7, 119.3, 118.4, 105.0, 59.1, 55.7, 47.6, 36.7, 26.4, 26.3, 16.2.

HRMS (ESI) m/z calcd. for C₂₃H₂₆N₃O [M+H]⁺ 360.2076, found 360.2071.

HPLC condition: Chiralcel AD-3, *i*-PrOH/*n*-hexane = 85/15, flow rate 0.8 mL/min. λ = 254 nm, t(minor) = 15.3 min, t(major) = 24.6 min, 92.5:7.5 er.

(S)(E)-4,4-dimethyl-N-(4-nitrophenyl)-2-(2-phenylprop-1-en-1-yl)pyrrolidine-1-carboxamide



2f, 62% yield, $[\alpha]_D^{27} = -14.7$ (c 0.1, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*) δ 8.11 (d, J = 8.8 Hz, 2H), 7.44 – 7.32 (m, 7H), 7.11 (s, 1H), 5.88 (d, J = 9.1 Hz, 1H), 4.73 (q, J = 8.5 Hz, 1H), 3.68 (d, J = 10.7 Hz, 1H), 3.24 (d, J = 10.8 Hz, 1H), 2.29 (s, 3H), 2.09 (dd, J = 12.7, 7.0 Hz, 1H), 1.73 (dd, J = 12.7, 9.1 Hz, 2H),

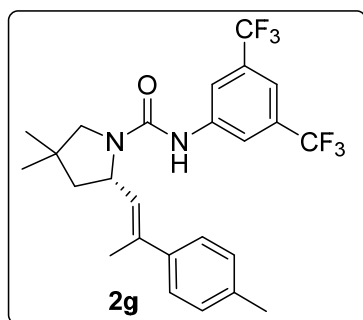
1.19 (s, 3H), 1.15 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 153.5, 145.5, 142.2, 141.7, 138.6, 129.4, 128.7, 128.2, 125.7, 125.2, 117.7, 59.2, 55.8, 47.7, 36.6, 26.3(0), 26.2(9), 16.2.

HRMS (ESI) m/z calcd. for C₂₂H₂₆N₃O₃ [M+H]⁺ 380.1974, found 380.1974.

HPLC condition: Chiralcel AD-3, *i*-PrOH/*n*-hexane = 85/15, flow rate 0.8 mL/min. λ = 254 nm, t(minor) = 16.0 min, t(major) = 25.8 min, 91:9 er.

(S)(E)-N-(3,5-bis(trifluoromethyl)phenyl)-4,4-dimethyl-2-(2-(*p*-tolyl)prop-1-en-1-yl)pyrrolidine-1-carboxamide



2g, 81% yield, $[\alpha]_D^{27} = -78.6$ (*c* 0.6, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*) δ 7.75 (s, 2H), 7.44 (s, 1H), 7.36 – 7.28 (m, 2H), 7.18 (d, *J* = 8.0 Hz, 2H), 7.06 (s, 1H), 5.89 – 5.77 (m, 1H), 4.72 (td, *J* = 9.1, 7.0 Hz, 1H), 3.66 (dd, *J* = 10.8, 1.6 Hz, 1H), 3.23 (d, *J* = 10.7 Hz, 1H), 2.36 (s, 3H), 2.26 (d, *J* = 1.3 Hz, 3H), 2.08 (ddd, *J* = 12.6, 7.1, 1.6 Hz, 1H), 1.72 (dd, *J* = 12.8, 9.1 Hz, 1H), 1.18 (s, 3H), 1.14 (s, 3H).

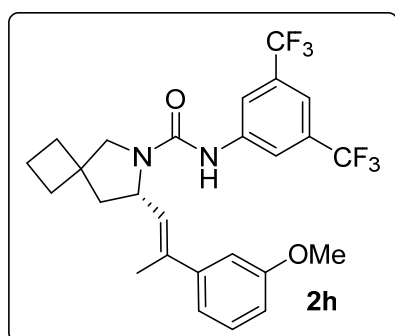
¹³C NMR (101 MHz, CDCl₃) δ 153.9, 140.8, 138.9, 138.5, 138.1, 132.1 (q, *J*_{C-F} = 33.1 Hz), 129.4, 128.7, 125.5, 123.2 (q, *J*_{C-F} = 271.0 Hz), 118.5 – 118.4 (m, 1C), 115.7 – 115.5 (m, 1C), 59.1, 55.8, 47.7, 36.6, 26.3(0), 26.2(8), 21.1, 16.2.

¹⁹F NMR (376 MHz, CDCl₃) δ -63.07.

HRMS (ESI) *m/z* calcd. for C₂₅H₂₇F₆N₂O [M+H]⁺ 485.2028, found 485.2028.

HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.4 mL/min. λ = 254 nm, *t*(minor) = 10.4 min, *t*(major) = 13.0 min, 92:8 er.

(*S*)(*E*)-N-(3,5-bis(trifluoromethyl)phenyl)-7-(2-(3-methoxyphenyl)prop-1-en-1-yl)-6-azaspiro[3.4]octane-6-carboxamide



2h, 80% yield, $[\alpha]_D^{27} = -80.6$ (*c* 0.8, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*) δ 7.76 (s, 2H), 7.45 (s, 1H), 7.29 (t, *J* = 8.0 Hz, 1H), 6.99 (ddd, *J* = 7.8, 1.8, 0.9 Hz, 1H), 6.94 (s, 1H), 6.93 – 6.92 (m, 1H), 6.86 (ddd, *J* = 8.2, 2.6, 0.9 Hz, 1H), 5.84 (dd, *J* = 9.1, 1.4 Hz, 1H), 4.64 (dt, *J* = 9.1, 7.2 Hz, 1H), 3.82 (s, 3H), 3.81 (d, *J* = 10.7 Hz, 1H), 3.50 (d, *J* = 10.7 Hz, 1H), 2.34 (ddd, *J* = 12.6, 7.2, 1.1 Hz, 1H), 2.25 (d, *J* = 1.3 Hz, 3H), 2.16 – 1.87 (m, 7H).

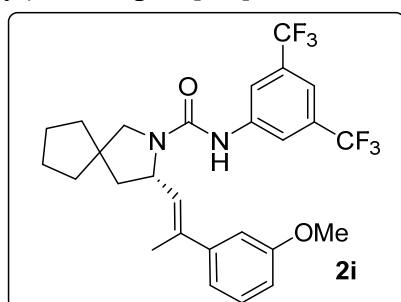
¹³C NMR (101 MHz, CDCl₃) δ 159.8, 153.8, 143.4, 140.8, 138.2, 132.1 (q, *J*_{C-F} = 33.1 Hz), 129.7, 129.5, 123.2 (q, *J*_{C-F} = 271.0 Hz), 118.5 – 118.4 (m, 1C), 118.1, 115.7 – 115.6 (m, 1C), 113.2, 111.8, 57.9, 55.6, 55.3, 45.8, 43.4, 32.4, 30.6, 16.3, 16.1.

¹⁹F NMR (376 MHz, CDCl₃) δ -63.08.

HRMS (ESI) *m/z* calcd. for C₂₆H₂₇F₆N₂O₂ [M+H]⁺ 513.1977, found 513.1980.

HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 0.5 mL/min. λ = 254 nm, *t*(minor) = 13.0 min, *t*(major) = 16.0 min, 94:6 er.

(*S*)(*E*)-N-(3,5-bis(trifluoromethyl)phenyl)-3-(2-(3-methoxyphenyl)prop-1-en-1-yl)-2-azaspiro[4.4]nonane-2-carboxamide



2i, 84% yield, $[\alpha]_D^{27} = -60.3$ (*c* 0.8, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*) δ 7.77 (s, 2H), 7.45 (s, 1H), 7.29 (t, *J* = 8.0 Hz, 1H), 7.04 – 6.97 (m, 2H), 6.93 (t, *J* = 2.1 Hz, 1H), 6.87 (ddd, *J* = 8.2, 2.5, 0.9 Hz, 1H), 5.88 (dd, *J* = 9.1, 1.4 Hz, 1H), 4.66 (td, *J* = 8.6, 7.1 Hz, 1H), 3.82 (s, 3H), 3.69 (d, *J* = 10.6 Hz, 1H), 3.33 (d, *J* = 10.6 Hz, 1H), 2.26 (d, *J* = 1.4

Hz, 3H), 2.19 (ddd, $J = 12.5, 7.1, 1.4$ Hz, 1H), 1.86 (dd, $J = 12.5, 8.4$ Hz, 1H), 1.77 – 1.49 (m, 8H).

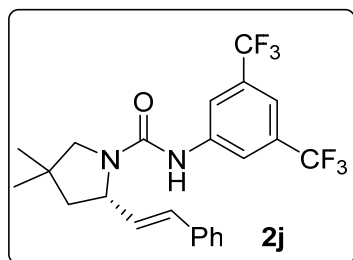
^{13}C NMR (101 MHz, CDCl_3) δ 159.8, 153.8, 143.4, 140.8, 138.4, 132.1 (q, $J_{\text{C-F}} = 33.1$ Hz), 129.8, 129.7, 123.2 (q, $J_{\text{C-F}} = 271.0$ Hz), 118.5 – 118.4 (m, 1C), 118.1, 115.7 – 115.6 (m, 1C), 113.2, 111.7, 57.9, 56.2, 55.3, 47.8, 45.9, 37.5, 36.3, 24.9, 24.8, 16.3.

^{19}F NMR (376 MHz, CDCl_3) δ -63.08.

HRMS (ESI) m/z calcd. for $\text{C}_{27}\text{H}_{29}\text{F}_6\text{N}_2\text{O}_2$ $[\text{M}+\text{H}]^+$ 527.2133, found 527.2136.

HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 0.5 mL/min. $\lambda = 254$ nm, $t(\text{minor}) = 10.9$ min, $t(\text{major}) = 15.1$ min, 90:10 er.

(*S*)(*E*)-*N*-(3,5-bis(trifluoromethyl)phenyl)-4,4-dimethyl-2-styrylpyrrolidine-1-carboxamide



2j, 54% yield, $[\alpha]_D^{27} = -50.9$ (*c* 0.5, CHCl_3).

^1H NMR (400 MHz, Chloroform-*d*) δ 7.79 (s, 2H), 7.48 – 7.42 (m, 3H), 7.42 – 7.37 (m, 2H), 7.37 – 7.31 (m, 1H), 7.11 (s, 1H), 6.83 (d, $J = 16.0$ Hz, 1H), 6.27 (dd, $J = 16.0, 8.4$ Hz, 1H), 4.52 (q, $J = 8.0$ Hz, 1H), 3.71 (d, $J = 10.6$ Hz, 1H), 3.27 (d, $J = 10.6$ Hz, 1H), 2.12 (ddd, $J = 12.8, 7.2, 1.6$ Hz, 1H), 1.82 (dd, $J = 12.8, 9.6$ Hz, 1H), 1.22 (s,

3H), 1.15 (s, 3H).

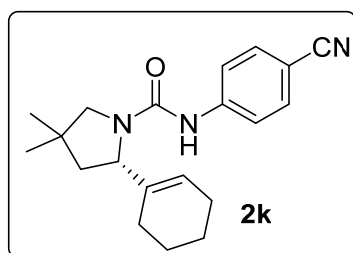
^{13}C NMR (151 MHz, CDCl_3) δ 154.0, 140.6, 135.3, 132.6, 132.4, 131.9 (q, $J_{\text{C-F}} = 33.1$ Hz), 129.0, 128.7, 126.5, 123.2 (q, $J_{\text{C-F}} = 271.0$ Hz), 118.6 – 118.5 (m, 1C), 115.8 – 115.7 (m, 1C), 60.1, 59.4, 48.2, 36.8, 26.3, 26.2.

^{19}F NMR (376 MHz, CDCl_3) δ -63.09.

HRMS (ESI) m/z calcd. for $\text{C}_{23}\text{H}_{23}\text{F}_6\text{N}_2\text{O}$ $[\text{M}+\text{H}]^+$ 457.1709, found 457.1708.

HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.6 mL/min. $\lambda = 254$ nm, $t(\text{major}) = 13.3$ min, $t(\text{minor}) = 20.8$ min, 90:10 er.

(*S*)-*N*-(4-cyanophenyl)-2-(cyclohex-1-en-1-yl)-4,4-dimethylpyrrolidine-1-carboxamide



2k, 25% yield (40% conversion), $[\alpha]_D^{27} = 14.4$ (*c* 0.14, CHCl_3). **^1H NMR (400 MHz, Chloroform-*d*)** δ 7.55 (d, $J = 8.8$ Hz, 2H), 7.45 (d, $J = 8.8$ Hz, 2H), 7.25 (s, 1H), 6.02 (t, $J = 1.7$ Hz, 1H), 4.27 (dd, $J = 9.9, 7.1$ Hz, 1H), 3.76 (dd, $J = 10.9, 1.8$ Hz, 1H), 3.08 (d, $J = 10.9$ Hz, 1H), 2.20 – 2.17 (m, 2H), 1.99 – 1.96 (m, 2H), 1.90 – 1.85 (m, 1H), 1.82 – 1.75 (m, 1H), 1.73 – 1.58 (m, 4H), 1.16 (s,

3H), 1.09 (s, 3H).

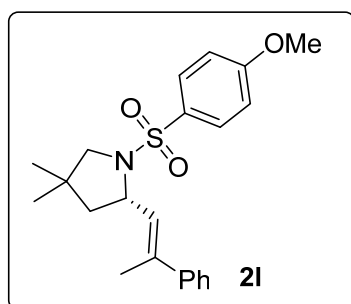
^{13}C NMR (101 MHz, CDCl_3) δ 153.8, 143.7, 139.5, 133.3, 126.4, 119.3, 118.2, 104.9, 63.9, 59.5, 46.1, 36.2, 26.1, 25.8, 25.3, 22.4(2), 22.4(1), 22.3.

HRMS (ESI) m/z calcd. for $\text{C}_{20}\text{H}_{26}\text{N}_3\text{O}$ $[\text{M}+\text{H}]^+$ 324.2076, found 324.2074.

HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 97/3, flow rate 0.3 mL/min. $\lambda = 254$ nm, $t(\text{major}) = 60.8$ min, $t(\text{minor}) = 69.5$ min, 76:24 er.

(*S*)(*E*)-1-((4-methoxyphenyl)sulfonyl)-4,4-dimethyl-2-(2-phenylprop-1-en-1-

yl)pyrrolidine



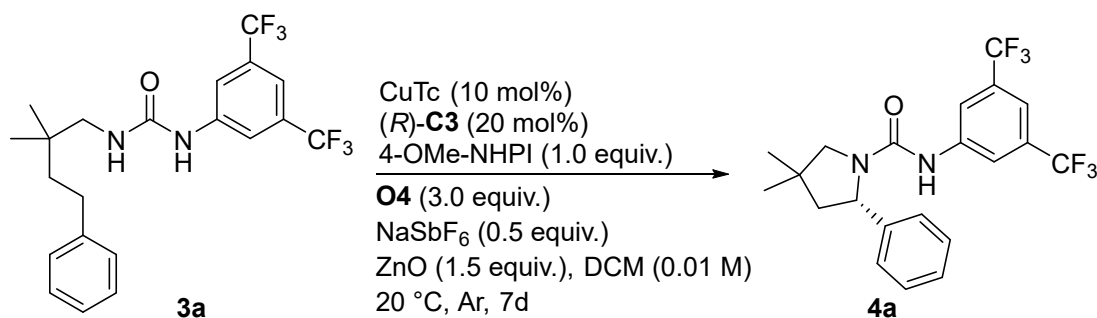
2l, 35% yield (75% conversion), $[\alpha]_D^{27} = -30.9$ (*c* 0.2, CHCl₃). ¹H NMR (400 MHz, Chloroform-*d*) δ 7.75 (d, *J* = 8.3 Hz, 2H), 7.44 – 7.10 (m, 5H), 6.91 (d, *J* = 8.4 Hz, 2H), 5.66 (d, *J* = 8.6 Hz, 1H), 4.54 (q, *J* = 8.3 Hz, 1H), 3.83 (s, 3H), 3.32 (d, *J* = 10.0 Hz, 1H), 3.23 (d, *J* = 10.0 Hz, 1H), 2.10 (s, 3H), 1.91 (dd, *J* = 12.7, 7.6 Hz, 1H), 1.57 (dd, *J* = 12.6, 8.7 Hz, 1H), 1.12 (s, 3H), 0.92 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 162.6, 142.9, 134.9, 131.2, 129.8, 129.5, 128.1, 127.0, 125.8, 113.9, 61.1, 58.0, 55.5, 47.7, 37.6, 26.6, 26.2, 16.0.

HRMS (ESI) *m/z* calcd. for C₂₂H₂₈NO₃S [M+H]⁺ 386.1790, found 386.1786.

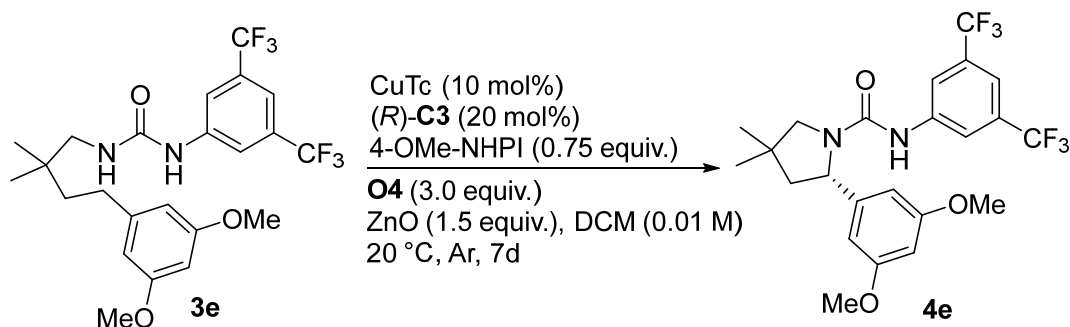
HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 0.5 mL/min. λ = 254 nm, *t*(major) = 16.0 min, *t*(minor) = 18.4 min, 87:13 er.

General procedure for enantioselective radical benzylic C–H amination:



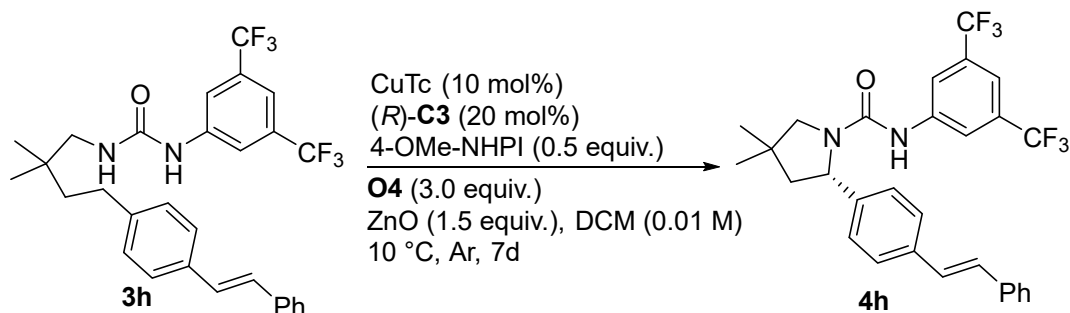
To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **3a** (43.2 mg, 0.1 mmol), CuTc (1.9 mg 10 mol%), (*R*)-**C3** (16 mg, 20 mol%), 4-OMe-NHPI (19 mg, 0.1 mmol), ZnO (12 mg, 0.15 mmol) and NaSbF₆ (13 mg, 0.05 mmol). The tube was evacuated and backfilled with argon for three times, the **O4** (72 μ L, 0.3 mmol) and dry DCM (10.0 mL) was added *via* syringe. The tube was stirred at 20 °C and (*R*)-**C3** (4 mg, 5 mol%) was added after 12 h and 24 h, respectively. After 7 days, the reaction stopped and the solvent was removed under reduced pressure, and the residue was purified by flash chromatography (petroleum ether/ ethyl acetate =20/1-10/1) to give the corresponding product **4a** in 38% isolated yield with 55% conversion (67% yield based on recovered starting material).

Procedure for enantioselective radical benzylic C–H amination of **3e**.



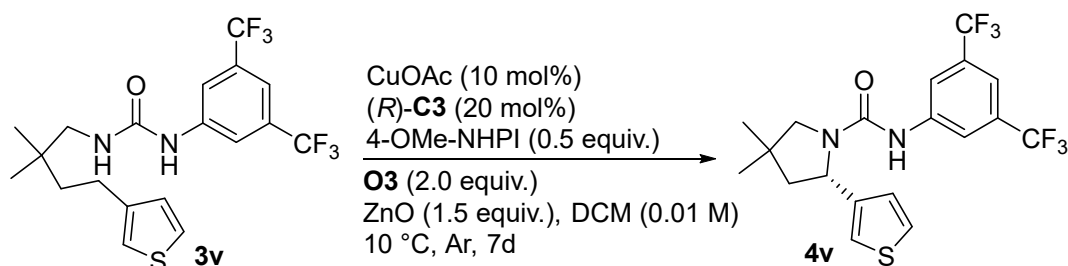
To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **3e** (49.3 mg, 0.1 mmol), CuTc (1.9 mg 10 mol%), (*R*)-**C3** (16 mg, 20 mol%), 4-OMe-NHPI (14.3 mg, 0.075 mmol), and ZnO (12 mg, 0.15 mmol). The tube was evacuated and backfilled with argon for three times, the **O4** (72 μ L, 0.3 mmol) and dry DCM (10.0 mL) was added *via* syringe. The tube was stirred at 10 °C and (*R*)-**C3** (4 mg, 5 mol%) was added after 12 h and 24 h, respectively. After 7 days, the reaction stopped and the solvent was removed under reduced pressure, and the residue was purified by flash chromatography (petroleum ether/ ethyl acetate =20/1-10/1) to give the corresponding product **4e** in 40% isolated yield with 53% conversion (76% yield based on recovered starting material).

Procedure for enantioselective radical benzylic C–H amination of **3h**.



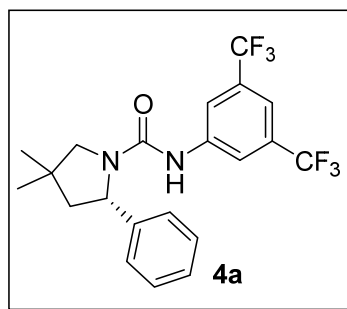
To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **3h** (53.5 mg, 0.1 mmol), CuTc (1.9 mg 10 mol%), (*R*)-**C3** (16 mg, 20 mol%), 4-OMe-NHPI (9.5 mg, 0.05 mmol), and ZnO (12 mg, 0.15 mmol). The tube was evacuated and backfilled with argon for three times, the **O4** (72 μ L, 0.3 mmol) and dry DCM (10.0 mL) was added *via* syringe. The tube was stirred at 10 °C and (*R*)-**C3** (4 mg, 5 mol%) was added after 12 h and 24 h, respectively. After 7 days, the reaction stopped and the solvent was removed under reduced pressure, and the residue was purified by flash chromatography (petroleum ether/ ethyl acetate =20/1-10/1) to give the corresponding product **4h** in 44% isolated yield with 60% conversion (73% yield based on recovered starting material).

Procedure for enantioselective radical benzylic C–H amination of **3v**.



To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **3v** (43.8 mg, 0.1 mmol), CuOAc (1.24 mg 10 mol%), (*R*)-**C3** (16 mg, 20 mol%), 4-OMe-NHPI (9.5 mg, 0.05 mmol), and ZnO (12 mg, 0.15 mmol). The tube was evacuated and backfilled with argon for three times, the **O3** (48 μ L, 0.2 mmol) and dry DCM (10.0 mL) was added *via* syringe. The tube was stirred at 10 °C and (*R*)-**C3** (4 mg, 5 mol%) was added after 12 h and 24 h, respectively. After 7 days, the reaction stopped and the solvent was removed under reduced pressure, and the residue was purified by flash chromatography (petroleum ether/ ethyl acetate =20/1-10/1) to give the corresponding product **4v** in 51% isolated yield with 71% conversion (72% yield based on recovered starting material).

(*S*)-*N*-(3,5-bis(trifluoromethyl)phenyl)-4,4-dimethyl-2-phenylpyrrolidine-1-carboxamide



4a, 69% yield based on recovered starting material, $[\alpha]_D^{27} = -63.1$ (c 0.5, CHCl₃).

¹H NMR (400 MHz, CDCl₃): δ 7.53 (s, 2H), 7.47-7.37 (m, 6H), 6.29 (s, 1H), 4.86 (dd, *J* = 9.2, 7.2 Hz, 1H), 3.85 (d, *J* = 10.4 Hz, 1H), 3.40 (d, *J* = 10.8 Hz, 1H), 2.28 (ddd, *J* = 12.7, 7.0, 1.5 Hz, 1H), 1.84 (dd, *J* = 12.4, 10.0 Hz, 1H), 1.19 (s, 3H), 1.18 (s, 3H).

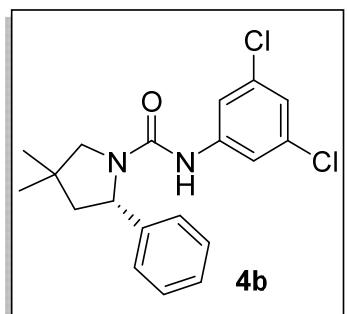
¹³C NMR (101 MHz, CDCl₃): δ 153.6, 142.1, 140.5, 131.9 (q, *J*_{C-F} = 33.3 Hz), 129.7, 128.6, 125.9, 123.1 (q, *J*_{C-F} = 271.0 Hz), 118.7 – 118.6 (m, 1C), 116.0 – 115.8 (m, 1C), 61.7, 60.3, 52.0, 37.3, 26.1, 25.9.

¹⁹F NMR (376 MHz, CDCl₃): δ -63.09.

HRMS (ESI) *m/z* calcd. for C₂₁H₂₁F₆N₂O [*M*+H]⁺ 431.1553, found 431.1549.

HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.6 mL/min. λ = 254 nm, *t*(major) = 10.6 min, *t*(minor) = 16.4 min, 95:5 er.

(*S*)-*N*-(3,5-dichlorophenyl)-4,4-dimethyl-2-phenylpyrrolidine-1-carboxamide



4b, 84% yield based on recovered starting material, $[\alpha]_D^{27} = -31.7$ (c 0.1, CHCl₃).

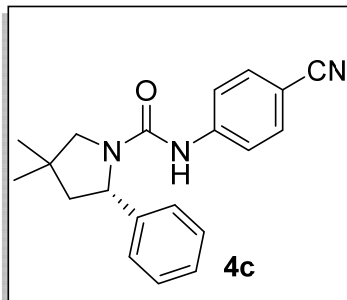
¹H NMR (400 MHz, Chloroform-*d*): δ 7.45 – 7.41 (m, 2H), 7.38 – 7.34 (m, 3H), 7.02 (s, 2H), 6.90 (t, *J* = 2.0 Hz, 1H), 6.04 (s, 1H), 4.80 (dd, *J* = 9.6, 6.8 Hz, 1H), 3.84 (d, *J* = 10.8 Hz, 1H), 3.37 (d, *J* = 10.8 Hz, 1H), 2.25 (ddd, *J* = 12.9, 7.0, 1.7 Hz, 1H), 1.81 (dd, *J* = 12.8, 9.6 Hz, 1H), 1.17 (s, 3H), 1.15 (s, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 153.6, 142.2, 140.9, 134.8, 129.7, 128.5, 125.9, 122.5, 117.1, 61.7, 60.2, 52.1, 37.3, 26.1, 25.9.

HRMS (ESI) m/z calcd. for C₁₉H₂₁Cl₂N₂O [M+H]⁺ 363.1025, found 363.1022.

HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 90/10, flow rate 1.0 mL/min. λ = 254 nm, t(major) = 6.8 min, t(minor) = 12.7 min, 91:9 er.

(S)-N-(4-cyanophenyl)-4,4-dimethyl-2-phenylpyrrolidine-1-carboxamide



4c, 72% yield based on recovered starting material, [α]_D²⁷ = −77.8 (*c* 0.5, CHCl₃).

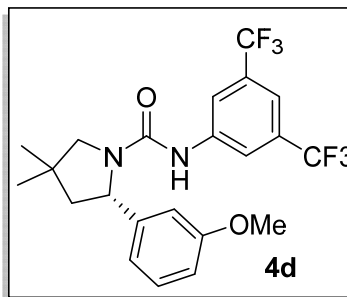
¹H NMR (400 MHz, Chloroform-*d*): δ 7.46 – 7.41 (m, 4H), 7.38 – 7.34 (m, 3H), 7.15 (d, *J* = 8.4 Hz, 2H), 6.25 (s, 1H), 4.83 (dd, *J* = 9.8, 7.0 Hz, 1H), 3.87 (d, *J* = 10.8 Hz, 1H), 3.38 (d, *J* = 10.8 Hz, 1H), 2.26 (ddd, *J* = 12.8, 7.0, 1.8 Hz, 1H), 1.83 (dd, *J* = 12.8, 10.0 Hz, 1H), 1.18 (s, 3H), 1.17 (s, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 153.5, 143.2, 142.1, 133.1, 129.7, 128.6, 126.0, 119.2, 118.5, 105.1, 61.8, 60.2, 52.1, 37.2, 26.1, 25.9.

HRMS (ESI) m/z calcd. for C₂₀H₂₂N₃O [M+H]⁺ 320.1757, found 320.1756.

HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 85/15, flow rate 0.6 mL/min. λ = 254 nm, t(major) = 16.1 min, t(minor) = 20.8 min, 92:8 er.

(S)-N-(3,5-bis(trifluoromethyl)phenyl)-2-(3-methoxyphenyl)-4,4-dimethylpyrrolidine-1-carboxamide



4d, 61% yield based on recovered starting material, [α]_D²⁷ = −44.5 (*c* 0.6, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*): δ 7.57 (s, 2H), 7.42 (s, 1H), 7.39 – 7.34 (m, 1H), 6.97 – 6.95 (m, 1H), 6.91 – 6.88 (m, 2H), 6.36 (s, 1H), 4.82 (dd, *J* = 9.6, 7.2 Hz, 1H), 3.85 – 3.82 (m, 4H), 3.38 (d, *J* = 10.4 Hz, 1H), 2.27 (ddd, *J* = 12.8, 7.0, 1.7 Hz, 1H), 1.84 (dd, *J* = 12.8, 9.6 Hz, 1H), 1.18 (s, 3H), 1.16 (s, 3H).

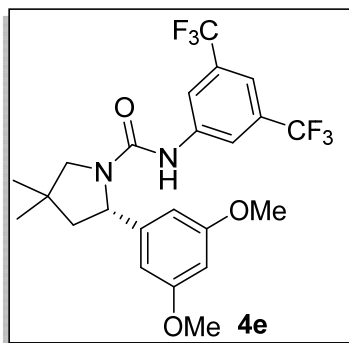
¹³C NMR (101 MHz, CDCl₃): δ 160.6, 153.6, 143.8, 140.5, 131.9 (q, *J*_{C-F} = 33.1 Hz), 130.8, 123.2 (q, *J*_{C-F} = 271.0 Hz), 118.7 – 118.6 (m, 1C), 118.0, 115.9 – 115.8 (m, 1C), 113.5, 111.9, 61.6, 60.2, 55.3, 51.8, 37.3, 26.1, 25.9.

¹⁹F NMR (376 MHz, CDCl₃): δ −63.07.

HRMS (ESI) m/z calcd. for C₂₂H₂₃F₆N₂O₂ [M+H]⁺ 461.1658, found 461.1655.

HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.6 mL/min. λ = 254 nm, t(major) = 15.1 min, t(minor) = 29.9 min, 95:5 er.

(S)-N-(3,5-bis(trifluoromethyl)phenyl)-2-(3,5-dimethoxyphenyl)-4,4-dimethylpyrrolidine-1-carboxamide



4e, 76% yield based on recovered starting material, $[\alpha]_D^{27} = -18.4$ (c 0.2, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*): δ 7.61 (s, 2H), 7.42 (s, 1H), 6.50 (d, *J* = 2.0 Hz, 2H), 6.47 (s, 1H), 6.43 (t, *J* = 2.0 Hz, 1H), 4.78 (dd, *J* = 9.6, 7.2 Hz, 1H), 3.85 – 3.80 (m, 7H), 3.36 (d, *J* = 10.4 Hz, 1H), 2.25 (ddd, *J* = 12.8, 7.0, 1.7 Hz, 1H), 1.84 (dd, *J* = 12.8, 10.0 Hz, 1H), 1.18 (s, 3H), 1.15 (s, 3H).

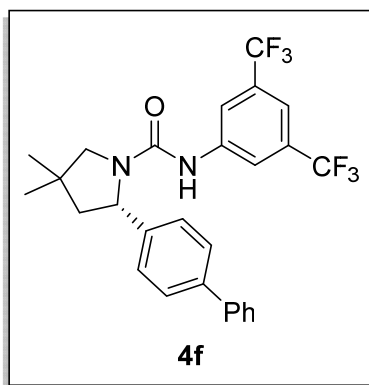
¹³C NMR (101 MHz, CDCl₃): δ 161.9, 153.8, 144.7, 140.6, 132.0 (q, *J*_{C-F} = 33.1 Hz), 123.2 (q, *J*_{C-F} = 271.0 Hz), 118.7 – 118.6 (m, 1C), 115.9 – 115.7 (m, 1C), 103.9, 99.5, 61.8, 60.2, 55.4, 51.5, 37.3, 26.1, 25.8.

¹⁹F NMR (376 MHz, CDCl₃): δ -63.07.

HRMS (ESI) *m/z* calcd. for C₂₃H₂₅F₆N₂O₃ [M+H]⁺ 491.1764, found 491.1762.

HPLC condition: Chiralcel AD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.8 mL/min. λ = 254 nm, *t*(major) = 7.3 min, *t*(minor) = 13.3 min, 94.5:5.5 er.

(*S*)-2-([1,1'-biphenyl]-4-yl)-*N*-(3,5-bis(trifluoromethyl)phenyl)-4,4-dimethylpyrrolidine-1-carboxamide



4f, 64% yield based on recovered starting material, $[\alpha]_D^{27} = -74.5$ (c 0.8, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*): δ 7.66 – 7.57 (m, 6H), 7.47 – 7.41 (m, 5H), 7.39 – 7.35 (m, 1H), 6.40 (s, 1H), 4.93 (t, *J* = 8.2 Hz, 1H), 3.84 (d, *J* = 10.4 Hz, 1H), 3.44 (d, *J* = 10.4 Hz, 1H), 2.31 (ddd, *J* = 12.8, 7.0, 1.7 Hz, 1H), 1.87 (dd, *J* = 12.8, 9.6 Hz, 1H), 1.20 (s, 3H), 1.19 (s, 3H).

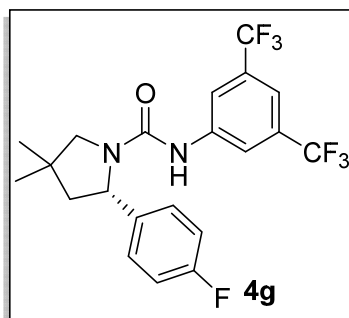
¹³C NMR (101 MHz, CDCl₃): δ 153.6, 141.5, 141.1, 140.5, 140.2, 132.0 (q, *J*_{C-F} = 33.1 Hz), 128.9, 128.3, 127.6, 127.1, 126.3, 123.1 (q, *J*_{C-F} = 271.3 Hz), 118.7 – 118.6 (m, 1C), 115.9 – 115.8 (m, 1C), 61.4, 60.3, 51.8, 37.5, 26.1, 25.9.

¹⁹F NMR (376 MHz, CDCl₃): δ -63.04.

HRMS (ESI) *m/z* calcd. for C₂₇H₂₅F₆N₂O [M+H]⁺ 507.1866, found 507.1866.

HPLC condition: Chiralcel AD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.5 mL/min. λ = 254 nm, *t*(major) = 10.5 min, *t*(minor) = 12.8 min, 94:6 er.

(*S*)-*N*-(3,5-bis(trifluoromethyl)phenyl)-2-(4-fluorophenyl)-4,4-dimethylpyrrolidine-1-carboxamide



4g, 80% yield based on recovered starting material, $[\alpha]_D^{27} = -61.1$ (*c* 0.6, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*): δ 7.64 (s, 2H), 7.44 (s, 1H), 7.35 – 7.31 (m, 2H), 7.11 (t, *J* = 8.6 Hz, 2H), 6.32 (s, 1H), 4.90 (dd, *J* = 8.8, 7.2 Hz, 1H), 3.78 (d, *J* = 10.8 Hz, 1H), 3.40 (d, *J* = 10.6 Hz, 1H), 2.26 (ddd, *J* = 12.8, 7.0, 1.7 Hz, 1H), 1.77 (dd, *J* = 12.8, 9.6 Hz, 2H), 1.18 (s, 3H), 1.17 (s, 3H).

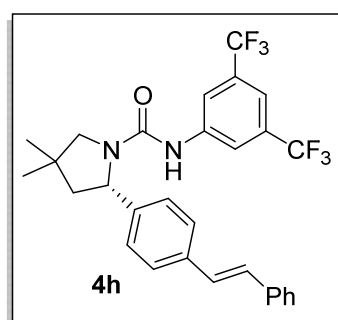
¹³C NMR (101 MHz, CDCl₃): δ 162.4 (q, *J*_{C-F} = 246.1 Hz), 153.5, 140.3, 138.0, 132.0 (q, *J*_{C-F} = 33.1 Hz), 127.4 (d, *J*_{C-F} = 8.1 Hz), 123.2 (q, *J*_{C-F} = 271.2 Hz), 118.7 – 118.6 (m, 1C), 116.5 (d, *J*_{C-F} = 21.5 Hz), 116.0 – 115.9 (m, 1C), 61.0, 60.3, 51.8, 37.6, 26.1, 25.9.

¹⁹F NMR (565 MHz, DMSO-*d*₆, 80 °C) δ -61.80, -116.86.

HRMS (ESI) *m/z* calcd. for C₂₁H₂₀F₇N₂O [*M*+*H*]⁺ 449.1458, found 449.1454.

HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 98/2, flow rate 0.4 mL/min. λ = 254 nm, *t*(major) = 26.9 min, *t*(minor) = 31.6 min, 94:6 er.

(*S*)-(*E*)-*N*-(3,5-bis(trifluoromethyl)phenyl)-4,4-dimethyl-2-(4-styrylphenyl)pyrrolidine-1-carboxamide



4h, 73% yield based on recovered starting material, $[\alpha]_D^{27} = -85$ (*c* 1.4, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*): δ 7.60 – 7.56 (m, 4H), 7.53 – 7.51 (m, 2H), 7.41 (s, 1H), 7.39 – 7.34 (m, 4H), 7.30 – 7.27 (m, 1H), 7.11 (d, *J* = 2.0 Hz, 2H), 6.36 (s, 1H), 4.91 – 4.86 (m, 1H), 3.84 (d, *J* = 10.4 Hz, 1H), 3.42 (d, *J* = 10.5 Hz, 1H), 2.28 (ddd, *J* = 12.8, 7.1, 1.6 Hz, 1H), 1.83 (dd, *J* = 12.8, 9.7 Hz, 1H), 1.19 (s, 3H), 1.18 (s, 3H).

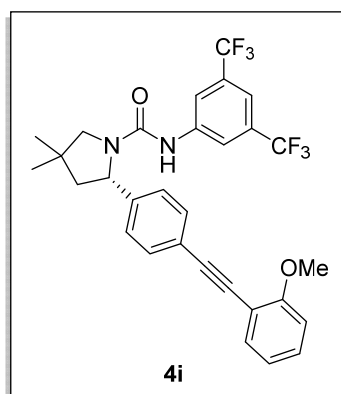
¹³C NMR (101 MHz, CDCl₃): δ 153.6, 140.4, 137.0, 135.2, 132.0 (q, *J*_{C-F} = 32.9 Hz), 129.6, 128.7, 127.9, 127.7, 127.6, 126.6, 126.2, 124.3, 123.1 (q, *J*_{C-F} = 271.1 Hz), 119.1 - 118.7 (m, 1C), 116.0 - 115.8 (m, 1C), 61.5, 60.3, 51.8, 37.4, 26.1, 25.9.

¹⁹F NMR (376 MHz, CDCl₃): δ -63.04.

HRMS (ESI) *m/z* calcd. for C₂₉H₂₇F₆N₂O [*M*+*H*]⁺ 533.2028, found 533.2028.

HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.5 mL/min. λ = 254 nm, *t*(major) = 15.5 min, *t*(minor) = 18.7 min, 96:4 er.

(*S*)-*N*-(3,5-bis(trifluoromethyl)phenyl)-2-(4-((2-methoxyphenyl)ethynyl)phenyl)-4,4-dimethylpyrrolidine-1-carboxamide



4i, 70% yield based on recovered starting material, $[\alpha]_D^{27} = -43$ (c 1.0, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*): δ 7.65 (s, 2H), 7.60 (d, *J* = 8.0 Hz, 2H), 7.49 (dd, *J* = 7.6, 1.8 Hz, 1H), 7.43 (s, 1H), 7.28 (d, *J* = 8.1 Hz, 3H), 6.96 – 6.90 (m, 2H), 6.32 (s, 1H), 4.92 – 4.88 (m, 1H), 3.91 (s, 3H), 3.83 – 3.81 (m, 1H), 3.41 (d, *J* = 10.4 Hz, 1H), 2.28 (ddd, *J* = 12.8, 7.2, 1.7 Hz, 1H), 1.82 (dd, *J* = 12.8, 9.6 Hz, 1H), 1.20 (s, 3H), 1.19 (s, 3H).

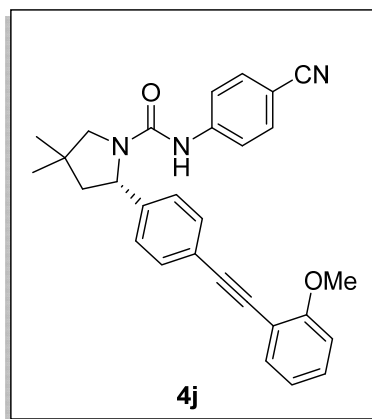
¹³C NMR (101 MHz, CDCl₃) δ 160.0, 153.6, 142.1, 140.4, 133.6, 132.8, 132.0 (q, *J*_{C-F} = 33.1 Hz), 130.0, 125.7, 123.8, 123.1 (q, *J*_{C-F} = 271.1 Hz), 120.5, 119.1 – 118.7 (m, 1C), 116.0 – 115.9 (m, 1C), 112.2, 110.8, 92.6, 86.6, 61.5, 60.3, 55.8, 51.7, 37.6, 26.1, 25.9.

¹⁹F NMR (376 MHz, CDCl₃) δ -63.0.

HRMS (ESI) *m/z* calcd. for C₃₀H₂₇F₆N₂O₂ [*M*+*H*]⁺ 561.1977, found 561.1978.

HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.5 mL/min. λ = 254 nm, *t*(major) = 17.7 min, *t*(minor) = 22.0 min, 96:4 er.

(*S*)-*N*-(4-cyanophenyl)-2-(4-((2-methoxyphenyl)ethynyl)phenyl)-4,4-dimethylpyrrolidine-1-carboxamide



4j, 74% yield based on recovered starting material, $[\alpha]_D^{27} = -154.6$ (c 0.6, CHCl₃).

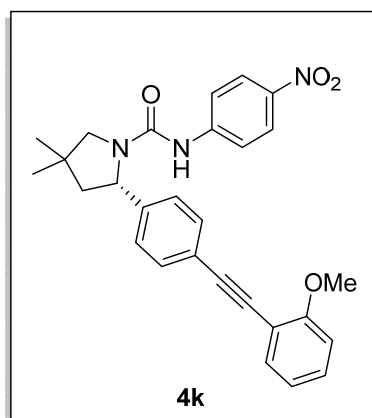
¹H NMR (400 MHz, Chloroform-*d*): δ 7.61 (d, *J* = 8.0 Hz, 2H), 7.50 (dd, *J* = 7.6, 1.6 Hz, 1H), 7.45 (d, *J* = 8.8 Hz, 2H), 7.35 – 7.31 (m, 3H), 7.20 (d, *J* = 8.4 Hz, 2H), 6.97 – 6.91 (m, 2H), 6.23 (s, 1H), 4.85 (dd, *J* = 9.6, 7.0 Hz, 1H), 3.92 (s, 3H), 3.86 (d, *J* = 10.8 Hz, 1H), 3.39 (d, *J* = 10.4 Hz, 1H), 2.27 (ddd, *J* = 12.9, 7.0, 1.6 Hz, 1H), 1.81 (dd, *J* = 12.8, 9.6 Hz, 1H), 1.18 (s, 3H), 1.17 (s, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 156.0, 153.4, 143.1, 142.0, 133.6, 133.1, 132.9, 130.1, 125.9, 123.9, 120.6, 119.2, 118.6, 112.0, 110.7, 105.3, 92.6, 86.8, 61.6, 60.2, 55.9, 51.9, 37.4, 26.1, 25.9.

HRMS (ESI) *m/z* calcd. for C₂₉H₂₈N₃O₂ [*M*+*H*]⁺ 450.2176, found 450.2175.

HPLC condition: Chiralcel AD-H, *i*-PrOH/*n*-hexane = 75/25, flow rate 1.0 mL/min. λ = 230 nm, *t*(minor) = 13.7 min, *t*(major) = 24.9 min, 97:3 er.

(*S*)-2-(4-((2-methoxyphenyl)ethynyl)phenyl)-4,4-dimethyl-*N*-(4-nitrophenyl)pyrrolidine-1-carboxamide



4k, 69% yield based on recovered starting material, $[\alpha]_D^{27} = -180$ (*c* 1.3, CHCl₃).

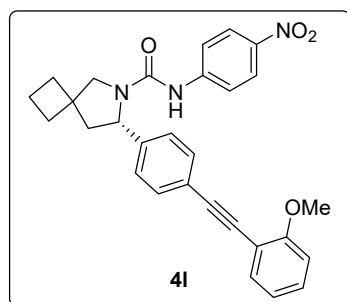
¹H NMR (400 MHz, Chloroform-*d*): δ 8.09 – 8.05 (m, 2H), 7.63 – 7.61 (m, 2H), 7.50 (dd, *J* = 7.6, 1.7 Hz, 1H), 7.36 – 7.31 (m, 3H), 7.24 (d, *J* = 10.5 Hz, 2H), 6.97 – 6.91 (m, 2H), 6.38 (s, 1H), 4.87 (dd, *J* = 9.6, 7.0 Hz, 1H), 3.92 (s, 3H), 3.86 (d, *J* = 10.8 Hz, 1H), 3.40 (d, *J* = 10.8 Hz, 1H), 2.28 (ddd, *J* = 12.8, 7.1, 1.7 Hz, 1H), 1.82 (dd, *J* = 12.8, 9.7 Hz, 1H), 1.19 (s, 3H), 1.17 (s, 3H).

¹³C NMR (101 MHz, CDCl₃): δ 160.0, 153.2, 145.0, 142.4, 141.9, 133.6, 132.9, 130.1, 125.9, 125.1, 124.0, 120.6, 117.9, 112.0, 110.7, 92.5, 86.8, 61.6, 60.3, 55.9, 51.9, 37.4, 26.1, 25.9.

HRMS (ESI) *m/z* calcd. for C₂₈H₂₈N₃O₄ [M+H]⁺ 470.2074, found 470.2075.

HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 85/15, flow rate 1.0 mL/min. λ = 280 nm, *t*(minor) = 26.6 min, *t*(major) = 31.7 min, 96.5:3.5 er.

(*S*)-7-(4-((2-methoxyphenyl)ethynyl)phenyl)-N-(4-nitrophenyl)-6-azaspiro[3.4]octane-6-carboxamide



4l, 68% yield based on recovered starting material, $[\alpha]_D^{27} = -123.3$ (*c* 0.4, CHCl₃).

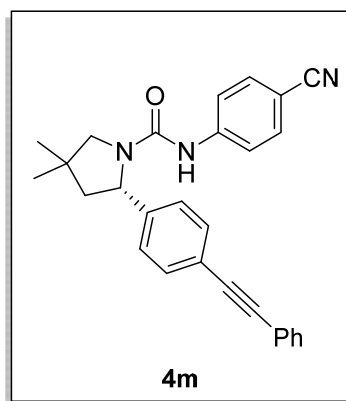
¹H NMR (400 MHz, Chloroform-*d*) δ 8.08 (d, *J* = 9.2 Hz, 2H), 7.63 (d, *J* = 8.3 Hz, 2H), 7.52 (dd, *J* = 7.6, 1.7 Hz, 1H), 7.40 – 7.32 (m, 3H), 7.28 (d, *J* = 5.6 Hz, 2H), 7.02 – 6.89 (m, 2H), 6.40 (s, 1H), 4.84 (t, *J* = 7.3 Hz, 1H), 4.01 (d, *J* = 10.8 Hz, 1H), 3.94 (s, 3H), 3.68 (d, *J* = 10.7 Hz, 1H), 2.56 (dd, *J* = 12.6, 7.3 Hz, 1H), 2.18 – 1.85 (m, 7H).

¹³C NMR (101 MHz, CDCl₃) δ 156.0, 153.3, 145.0, 142.4, 141.5, 133.6, 132.7, 130.1, 125.9, 125.0, 123.8, 120.6, 118.0, 112.0, 110.7, 92.6, 86.8, 61.2, 59.0, 55.9, 49.3, 43.7, 32.6, 30.4, 16.1.

HRMS (ESI) *m/z* calcd. for C₂₉H₂₈N₃O₄ [M+H]⁺ 480.2080, found 480.2081.

HPLC condition: Chiralcel AD-H, *i*-PrOH/*n*-hexane = 70/30, flow rate 1.0 mL/min. λ = 280 nm, *t*(minor) = 18.3 min, *t*(major) = 27.0 min, 96:4 er.

(*S*)-N-(4-cyanophenyl)-4,4-dimethyl-2-(4-(phenylethynyl)phenyl)pyrrolidine-1-carboxamide



4m, 80% yield based on recovered starting material, $[\alpha]_D^{27} = -107$ (c 1.1, CHCl₃).

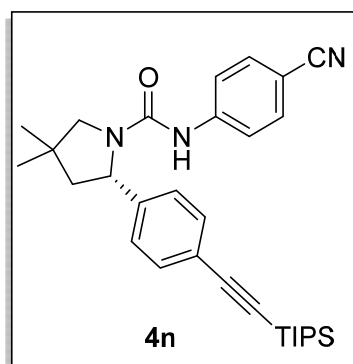
¹H NMR (400 MHz, Chloroform-*d*): δ 7.58 (d, *J* = 8.4 Hz, 2H), 7.54 – 7.52 (m, 2H), 7.46 – 7.43 (m, 2H), 7.37 – 7.33 (m, 5H), 7.23 (d, *J* = 8.4 Hz, 2H), 6.28 (s, 1H), 4.88 (dd, *J* = 9.6, 7.2 Hz, 1H), 3.82 (d, *J* = 10.4 Hz, 1H), 3.39 (d, *J* = 10.4 Hz, 1H), 2.26 (ddd, *J* = 12.8, 7.1, 1.7 Hz, 1H), 1.80 (dd, *J* = 12.8, 9.6 Hz, 1H), 1.17 (s, 3H), 1.16 (s, 3H).
¹³C NMR (101 MHz, CDCl₃): δ 153.4, 143.1, 142.3, 133.1, 132.7, 131.6, 128.6, 128.4, 125.9, 123.4, 122.9, 119.2, 118.6, 105.3, 90.3, 88.6, 61.5, 60.3, 51.7, 37.5, 26.1,

25.9.

HRMS (ESI) *m/z* calcd. for C₂₈H₂₆N₃O [M+H]⁺ 420.2070, found 420.2067.

HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 80/20, flow rate 1.0 mL/min. λ = 254 nm, *t*(minor) = 11.8 min, *t*(major) = 15.4 min, 97:3 er.

(*S*)-*N*-(4-cyanophenyl)-4,4-dimethyl-2-(4-((triisopropylsilyl)ethynyl)phenyl)pyrrolidine-1-carboxamide



4n, 81% yield based on recovered starting material, $[\alpha]_D^{27} = -116$ (c 0.6, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*): δ 7.52 (d, *J* = 7.6 Hz, 2H), 7.46 (d, *J* = 8.4 Hz, 2H), 7.29 (d, *J* = 8.0 Hz, 2H), 7.24 (d, *J* = 8.4 Hz, 2H), 6.24 (s, 1H), 4.85 (dd, *J* = 9.5, 7.0 Hz, 1H), 3.82 (d, *J* = 10.6 Hz, 1H), 3.38 (d, *J* = 10.6 Hz, 1H), 2.27 – 2.22 (m, 1H), 1.75 (dd, *J* = 12.8, 9.6 Hz, 1H), 1.16 (s, 3H), 1.15 (s, 3H), 1.13 (s, 21H).

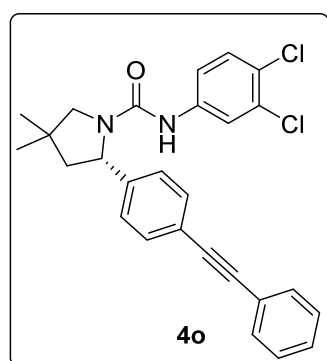
¹³C NMR (101 MHz, CDCl₃): δ 153.4, 143.1, 142.4, 133.2, 133.1, 128.4, 125.7, 119.2, 118.6, 106.2, 105.3,

91.8, 61.5, 60.2, 51.8, 37.5, 26.1, 25.9, 18.7, 11.3.

HRMS (ESI) *m/z* calcd. for C₃₁H₄₂N₃OSi [M+H]⁺ 500.3097, found 500.3095.

HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 80/20, flow rate 1.0 mL/min. λ = 254 nm, *t*(minor) = 5.7 min, *t*(major) = 9.1 min, 95.5:4.5 er.

(*S*)-*N*-(3,4-dichlorophenyl)-4,4-dimethyl-2-(4-(phenylethynyl)phenyl)pyrrolidine-1-carboxamide



4o, 75% yield based on recovered starting material, $[\alpha]_D^{27} = -79$ (c 0.2, CHCl₃).

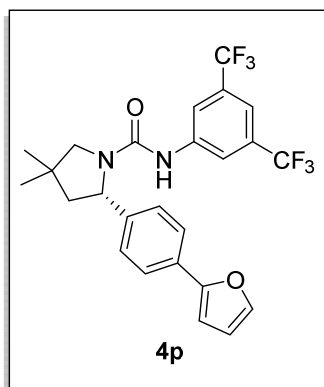
¹H NMR (400 MHz, Chloroform-*d*) δ 7.65 – 7.52 (m, 4H), 7.46 (d, *J* = 2.6 Hz, 1H), 7.42 – 7.31 (m, 5H), 7.23 (d, *J* = 8.8 Hz, 1H), 6.99 – 6.82 (m, 1H), 6.05 (s, 1H), 4.88 (t, *J* = 8.2 Hz, 1H), 3.83 (d, *J* = 10.4 Hz, 1H), 3.40 (d, *J* = 10.4 Hz, 1H), 2.28 (dd, *J* = 12.8, 7.1 Hz, 1H), 1.80 (dd, *J* = 12.8, 9.5 Hz, 1H), 1.19 (s, 3H), 1.18 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 153.8, 142.5, 138.4, 132.7, 132.5, 131.6, 130.2, 128.5, 128.4, 125.9, 125.8, 123.3, 123.0, 120.7, 118.3, 90.1, 88.7, 61.4, 60.3, 51.7, 37.6, 26.2, 25.9.

HRMS (ESI) m/z calcd. for C₂₇H₂₅Cl₂N₂O [M+H]⁺ 463.1338, found 463.1338.

HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 1.0 mL/min. λ = 254 nm, t(major) = 17.9 min, t(minor) = 28.4 min, 95:5 er.

(*S*)-*N*-(3,5-bis(trifluoromethyl)phenyl)-2-(4-(furan-2-yl)phenyl)-4,4-dimethylpyrrolidine-1-carboxamide



4p, 65% yield based on recovered starting material, [α]_D²⁷ = -52.2 (c 0.7, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*): δ 7.73 (d, *J* = 8.0 Hz, 2H), 7.62 (s, 2H), 7.48 (s, 1H), 7.41 – 7.37 (m, 3H), 6.68 (d, *J* = 3.3 Hz, 1H), 6.48 (t, *J* = 2.5 Hz, 1H), 6.34 (s, 1H), 4.89 (dd, *J* = 9.5, 7.1 Hz, 1H), 3.84 (d, *J* = 10.5 Hz, 1H), 3.41 (d, *J* = 10.5 Hz, 1H), 2.28 (dd, *J* = 12.8, 7.1 Hz, 1H), 1.83 (dd, *J* = 12.8, 9.6 Hz, 1H), 1.19 (s, 3H), 1.17 (s, 3H).

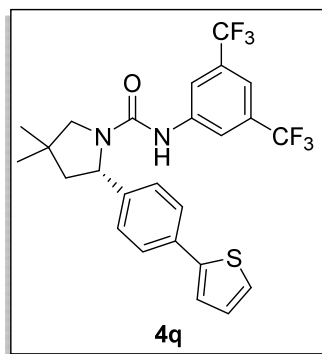
¹³C NMR (101 MHz, CDCl₃): δ 153.6, 153.1, 142.4, 140.9, 140.4, 132.0 (q, *J*_{C-F} = 32.8 Hz), 131.1, 126.2, 124.9, 123.1 (q, *J*_{C-F} = 271.0 Hz), 118.7 – 118.6 (m, 1C), 115.9 – 115.8 (m, 1C), 111.8, 105.6, 61.5, 60.3, 51.8, 37.5, 26.1, 25.9.

¹⁹F NMR (376 MHz, CDCl₃): δ -63.05.

HRMS (ESI) m/z calcd. for C₂₅H₂₃F₆N₂O₂ [M+H]⁺ 497.1664, found 497.1667.

HPLC condition: Chiralcel ID, *i*-PrOH/*n*-hexane = 96/4, flow rate 0.4 mL/min. λ = 254 nm, t(major) = 16.1 min, t(minor) = 20.3 min, 91:9 er.

(*S*)-*N*-(3,5-bis(trifluoromethyl)phenyl)-4,4-dimethyl-2-(4-(thiophen-2-yl)phenyl)pyrrolidine-1-carboxamide



4q, 66% yield based on recovered starting material, [α]_D²⁷ = -71 (c 1.1, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*): δ 7.67 – 7.64 (m, 4H), 7.42 (s, 1H), 7.36 (d, *J* = 8.0 Hz, 2H), 7.31 (dd, *J* = 9.0, 4.4 Hz, 2H), 7.09 (t, *J* = 4.4 Hz, 1H), 6.38 (s, 1H), 4.91 (t, *J* = 8.3 Hz, 1H), 3.82 (d, *J* = 10.4 Hz, 1H), 3.42 (d, *J* = 10.4 Hz, 1H), 2.29 (dd, *J* = 12.9, 7.1 Hz, 1H), 1.83 (dd, *J* = 12.8, 9.6 Hz, 1H), 1.19 (s, 3H), 1.18 (s, 3H).

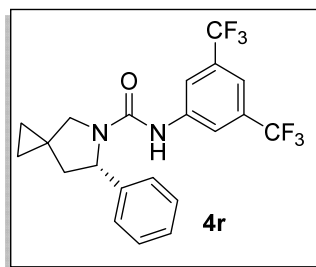
¹³C NMR (101 MHz, CDCl₃): δ 153.6, 143.4, 141.2, 140.4, 134.6, 132.0 (q, *J*_{C-F} = 33.1 Hz), 128.1, 127.0, 126.3, 125.3, 123.5, 123.1 (q, *J*_{C-F} = 270.9 Hz), 118.8 – 118.7 (m, 1C), 115.9 – 115.8 (m, 1C), 61.4, 60.3, 51.7, 37.6, 26.1, 25.9.

¹⁹F NMR (376 MHz, CDCl₃): δ -63.03.

HRMS (ESI) m/z calcd. for C₂₅H₂₃F₆N₂OS [M+H]⁺ 514.1435, found 514.1439.

HPLC condition: Chiralcel ID, *i*-PrOH/*n*-hexane = 96/4, flow rate 0.4 mL/min. λ = 254 nm, t(major) = 16.6 min, t(minor) = 20.9 min, 94:6 er.

(S)-N-(3,5-bis(trifluoromethyl)phenyl)-6-phenyl-5-azaspiro[2.4]heptane-5-carboxamide



4r, 60% yield based on recovered starting material, $[\alpha]_D^{27} = -17$ (*c* 0.3, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*): δ 7.59 (s, 2H), 7.46 – 7.42 (m, 5H), 7.39 – 7.36 (m, 1H), 6.30 (s, 1H), 4.99 (dd, *J* = 7.9, 4.9 Hz, 1H), 3.79 (d, *J* = 10.3 Hz, 1H), 3.63 (d, *J* = 10.3 Hz, 1H), 2.50 (dd, *J* = 12.6, 7.9 Hz, 1H), 1.89 (dd, *J* = 12.6, 4.9 Hz, 1H), 0.76 – 0.67 (m, 2H), 0.62 – 0.58 (m, 1H),

0.55 – 0.51 (m, 1H).

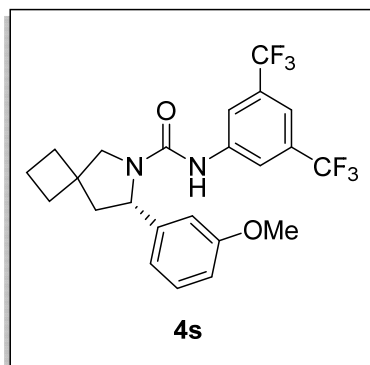
¹³C NMR (101 MHz, CDCl₃): δ 153.3, 142.0, 140.4, 132.0 (q, *J*_{C-F} = 33.1 Hz), 129.5, 128.4, 126.0, 123.2 (q, *J*_{C-F} = 271.0 Hz), 118.7 – 118.6 (m, 1C), 116.0 – 115.8 (m, 1C), 62.3, 55.3, 45.0, 19.8, 11.8, 9.0.

¹⁹F NMR (376 MHz, CDCl₃): δ -63.07.

HRMS (ESI) *m/z* calcd. for C₂₁H₁₉F₆N₂O [M+H]⁺ 429.1396, found 429.1391.

HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.6 mL/min. λ = 254 nm, *t*(major) = 12.3 min, *t*(minor) = 16.0 min, 95:5 er.

(S)-N-(3,5-bis(trifluoromethyl)phenyl)-7-(3-methoxyphenyl)-6-azaspiro[3.4]octane-6-carboxamide



4s, 69% yield based on recovered starting material, $[\alpha]_D^{27} = -61$ (*c* 0.25, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*): δ 7.58 (s, 2H), 7.42 – 7.41 (m, 1H), 7.37 – 7.33 (m, 1H), 6.95 – 6.92 (m, 1H), 6.90 – 6.87 (m, 2H), 6.36 (s, 1H), 4.76 (t, *J* = 7.2 Hz, 1H), 3.98 (d, *J* = 10.7 Hz, 1H), 3.82 (s, 3H), 3.63 (d, *J* = 10.6 Hz, 1H), 2.53 (ddd, *J* = 12.6, 7.3, 1.2 Hz, 1H), 2.13 – 1.89 (m, 7H).

¹³C NMR (101 MHz, CDCl₃): δ 160.5, 153.7, 143.5, 140.5, 131.9 (q, *J*_{C-F} = 33.1 Hz), 130.7, 123.1 (q, *J*_{C-F} =

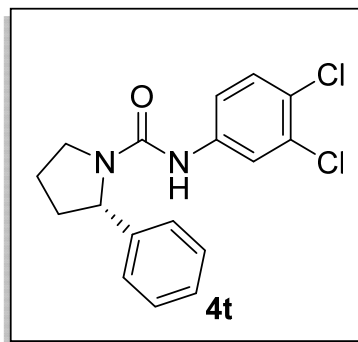
271.0 Hz), 118.7 – 118.6 (m, 1C), 118.0, 115.9 – 115.7 (m, 1C), 113.4, 111.8, 61.2, 59.9, 55.3, 49.3, 43.6, 32.8, 30.2, 16.1.

¹⁹F NMR (376 MHz, CDCl₃): δ -63.07.

HRMS (ESI) *m/z* calcd. for C₂₃H₂₃F₆N₂O₂ [M+H]⁺ 473.1664, found 473.1661.

HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 0.5 mL/min. λ = 254 nm, *t*(major) = 14.0 min, *t*(minor) = 20.3 min, 92:8 er.

(S)-N-(3,4-dichlorophenyl)-2-phenylpyrrolidine-1-carboxamide



4t, 37% yield based on recovered starting material, $[\alpha]_D^{27} = 43$ (*c* 0.1, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*) δ 7.40 (d, *J* = 7.5 Hz, 2H), 7.39 – 7.29 (m, 4H), 7.20 (d, *J* = 8.8 Hz, 1H), 6.92 (d, *J* = 8.8 Hz, 1H), 6.04 (s, 1H), 4.84 (d, *J* = 8.0 Hz, 1H), 3.85 – 3.71 (m, 2H), 2.05 – 2.44 (m, 1H), 2.04 – 1.90 (m, 3H).

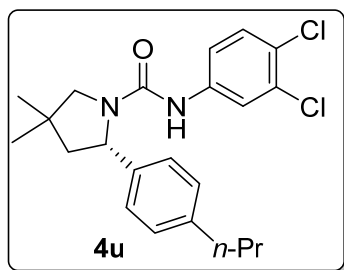
¹³C NMR (101 MHz, CDCl₃) δ 153.7, 142.2, 138.6, 132.4, 130.1, 129.4, 128.4, 126.9, 125.8, 120.7, 118.4,

61.5, 47.6, 26.3, 23.2.

HRMS (ESI) *m/z* calcd. for C₁₇H₁₇Cl₂N₂O [M+H]⁺ 335.0712, found 335.0710.

HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.6 mL/min. λ = 254 nm, *t*(minor) = 46.2 min, *t*(minor) = 53.8 min, 92.5:7.5 er.

(S)-N-(3,4-dichlorophenyl)-4,4-dimethyl-2-(4-propylphenyl)pyrrolidine-1-carboxamide



4u, 67% yield based on recovered starting material, $[\alpha]_D^{27} = -70$ (*c* 1.0, CHCl₃).

¹H NMR (400 MHz, Chloroform-*d*) δ 7.29 – 7.20 (m, 5H), 7.17 (d, *J* = 8.8 Hz, 1H), 6.82 (dd, *J* = 8.4, 2.4 Hz, 1H), 6.06 (s, 1H), 4.76 (dd, *J* = 10.0, 6.8 Hz, 1H), 3.83 (d, *J* = 10.4 Hz, 1H), 3.35 (d, *J* = 10.8 Hz, 1H), 2.61 (t, *J* = 7.6 Hz, 2H), 2.22 (ddd, *J* = 12.8, 6.8, 1.6 Hz, 1H), 1.81

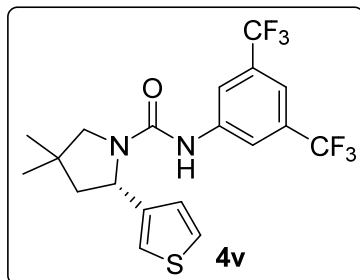
(dd, *J* = 12.8, 9.6 Hz, 1H), 1.65 (q, *J* = 7.6 Hz, 2H), 1.16 (s, 3H), 1.15 (s, 3H), 0.94 (t, *J* = 7.2 Hz, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 153.9, 143.2, 139.3, 138.7, 132.3, 130.1, 129.7, 125.9, 125.4, 120.5, 118.2, 61.5, 60.1, 52.2, 37.6, 37.1, 26.1, 25.9, 24.5, 13.8.

HRMS (ESI) *m/z* calcd. for C₂₂H₂₇Cl₂N₂O [M+H]⁺ 405.1495, found 405.1495.

HPLC condition: Chiralcel AD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.5 mL/min. λ = 254 nm, *t*(major) = 32.7 min, *t*(minor) = 37.9 min, 90.5:9.5 er.

(S)-N-(3,5-bis(trifluoromethyl)phenyl)-4,4-dimethyl-2-(thiophen-3-yl)pyrrolidine-1-carboxamide



4v, 72% yield based on recovered starting material, $[\alpha]_D^{27} = -11$ (*c* 0.4, CHCl₃) **¹H NMR (400 MHz, Chloroform-*d*)** δ 7.58 (s, 2H), 7.46 (dd, *J* = 5.1, 2.9 Hz,

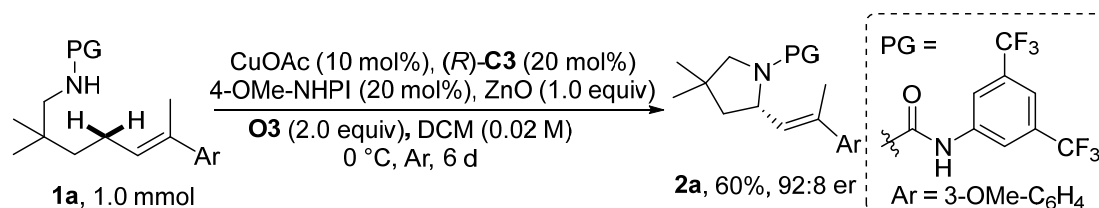
1H), 7.43 (s, 1H), 7.34 (dd, *J* = 3.0, 1.4 Hz, 1H), 7.07 (dd, *J* = 5.0, 1.4 Hz, 1H), 6.50 (s, 1H), 4.99 (dd, *J* = 9.5, 7.0 Hz, 1H), 3.79 (d, *J* = 10.6 Hz, 1H), 3.36 (d, *J* = 10.6 Hz, 1H), 2.23 (ddd, *J* = 12.7, 7.1, 1.7 Hz, 1H), 1.89 (dd, *J* = 12.7, 9.5 Hz, 1H), 1.18 (s, 3H), 1.15 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 152.7, 142.5, 139.5, 131.0 (q, $J_{\text{C-F}} = 33.1$ Hz), 127.8, 124.3, 122.1 (q, $J_{\text{C-F}} = 271$ Hz), 120.5, 117.7 – 117.6 (m, 1C), 114.8 – 114.7 (m, 1C), 58.7, 56.3, 49.7, 36.0, 25.1, 24.9.

HRMS (ESI) m/z calcd. for $\text{C}_{19}\text{H}_{19}\text{F}_6\text{N}_2\text{OS}$ $[\text{M}+\text{H}]^+$ 437.1122, found 437.1121.

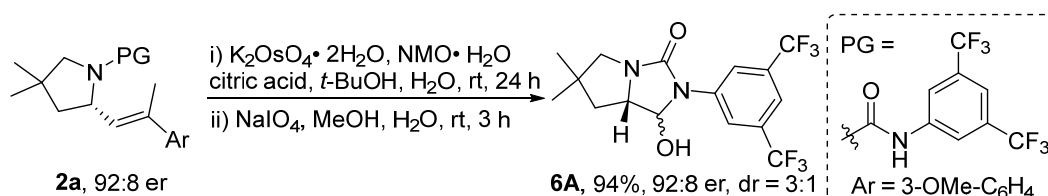
HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 0.6 mL/min. $\lambda = 254$ nm, $t(\text{major}) = 9.3$ min, $t(\text{minor}) = 12.4$ min, 68:32 er.

Preparative scale enantioselective radical allylic C–H amination under conditions A:



To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **1a** (502 mg, 1.0 mmol), CuOAc (12.4 mg 10 mol%), (*R*)-**C3** (160 mg, 20 mol%), 4-OMe-NHPI (40 mg, 20 mol%) and ZnO (80 mg, 1.0 mmol). The tube was evacuated and backfilled with argon for three times, the **O3** (480 μL , 2.0 mmol) and dry DCM (50.0 mL) was added *via* syringe. The tube was stirred at 10 °C for 3 days. After completion, the reaction mixture was filtered through celite and the filtrate was evaporated under reduced pressure, and the residue was purified by flash chromatography (petroleum ether/ ethyl acetate = 20/1–10/1) to give the corresponding product **2a** (300 mg) in 60% isolated yield with 92:8 er.

Versatile transformation¹⁰

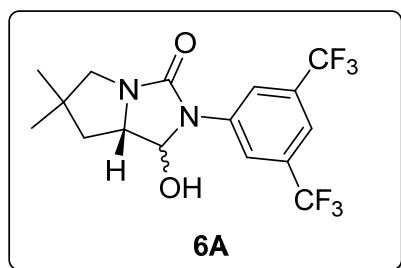


To a flame-dried Schlenk tube equipped with a magnetic stir bar was added **2a** (50.0 mg, 0.1 mmol). Then *t*-BuOH (1.0 mL) and deionized water (1.0 mL) was added while vigorous stirring followed by the addition of citric acid (38.5 mg, 0.2 mmol). Then $\text{K}_2\text{OsO}_4 \cdot 2\text{H}_2\text{O}$ (1.5 mg, 4 mol%) and $\text{NMO} \cdot \text{H}_2\text{O}$ (17.5 mg, 0.15 mmol) were sequentially added and the resulting mixture was vigorously stirred in air at rt for ca. 24 h. The resulting mixture was evaporated under reduced pressure until most of the *t*-BuOH had been removed. Then the mixture was diluted with HCl (1 M) solution and EtOAc and the separated aqueous layer was extracted with EtOAc. The combined organic layer was dried over anhydrous Na_2SO_4 , filtered through a cotton plug and evaporated under reduced pressure. The residue thus obtained was used directly for the

next step without purification.

To a solution of the crude product obtained above in MeOH (1.5 mL) was added deionized H₂O (0.5 mL) and NaIO₄ (42.8 mg, 0.2 mmol) sequentially. the resulting mixture was vigorously stirred in air at rt for ca. 3 h. Then the reaction mixture was repetitively diluted with MeCN and evaporated under reduced pressure until a dried residue was obtained. This residue was mixed with EtOAc and the resulting mixture was sonicated until all the solid was well dispersed in EtOAc. The suspension thus obtained was filtered through a celite pad and the filtrate was evaporated under reduced pressure, and the residue was purified by flash column chromatography (petroleum ether/ ethyl acetate =20/1-8/1) to give the corresponding product **6A** (36.0 mg) in 94% isolated yield.

2-(3,5-bis(trifluoromethyl)phenyl)-1-hydroxy-6,6-dimethylhexahydro-3H-pyrrolo[1,2-c]imidazol-3-one



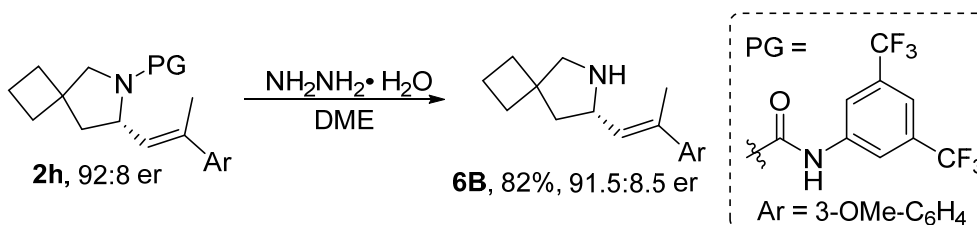
¹H NMR (400 MHz, Chloroform-*d*) δ 8.21 (s, 2H), 7.55 (s, 1H), 5.59 (*dd*, *J* = 5.2 Hz, 0.2H), 5.44 (*d*, *J* = 9.1 Hz, 0.8H), 5.14 (*d*, *J* = 12.0 Hz, 0.2H), 4.22 – 4.19 (m, 1H), 3.74 (*dd*, *J* = 10.5, 6.2 Hz, 0.8H), 3.40 (*d*, *J* = 11.4 Hz, 0.8H), 2.76 (*d*, *J* = 11.4 Hz, 1H), 2.56 (*d*, *J* = 10.0 Hz, 0.2H), 1.88 (*dd*, *J* = 6.4 Hz, 0.8H), 1.80 (*d*, *J* = 11.5 Hz, 0.2H), 1.58 (*dd*, *J* = 6.0 Hz, 0.2H), 1.31 (*d*, *J* = 11.5 Hz, 0.8H), 1.09 (*d*, *J* = 13.1 Hz, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 160.1, 156.6, 141.1, 139.8, 132.1 (*q*, *J* = 33.0 Hz, 1C), 123.1 (*q*, *J* = 271.1 Hz, 1C), 118.9(7) -118.9(2) (*m*, 1C), 117.4, 117.0 - 116.8(*m*, 1C), 115.6, 83.3, 82.0, 66.0, 60.2, 59.3, 56.3, 43.6, 42.0, 40.4, 37.8, 28.5, 27.7, 27.3, 26.2. (The italics data represents minor diastereomer).

¹⁹F NMR (376 MHz, CDCl₃) δ -62.92 (minor), -62.99 (major).

HRMS (ESI) *m/z* calcd. for C₁₆H₁₇F₆N₂O₂ [M+H]⁺ 383.1189, found 383.1186.

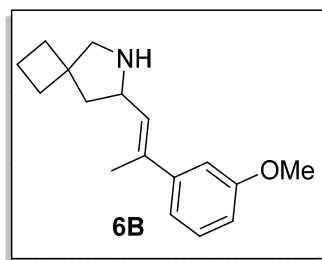
HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 96/4, flow rate 0.4 mL/min. λ = 254 nm, minor diastereomer: *t*(minor) = 15.3 min, *t*(major) = 17.4 min, 92:8 er; major diastereomer: *t*(minor) = 20.0 min, *t*(major) = 21.8 min, 92:8 er.



To a flame-dried Schlenk tube equipped with a magnetic stir bar was charged with **2h** (25.6 mg, 0.05 mmol), hydrazine hydrate (30 μ L, 0.5 mmol) and 1,2-Dimethoxyethane (2.0 mL). The sealed tube was then stirred at 80 °C for 96 h. Upon completion (monitored by TLC), the reaction mixture was cooled down to temperature. Then the crude product was concentrated *in vacuo* and purified by flash column chromatography using dichloromethane/methanol (50/1) as the eluent to give **6B** (10.6 mg) in 82%

isolated yield.

(E)-7-(2-(3-methoxyphenyl)prop-1-en-1-yl)-6-azaspiro[3.4]octane



¹H NMR (400 MHz, Chloroform-*d*) δ 7.20 (t, *J* = 7.9 Hz, 1H), 7.01 – 6.98 (m, 2H), 6.82 – 6.79 (m, 1H), 6.05 – 5.97 (m, 1H), 4.51 – 4.45 (m, 1H), 3.81 (s, 3H), 3.66 (brs, 1H), 3.22 (s, 2H), 2.25 (dd, *J* = 12.8, 6.4 Hz, 1H), 2.10 (d, *J* = 1.2 Hz, 3H), 2.07 – 1.94 (m, 5H), 1.91 – 1.83 (m, 2H).

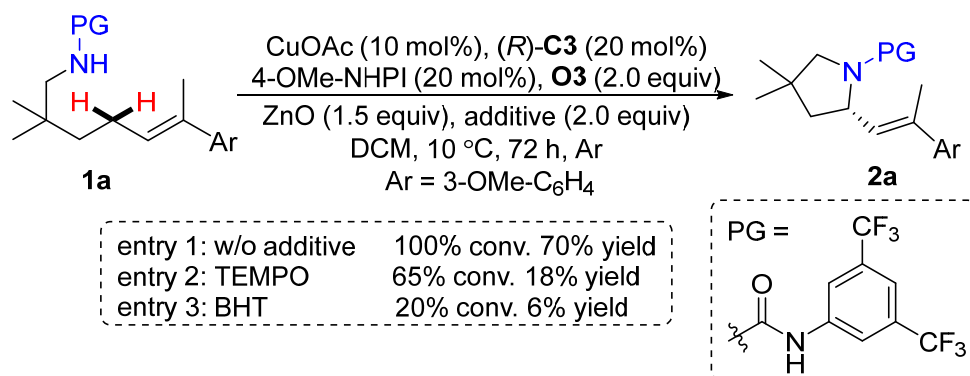
¹³C NMR (101 MHz, CDCl₃) δ 159.6, 143.3, 141.8, 129.3, 122.0, 118.5, 113.6, 111.7, 56.9, 55.3, 54.9, 45.0, 44.3, 33.5,

29.9, 16.6, 16.0.

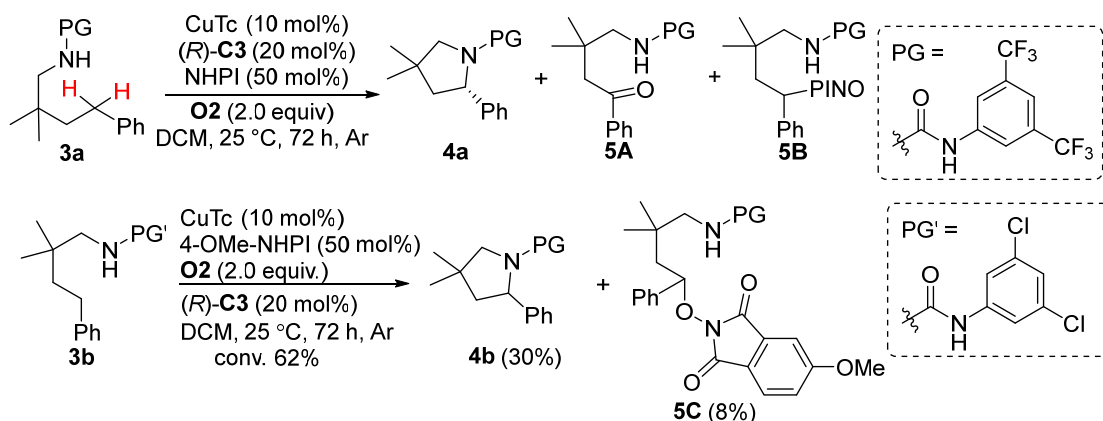
HRMS (ESI) *m/z* calcd. for C₁₇H₂₄NO [M+H]⁺ 258.1852, found 258.1848.

HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.5 mL/min. λ = 254 nm, *t*(minor) = 24.7 min, *t*(major) = 36.4 min, 91.5:8.5 er.

Mechanistic studies:



To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **1a** (50.5 0.1 mmol), CuOAc (1.24 mg 10 mol%), (*R*)-**C3** (16.0 mg, 20 mol%), 4-OMe-NHPI (4.0 mg, 20 mol%), ZnO (12.0 mg, 0.15 mmol) and additive (0.2 mmol). The tube was evacuated and backfilled with argon for three times, the **O3** (48 μ L, 0.2 mmol) and dry DCM (8.0 mL) was added *via* syringe. The tube was stirred at 10 °C for 3 days. The solvent was removed under reduced pressure, and the conversion and yield were determined by crude ¹H NMR using 1,3,5-trimethoxybenzene as internal standard.

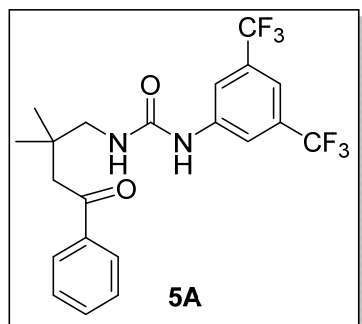


To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **3a** (43.2 mg, 0.1 mmol), CuTc (1.9 mg 10 mol%), (R)-C3 (16.0 mg, 20 mol%) and NHPI (8.2 mg, 50 mol%). The tube was evacuated and backfilled with argon for three times, the **O2** (54 μ L, 0.3 mmol) and dry DCM (8.0 mL) was added *via* syringe. The tube was stirred at rt for 3 days. The reaction stopped and the solvent was removed under reduced pressure, and the residue was purified by flash column chromatography (petroleum ether/ethyl acetate =20/1-10/1) to give the corresponding product **4a** in 34% isolated yield with 65% conversion along with the detection of byproducts **5A** and **5B**.

To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **3b** (36.5 0.1 mmol), CuTc (1.9 mg 10 mol%), (R)-C3 (16.0 mg, 20 mol%), and 4-OMe-NHPI (9.8 mg, 50 mol%). The tube was evacuated and backfilled with argon for three times, the **O2** (36 μ L, 0.2 mmol) and dry DCM (8.0 mL) was added *via* syringe. The tube was stirred at rt for 3 days. The solvent was removed under reduced pressure, and the residue was purified by flash column chromatography (petroleum ether/ethyl acetate =20/1-10/1) to give the corresponding product **4b** in 30% isolated yield with 62% conversion along with the detection of byproducts **5C** (4.5 mg, 8%).

1-(3,5-bis(trifluoromethyl)phenyl)-3-(2,2-dimethyl-4-oxo-4-phenylbutyl)urea

¹H NMR (400 MHz, Chloroform-*d*) δ 7.95 (d, *J* = 7.7 Hz, 2H), 7.90 (s, 2H), 7.59 (t, *J* = 7.4 Hz, 1H), 7.46 (t, *J* = 7.6 Hz, 2H), 7.42 (s, 1H), 6.05 (s, 1H), 3.28 (d, *J* = 6.7 Hz, 2H), 2.94 (s, 2H), 1.04 (s, 6H).



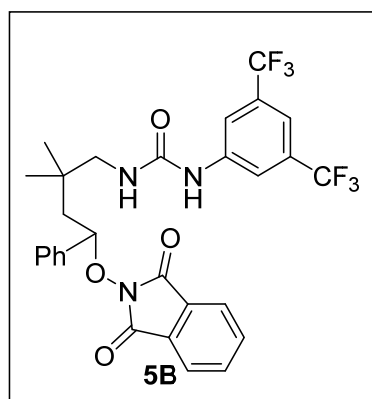
¹³C NMR (101 MHz, CDCl₃) δ 202.9, 155.9, 141.1, 137.6, 133.9, 132.0 (q, *J*_{C-F} = 33.1 Hz), 128.8, 128.5, 123.3 (q, *J*_{C-F} = 271.1 Hz), 118.5, 115.5, 49.7, 36.1, 29.72, 26.5.

¹⁹F NMR (376 MHz, CDCl₃): δ -63.11.

HRMS (ESI) *m/z* calcd. for C₂₁H₂₁F₆N₂O₂ [M+H]⁺

447.1507, found 447.1504.

1-(3,5-bis(trifluoromethyl)phenyl)-3-(4-((1,3-dioxoisindolin-2-yl)oxy)-2,2-dimethyl-4-phenylbutyl)urea.



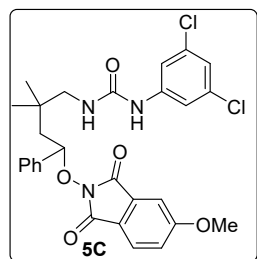
¹H NMR (400 MHz, Chloroform-*d*) δ 8.02 (s, 2H), 7.97 (s, 1H), 7.73 (s, 4H), 7.46 (s, 1H), 7.43 (dd, J = 6.6, 3.0 Hz, 2H), 7.35 – 7.29 (m, 3H), 6.33 (s, 1H), 5.57 (dd, J = 9.1, 1.7 Hz, 1H), 3.57 (dd, J = 14.0, 7.1 Hz, 1H), 3.36 (dd, J = 14.2, 5.9 Hz, 1H), 2.42 (dd, J = 15.8, 9.1 Hz, 1H), 1.76 (dd, J = 15.8, 1.7 Hz, 1H), 1.15 (s, 3H), 1.08 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 164.2, 155.4, 141.4, 138.5, 134.9, 132.0 (q, J_{C-F} = 33.1 Hz), 129.5, 128.5, 128.2, 128.1, 123.7, 123.4 (q, J_{C-F} = 271.1 Hz), 118.1, 115.0, 87.0, 48.9, 46.9, 34.8, 28.6, 24.4.

¹⁹F NMR (376 MHz, CDCl₃): δ -62.99.

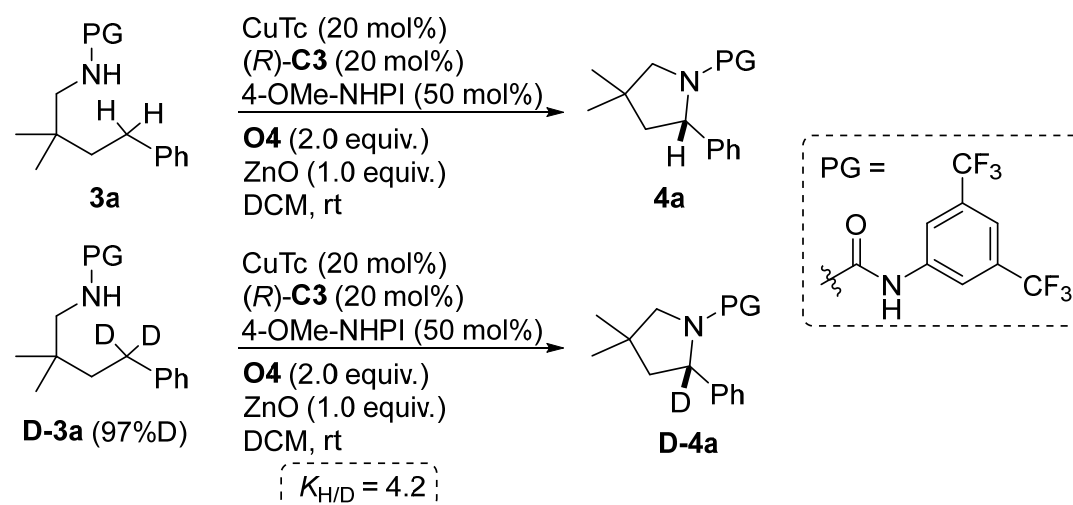
HRMS (ESI) m/z calcd. for C₂₉H₂₆F₆N₃O₄ [M+H]⁺ 594.1828, found 594.1827.

1-(3,5-dichlorophenyl)-3-(4-((5-methoxy-1,3-dioxoisindolin-2-yl)oxy)-2,2-dimethyl-4-phenylbutyl)urea



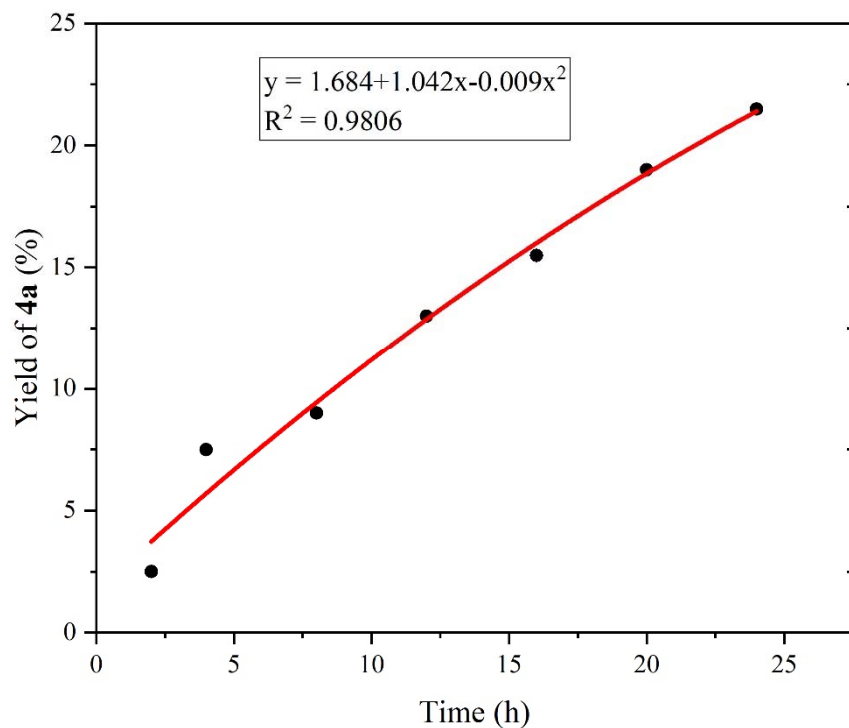
¹H NMR (400 MHz, Chloroform-*d*) δ 7.66 (d, J = 8.3 Hz, 1H), 7.49 (d, J = 1.6 Hz, 2H), 7.47 (s, 1H), 7.42 – 7.40 (m, 2H), 7.31 – 7.29 (m, 3H), 7.19 (d, J = 2.4 Hz, 1H), 7.14 (dd, J = 8.0, 2.0 Hz, 1H), 6.96 (t, J = 2.0 Hz, 1H), 6.26 (s, 1H), 5.57 – 5.47 (m, 1H), 3.90 (s, 3H), 3.56 (dd, J = 14.0, 7.6 Hz, 1H), 3.26 (dd, J = 14.4, 5.2 Hz, 1H), 2.39 (dd, J = 16.0, 9.2 Hz, 1H), 1.74 (d, J = 12.0 Hz, 1H), 1.11 (s, 3H), 1.06 (s, 3H). **¹³C NMR (101 MHz, CDCl₃)** δ 165.3, 164.5, 163.9, 155.2, 141.9, 138.7, 134.9, 130.8, 129.4, 128.4, 128.2, 125.7, 121.8, 120.0, 119.7, 116.8, 109.1, 86.9, 56.2, 48.8, 47.4, 34.8, 28.9, 24.3.

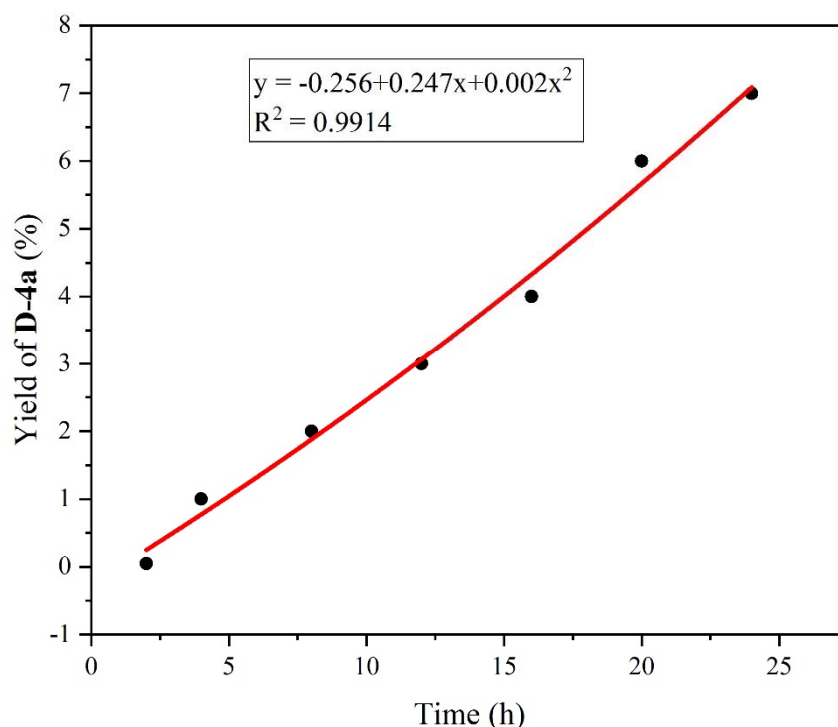
HRMS (ESI) m/z calcd. for C₂₈H₂₈Cl₂N₃O₅ [M+H]⁺ 556.1401, found 556.1404.



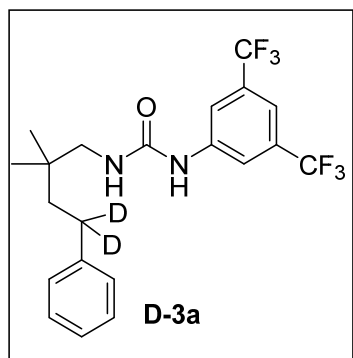
To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **3a** (43.3 mg, 0.1 mmol) or **D-3a** (43.5 mg, 0.1 mmol), CuTc (3.8 mg 20 mol%), (*R*)-**C3** (16 mg,

20 mol%), 4-OMe-NHPI (10 mg, 50 mol%) and ZnO (8.0 mg, 0.1 mmol). The tube were evacuated and backfilled with argon for three times, the **O4** (48 μ L, 0.2 mmol) and dry DCM (6.0 mL) was added *via* syringe. The tube was stirred at rt and the yield of **4a** and **D-4a** were determined by crude ^1H NMR using 1,3,5-trimethoxybenzene as internal standard at different times to determine the intermolecular KIE value.





1-(3,5-bis(trifluoromethyl)phenyl)-3-(2,2-dimethyl-4-phenylbutyl-4,4-d₂)urea



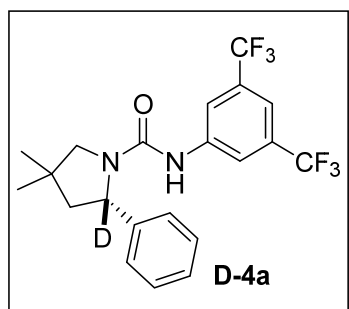
¹H NMR (400 MHz, Chloroform-*d*) δ 8.49 (s, 1H), 7.68 (s, 2H), 7.41 (s, 1H), 7.34 – 7.01 (m, 5H), 6.15 (t, *J* = 6.4 Hz, 1H), 3.13 (d, *J* = 6.0 Hz, 2H), 2.52 (t, *J* = 8.8 Hz, 0.07 H), 1.48 (s, 2H), 0.93 (s, 6H).

¹³C NMR (101 MHz, CDCl₃) δ 156.5, 142.4, 140.3, 132.1 (q, *J*_{C-F} = 33.1 Hz), 128.4, 128.1, 125.0, 123.0 (q, *J*_{C-F} = 271.1 Hz), 118.6, 115.9, 50.1, 41.9, 34.5, 24.6.

¹⁹F NMR (376 MHz, CDCl₃): δ -63.35.

HRMS (ESI) *m/z* calcd. for C₂₁H₂₁D₂F₆N₂O [M+H]⁺ 435.1835, found 435.1832.

(S)-N-(3,5-bis(trifluoromethyl)phenyl)-4,4-dimethyl-2-phenylpyrrolidine-2-d-1-carboxamide

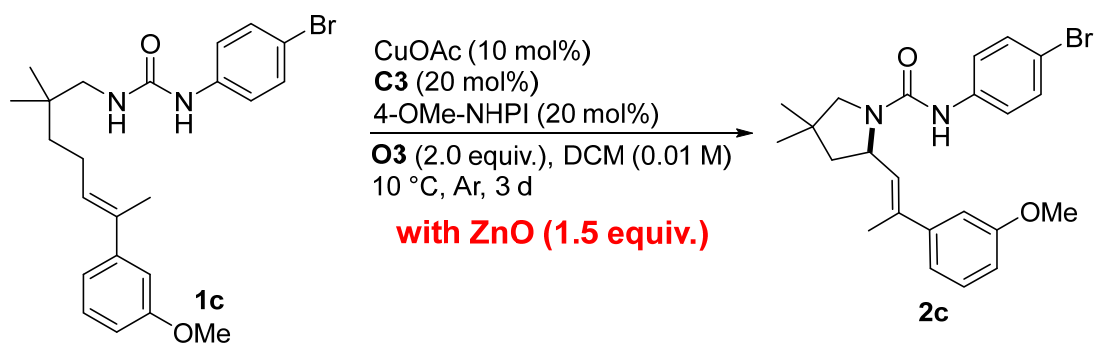


¹H NMR (400 MHz, Chloroform-*d*) δ 7.53 (s, 2H), 7.49 – 7.33 (m, 6H), 6.28 (s, 1H), 3.85 (d, *J* = 10.4 Hz, 1H), 3.40 (d, *J* = 10.4 Hz, 1H), 2.27 (dd, *J* = 12.4, 1.8 Hz, 1H), 1.84 (d, *J* = 12.4 Hz, 1H), 1.18 (s, 3H), 1.17 (s, 3H)

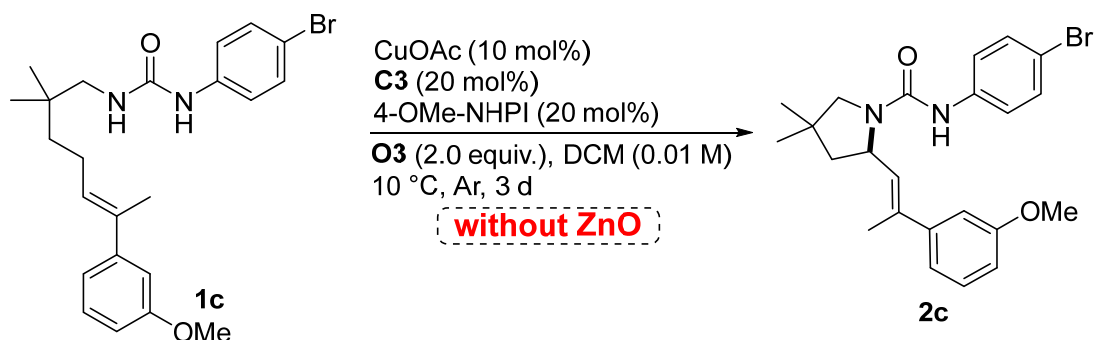
¹³C NMR (101 MHz, CDCl₃) δ 153.6, 142.0, 140.5, 131.9 (q, *J*_{C-F} = 33.3 Hz), 129.7, 128.6, 125.9, 123.1 (q, *J*_{C-F} = 271.0 Hz), 118.7 – 118.6 (m, 1C), 115.9 – 115.7 (m, 1C), 61.3 (t, *J*_{C-D} = 21.3 Hz), 60.3, 51.9, 37.3, 26.1, 25.9.

HRMS (ESI) m/z calcd. for $C_{21}H_{20}DF_6N_2O$ $[M+H]^+$ 432.1615, found 432.1613.

Non-linear effect experiments:



To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **1c** (22.0 mg, 0.05 mmol), CuOAc (0.62 mg 10 mol%), **C3** (8.0 mg, 20 mol%), 4-OMe-NHPI (2.0 mg, 20 mol%) and ZnO (6.0 mg, 0.075 mmol). The tube was evacuated and backfilled with argon for three times, the **O3** (24 μ L, 0.1 mmol) and dry DCM (5.0 mL) was added *via* syringe. The tube was stirred at 10 °C for 3 days. After completion, the reaction mixture was filtered through a pad of celite. The filtrate was used to collect the ee value by HPLC analysis on a Chiral column.

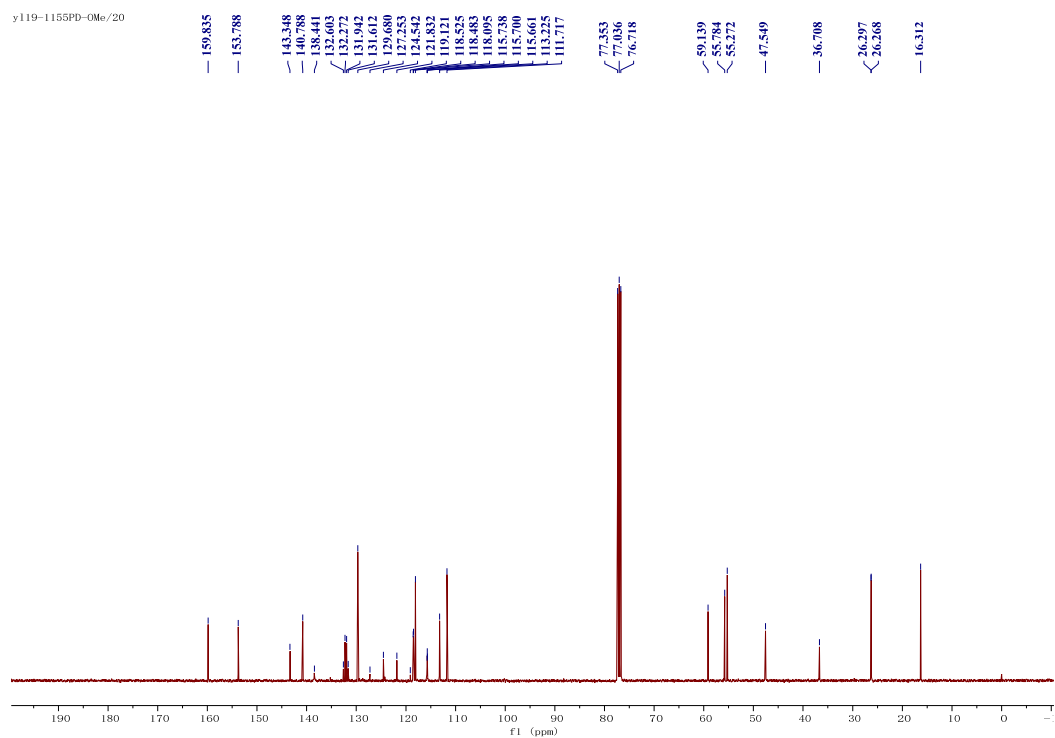
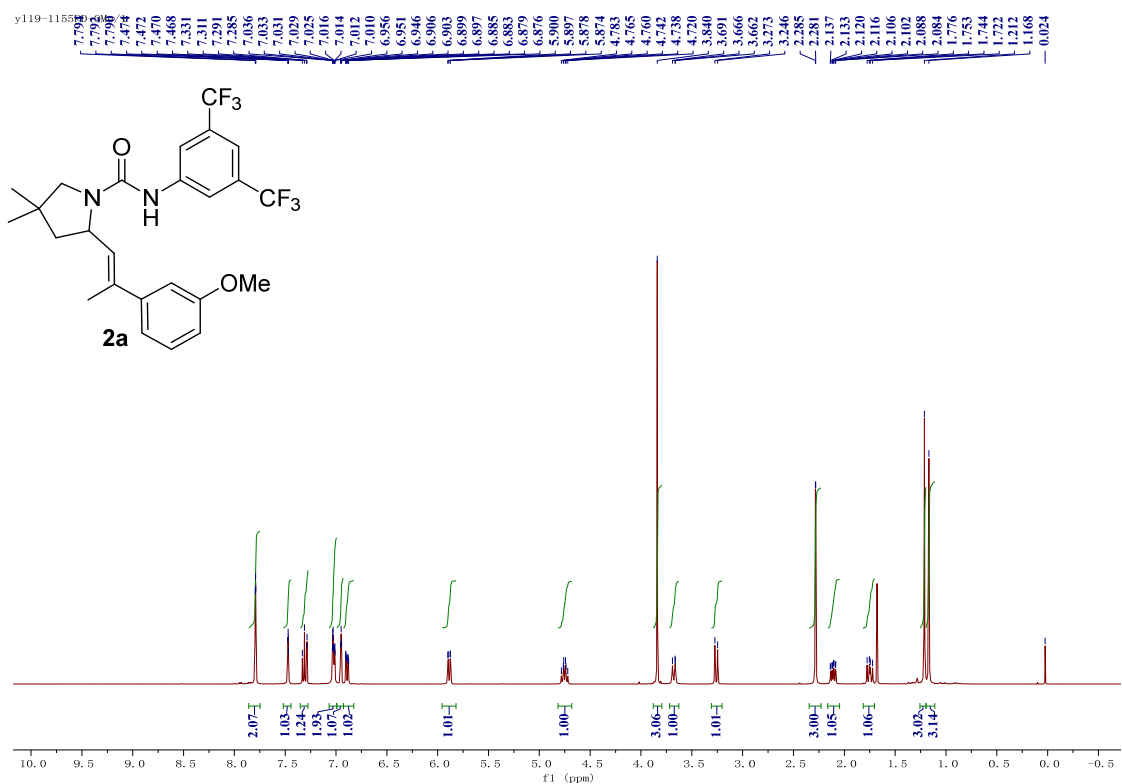


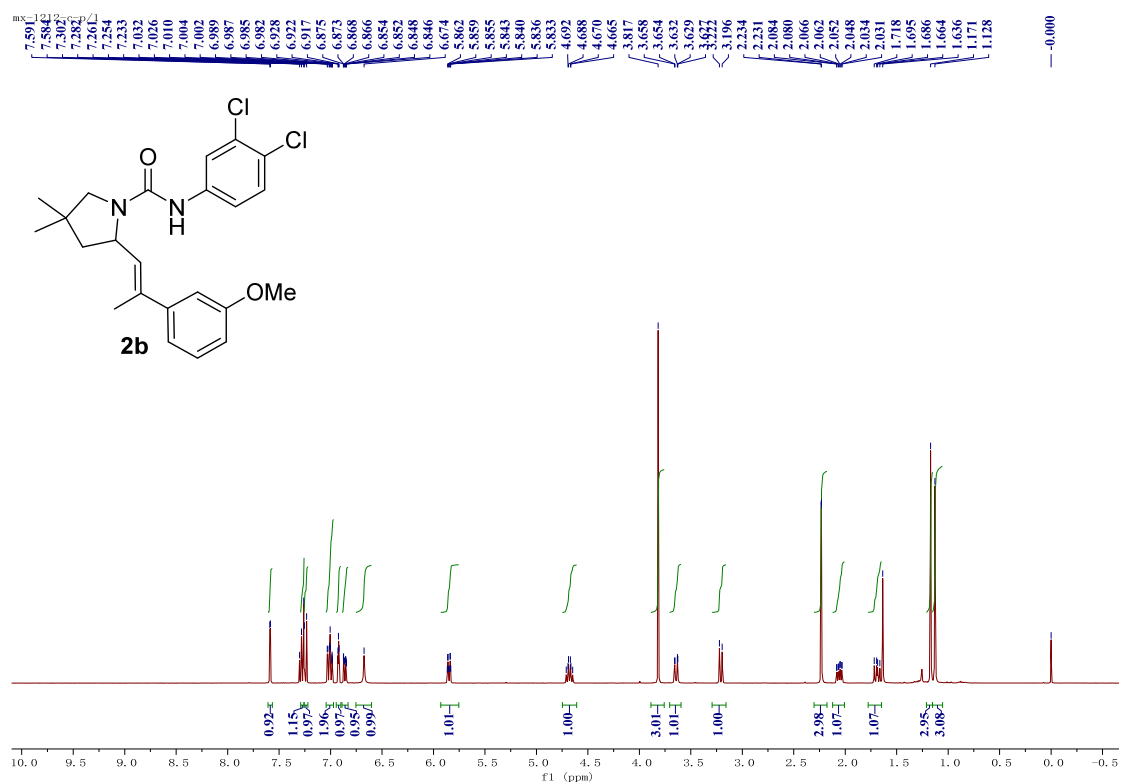
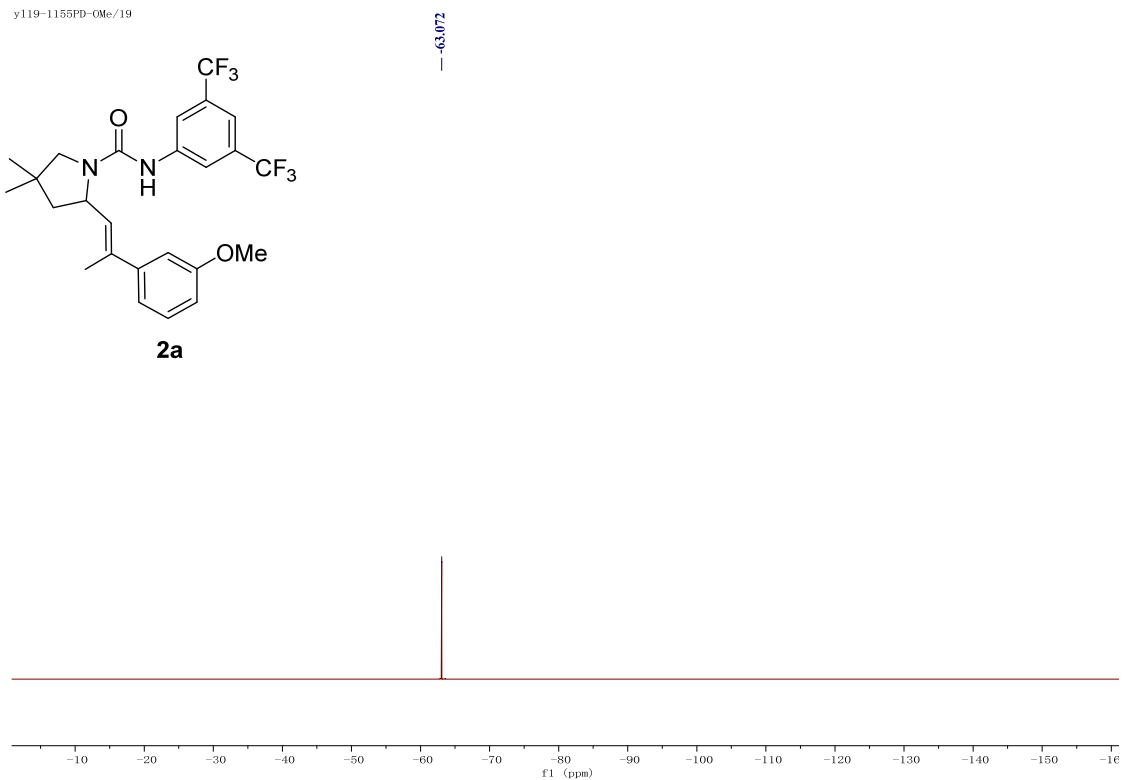
To a flame-dried Schlenk tube equipped with a magnetic stir bar were added **1c** (22.0 mg, 0.05 mmol), CuOAc (0.62 mg 10 mol%), **C3** (8.0 mg, 20 mol%), and 4-OMe-NHPI (2.0 mg, 20 mol%). The tube was evacuated and backfilled with argon for three times, the **O3** (24 μ L, 0.1 mmol) and dry DCM (5.0 mL) was added *via* syringe. The tube was stirred at 10 °C for 3 days. After completion, the reaction mixture was filtered through a pad of celite. The filtrate was used to collect the ee value by HPLC analysis on a Chiral column.

References:

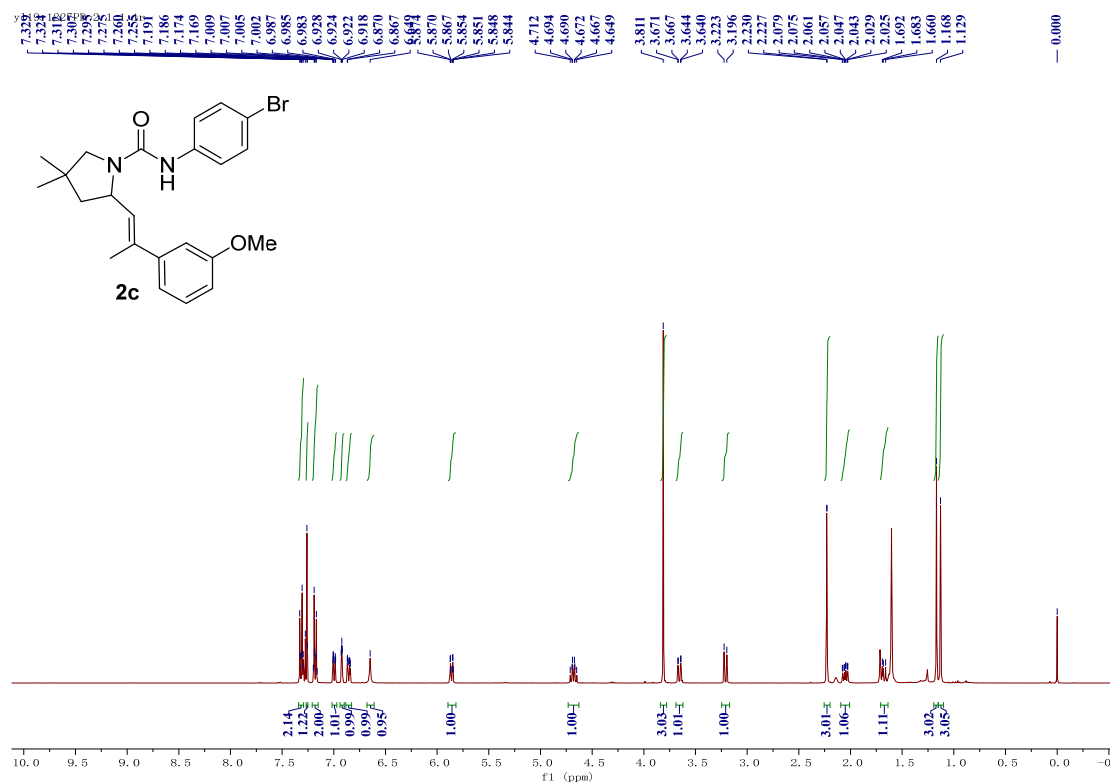
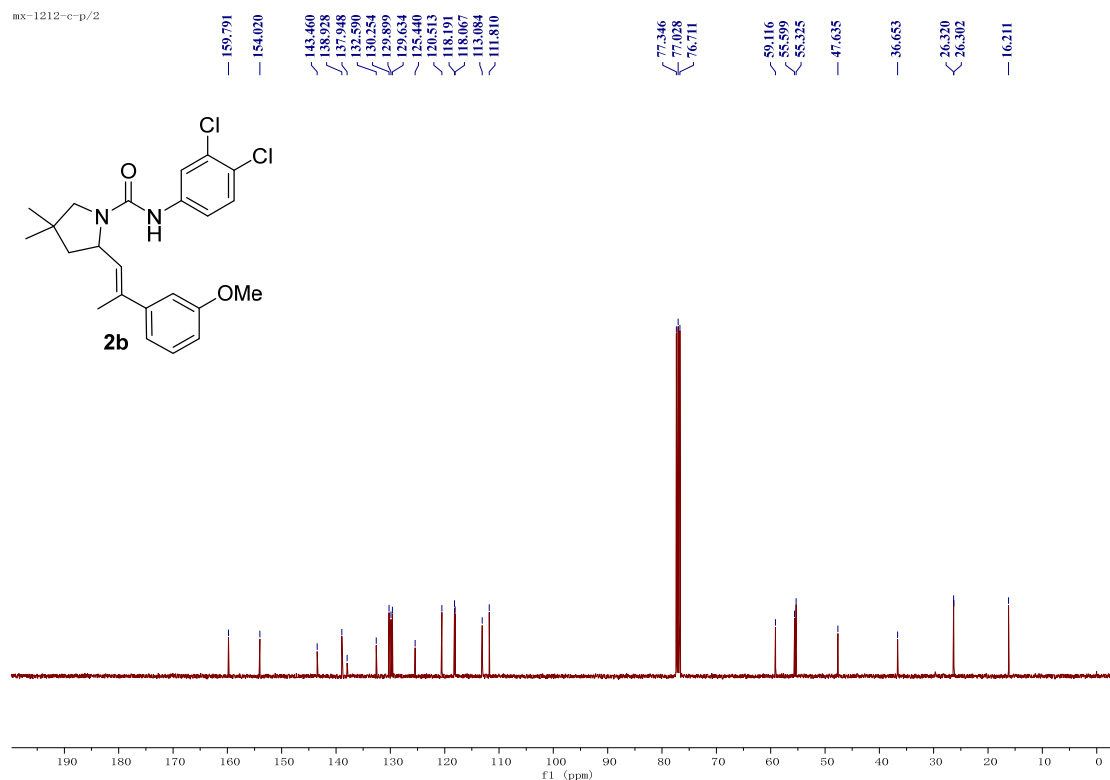
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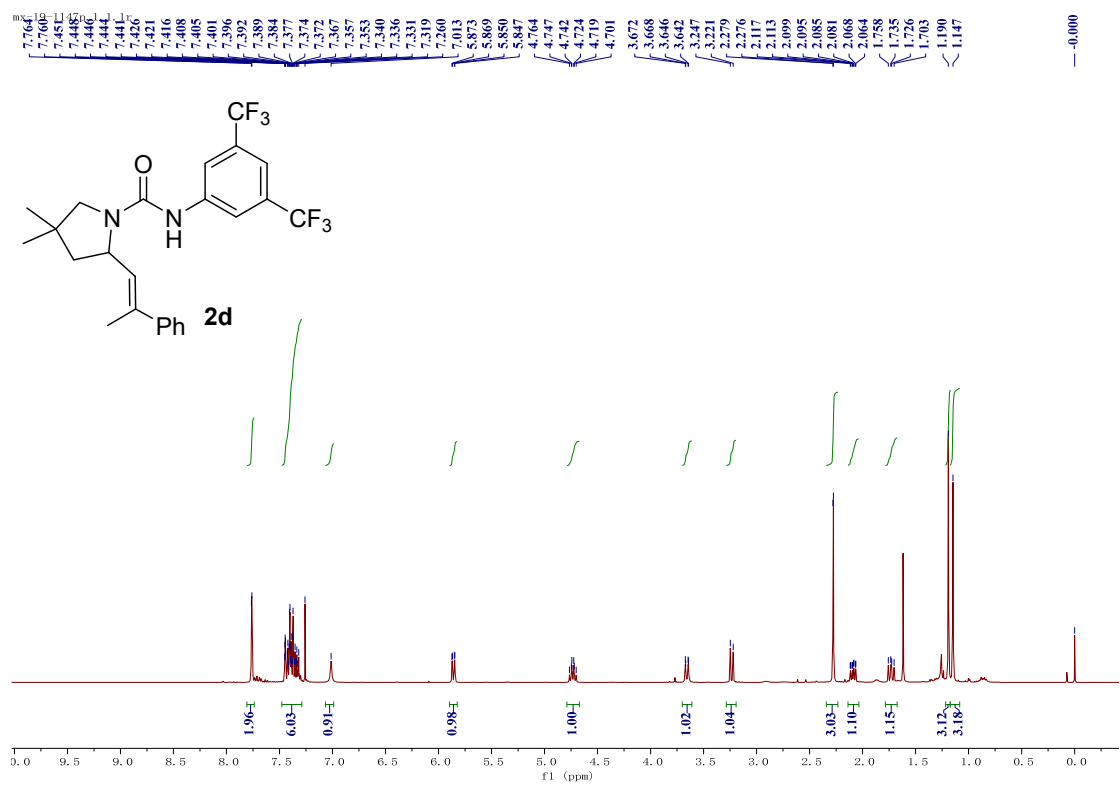
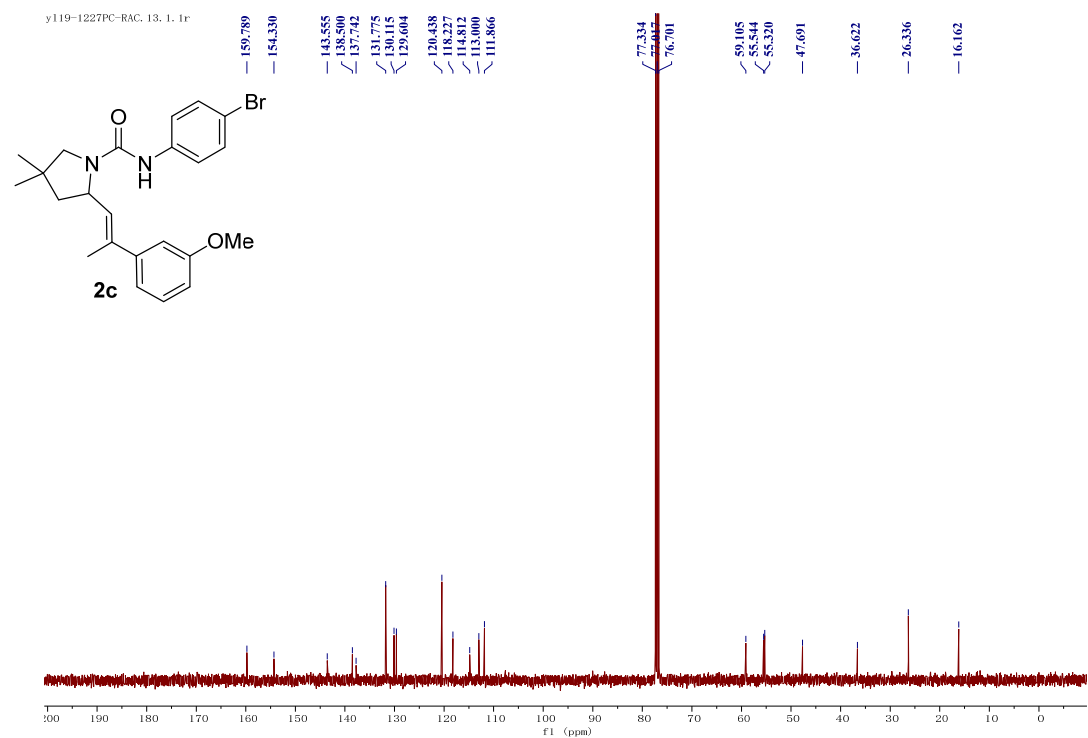
NMR Spectra



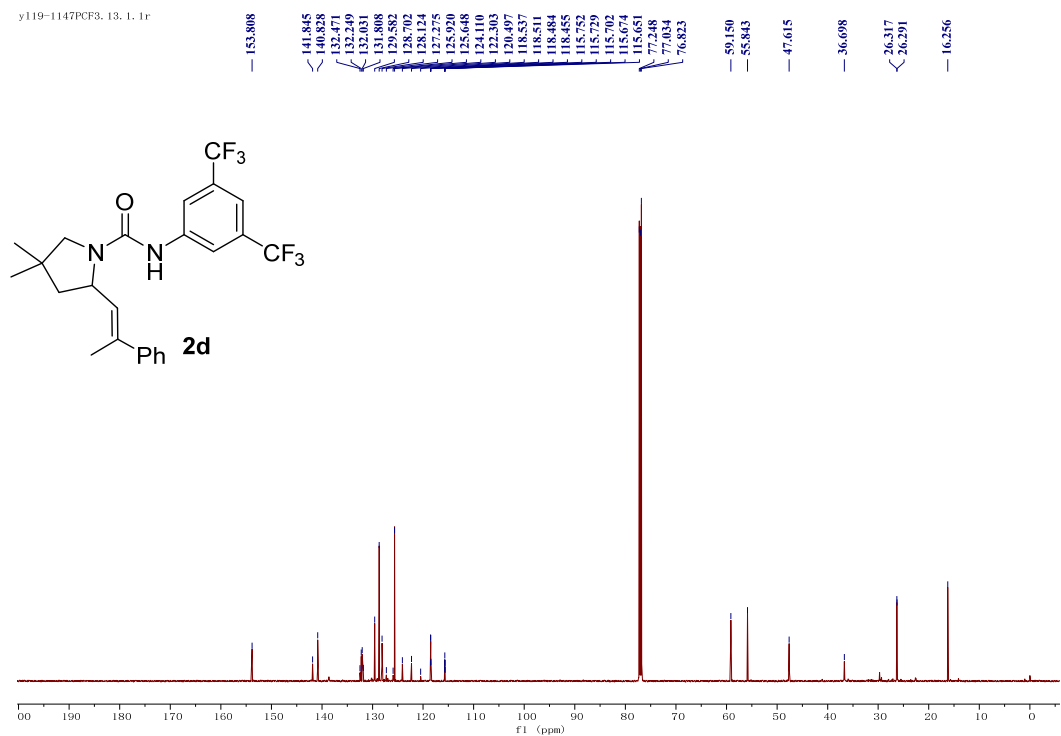


mx-1212-c-p/2

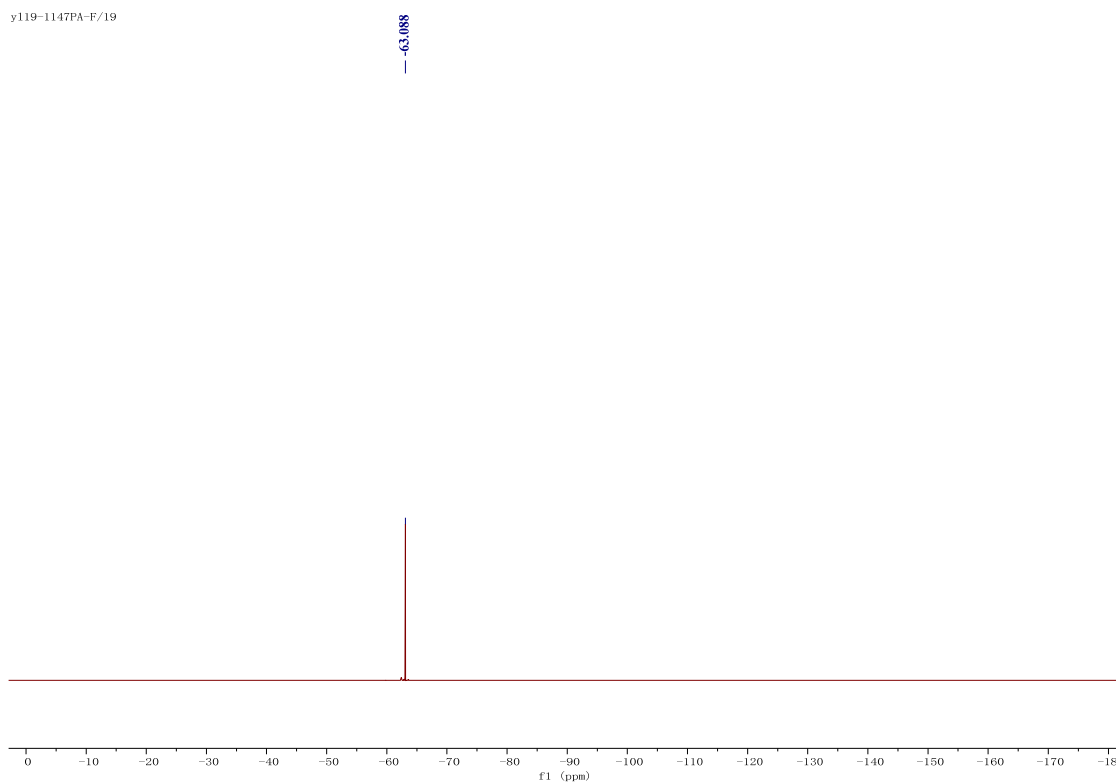


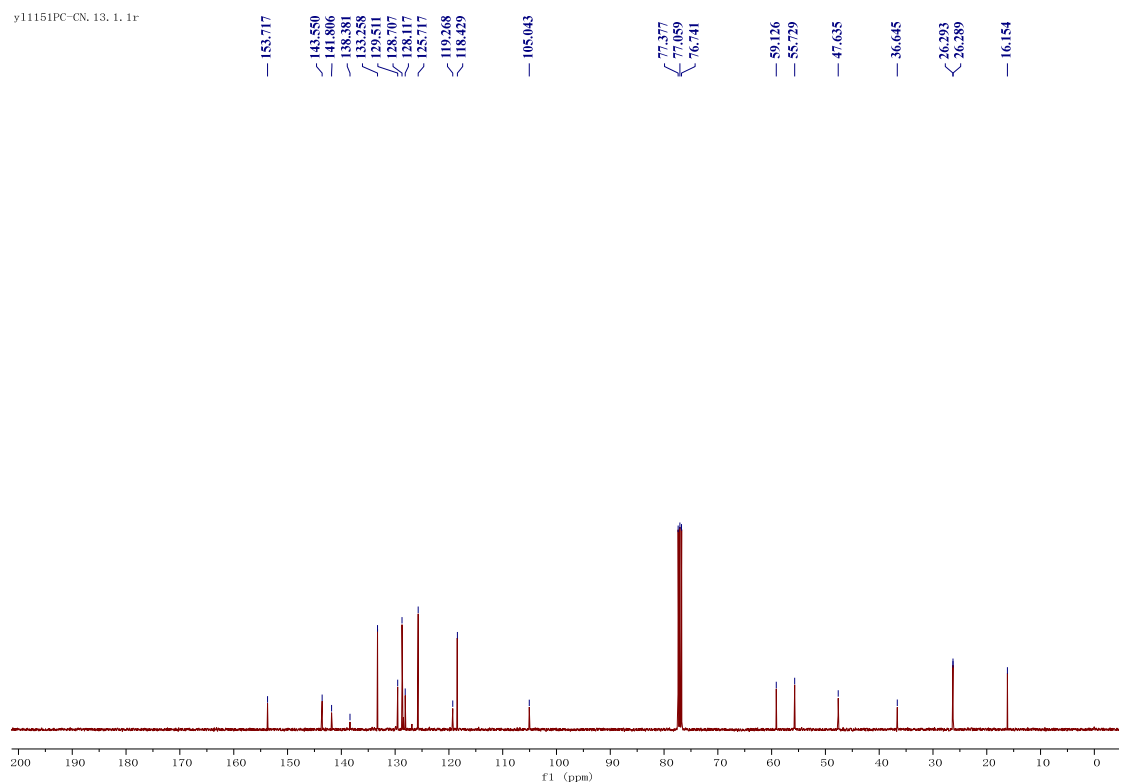
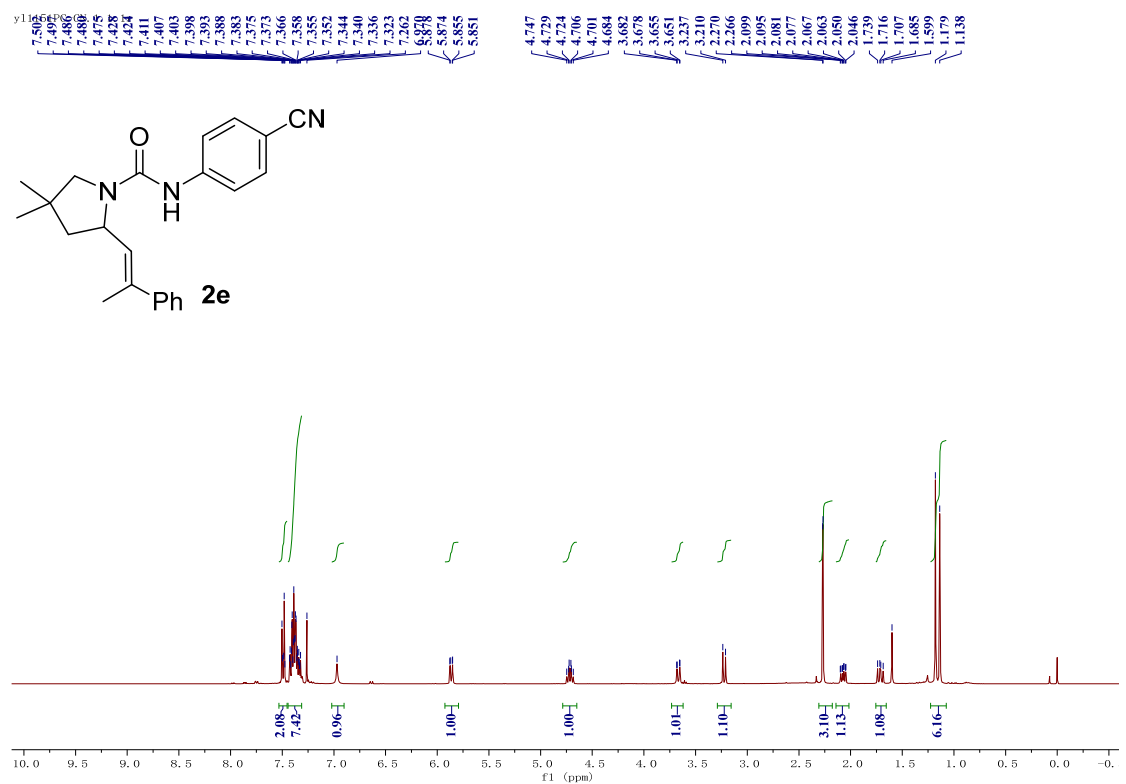


y119-1147PCF3, 13, 1, 1r

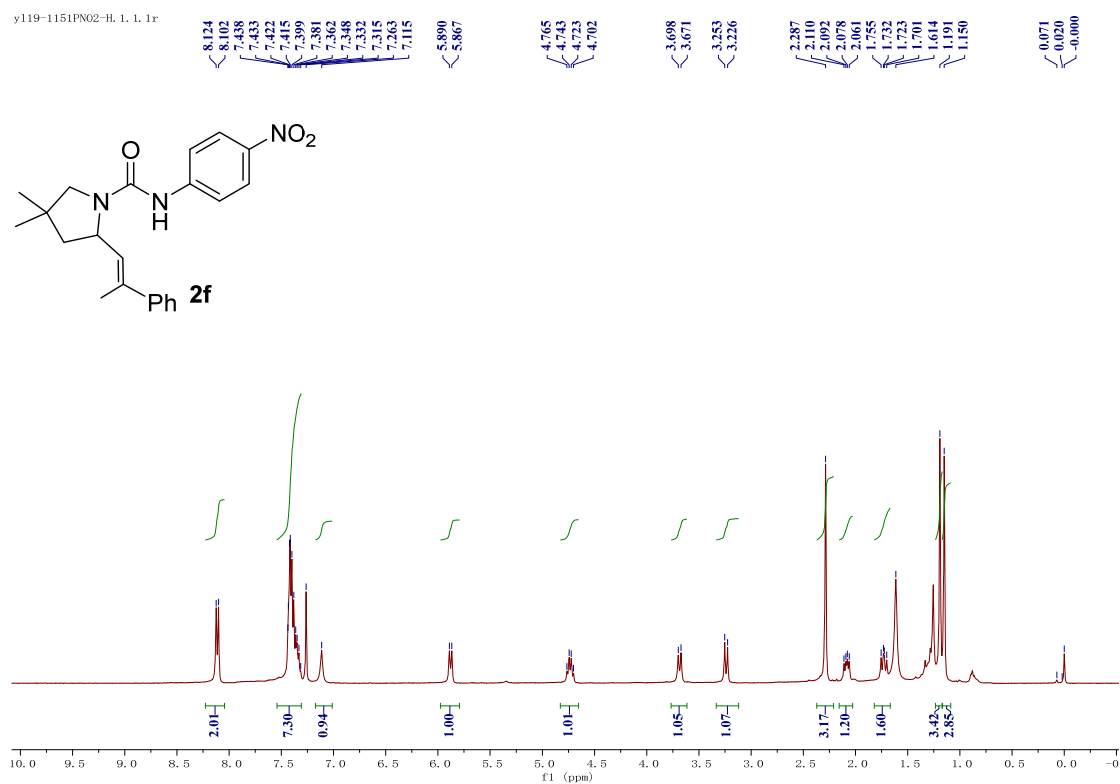


y119-1147PA-F/19

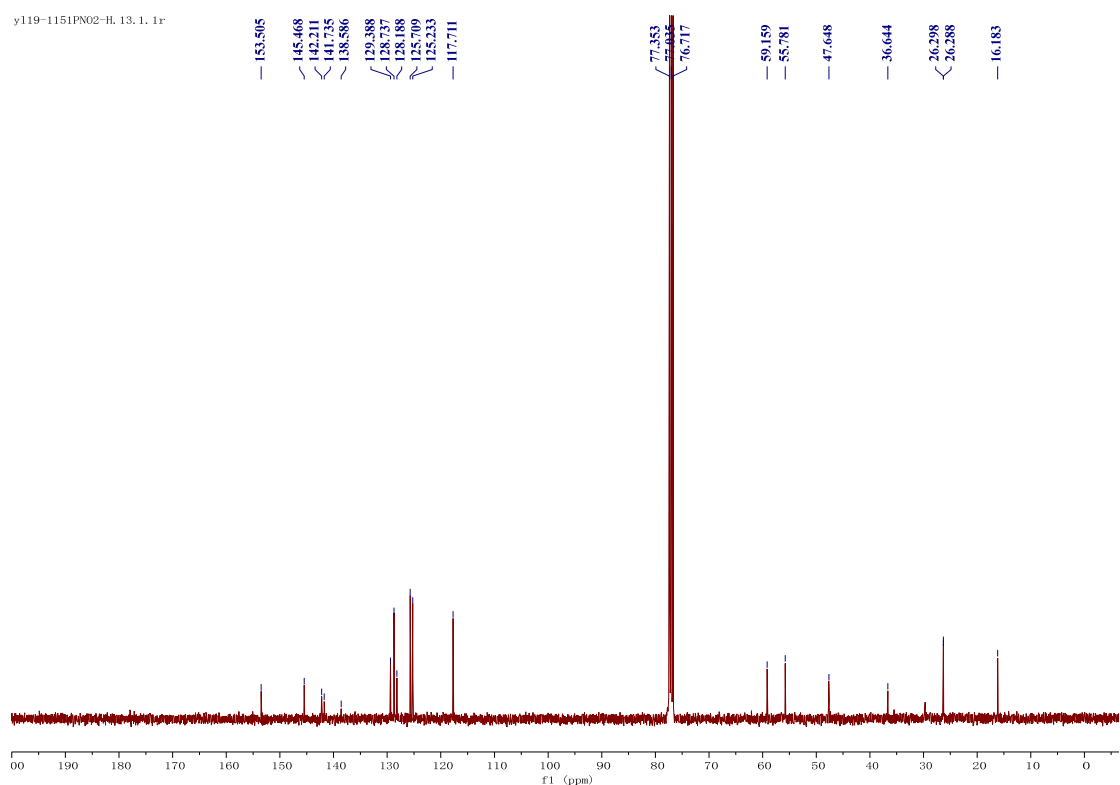




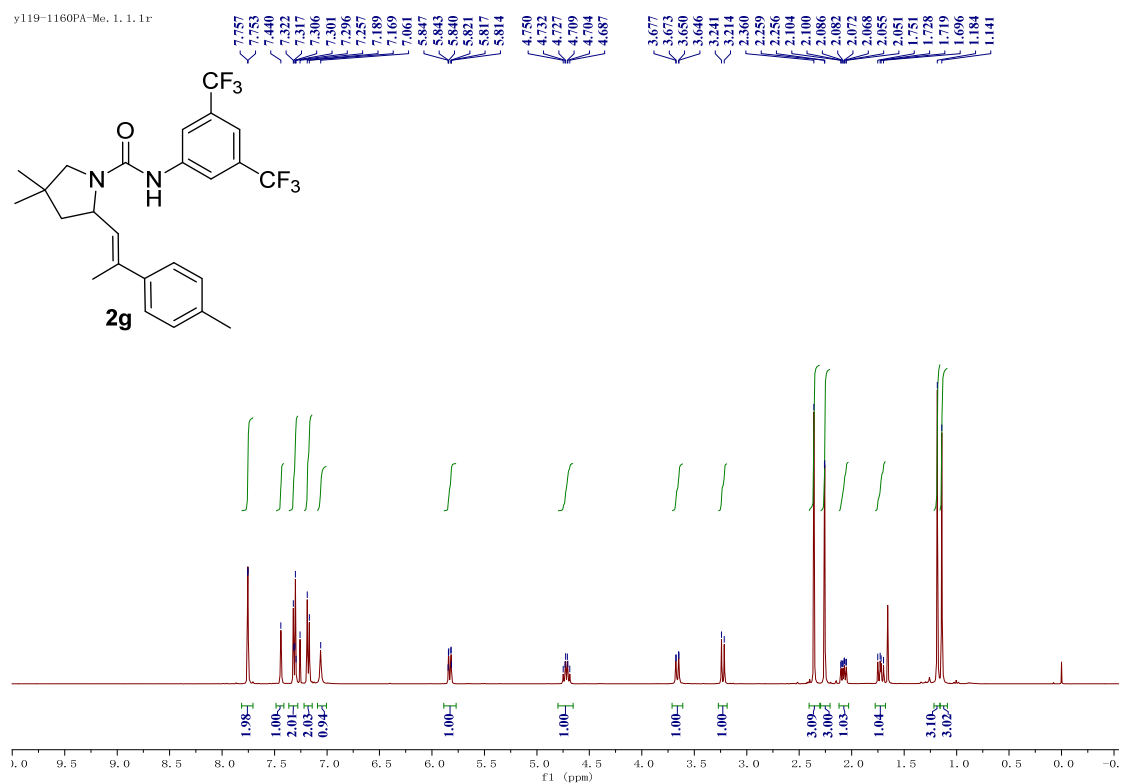
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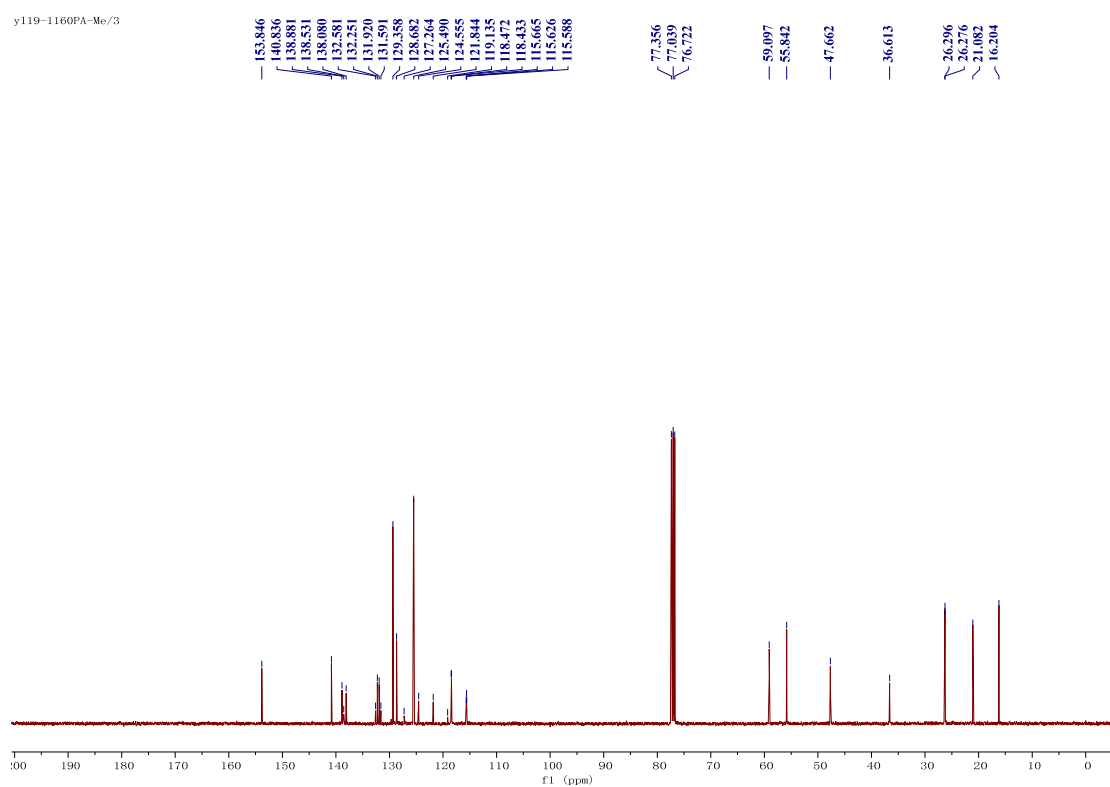
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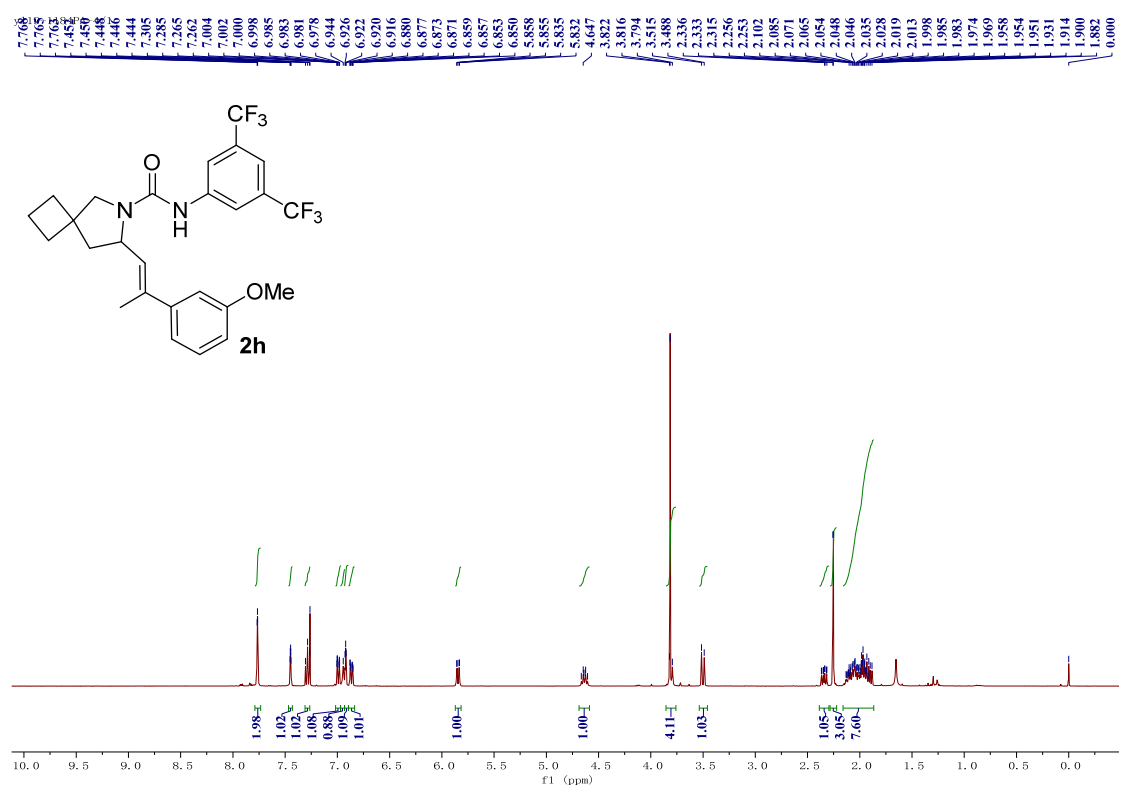
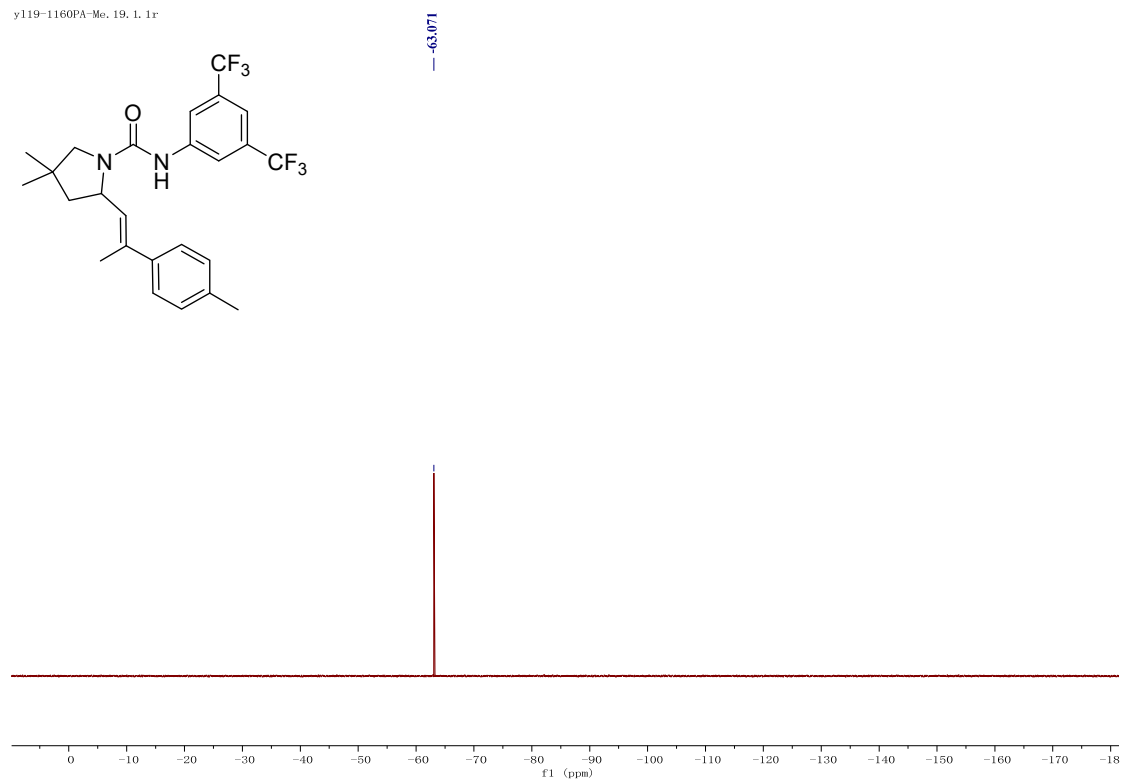
y119-1160PA-Me, 1, 1, 1r



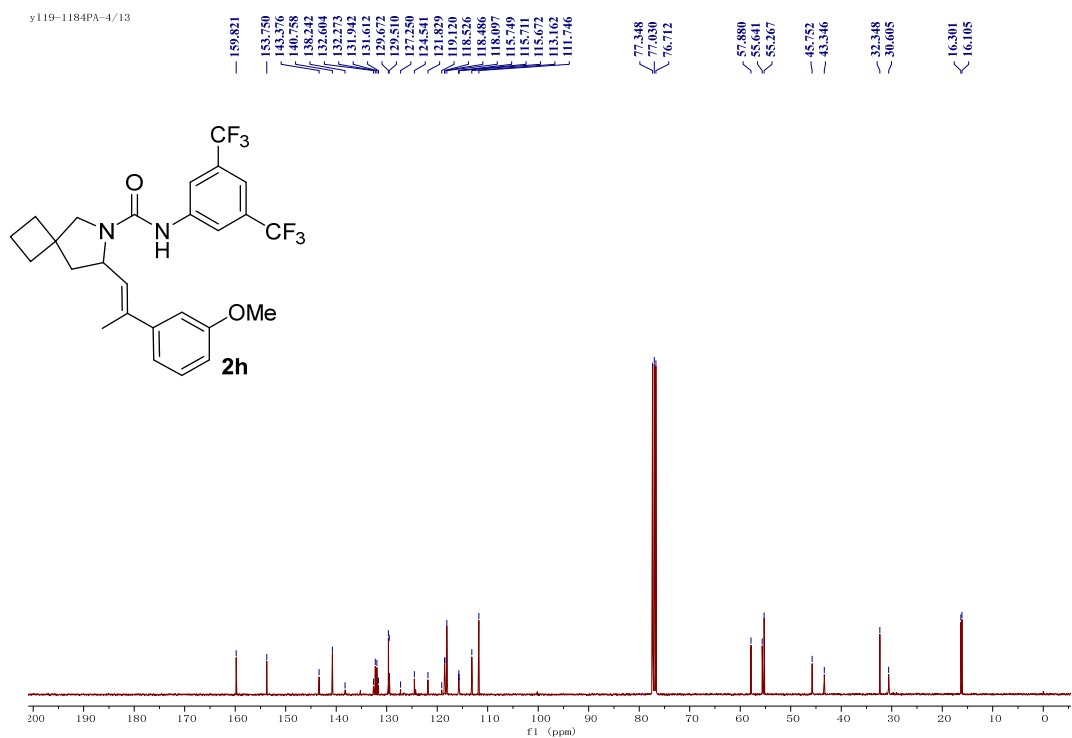
y119-1160PA-Me/3



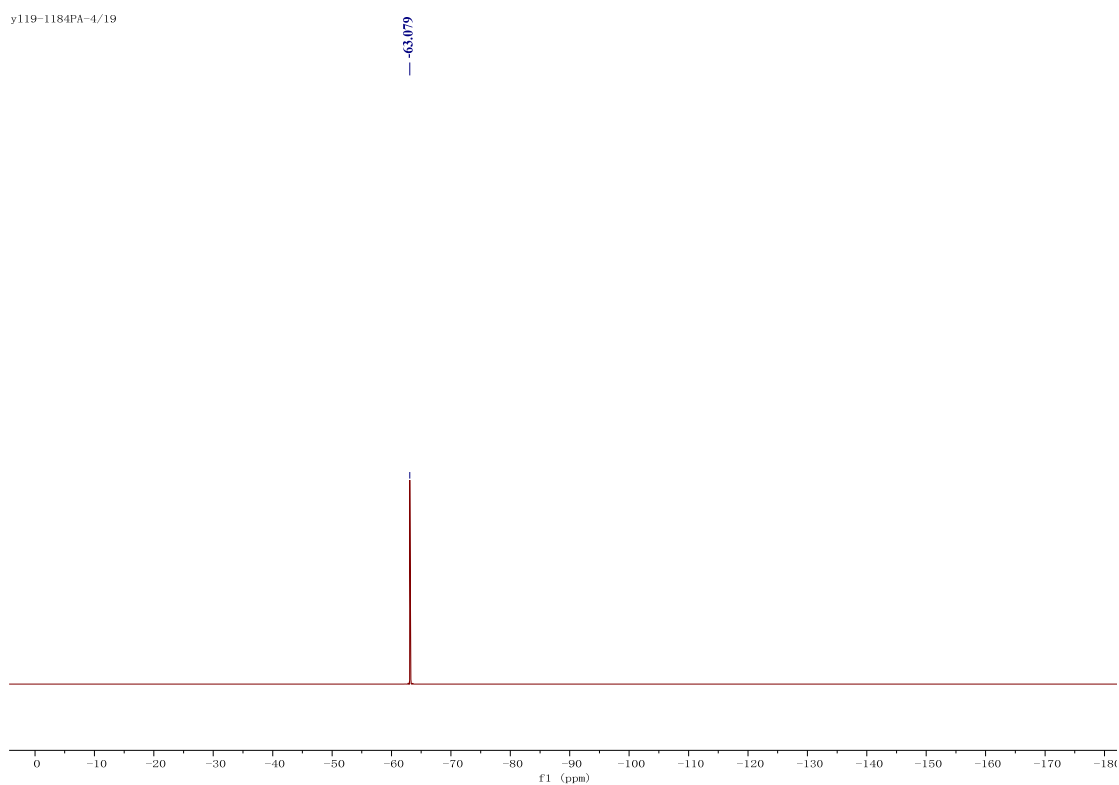
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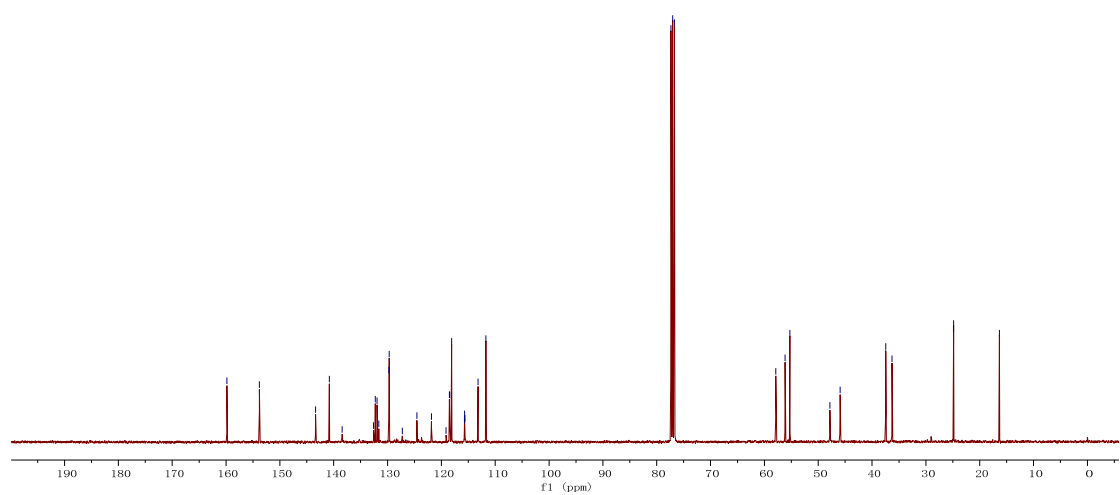
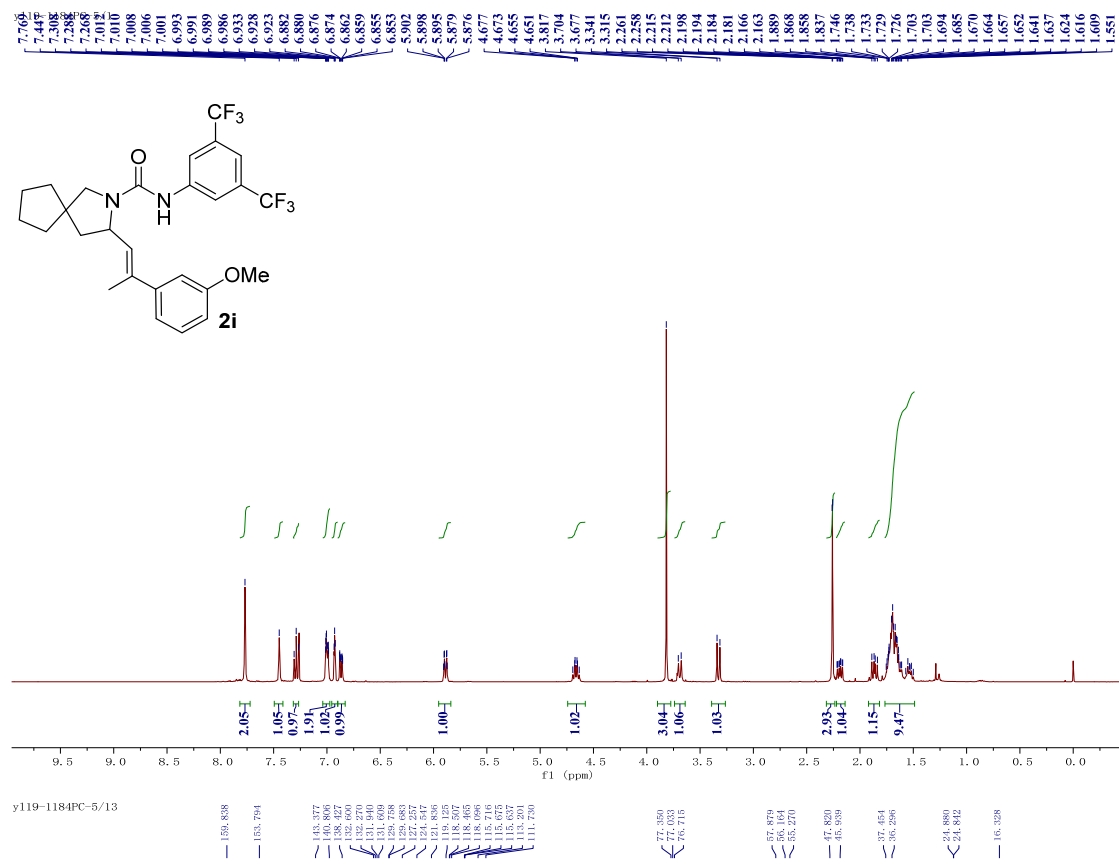


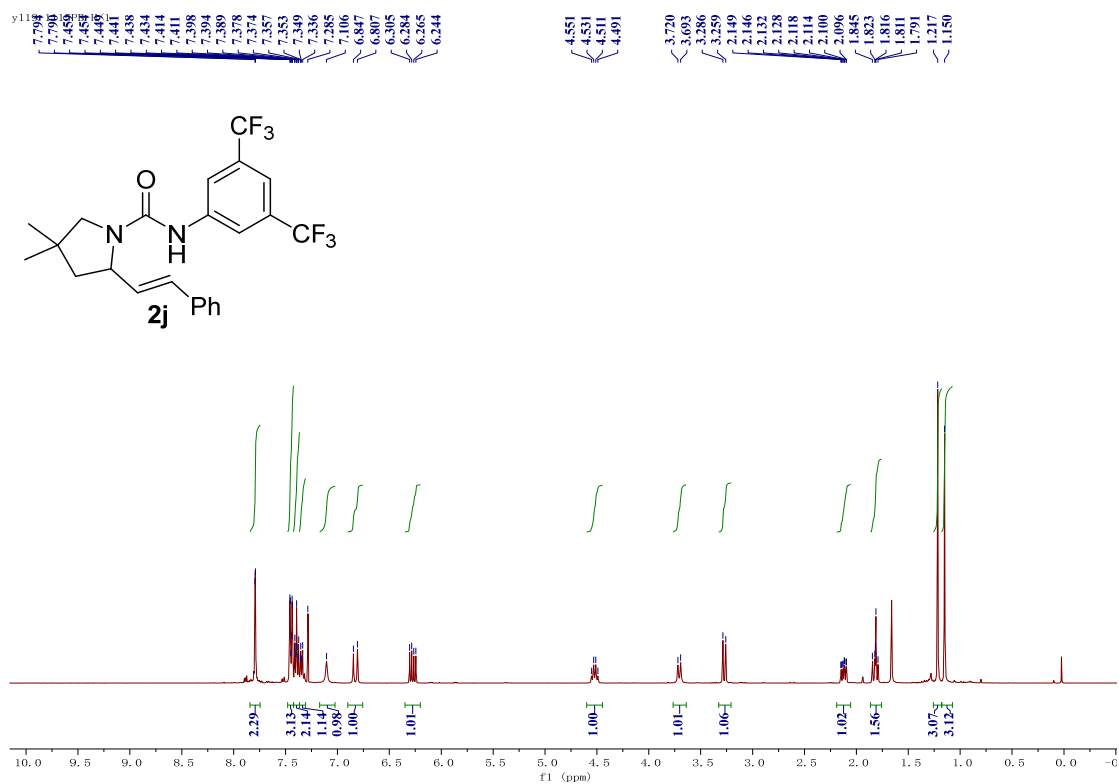
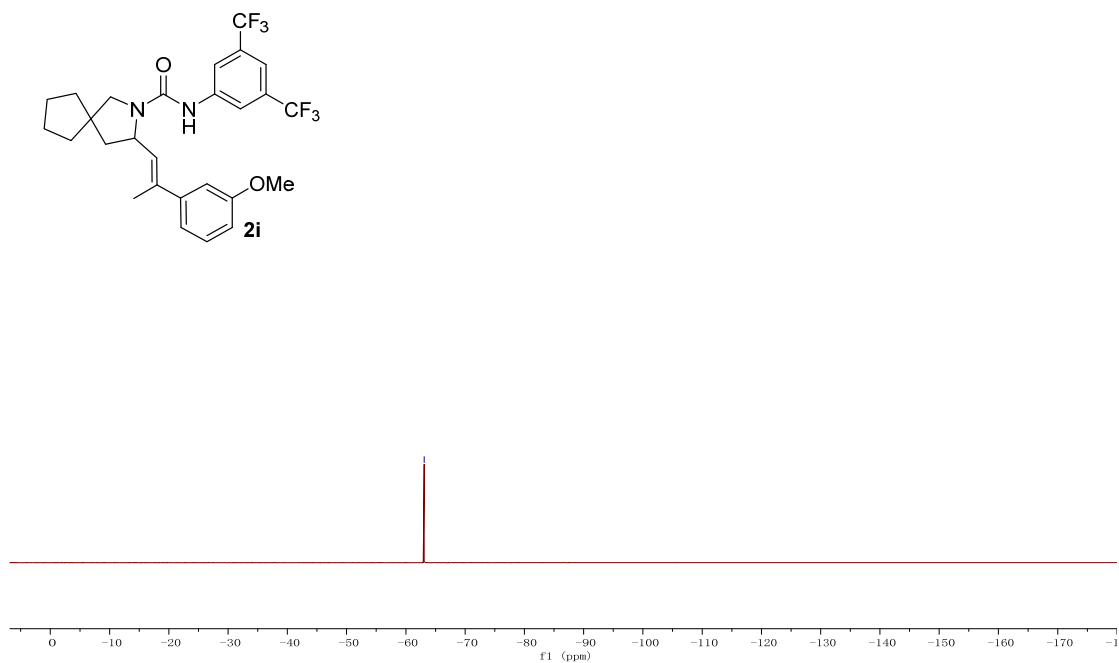
y119-1184PA-4/13



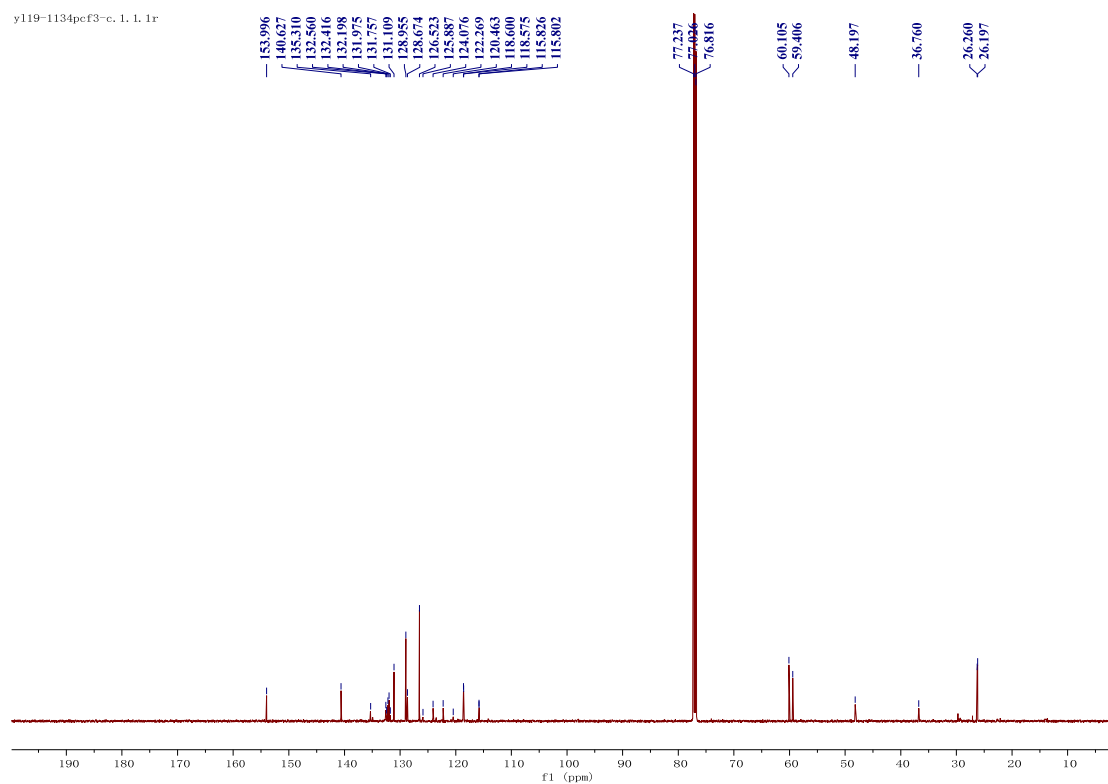
y119-1184PA-4/19



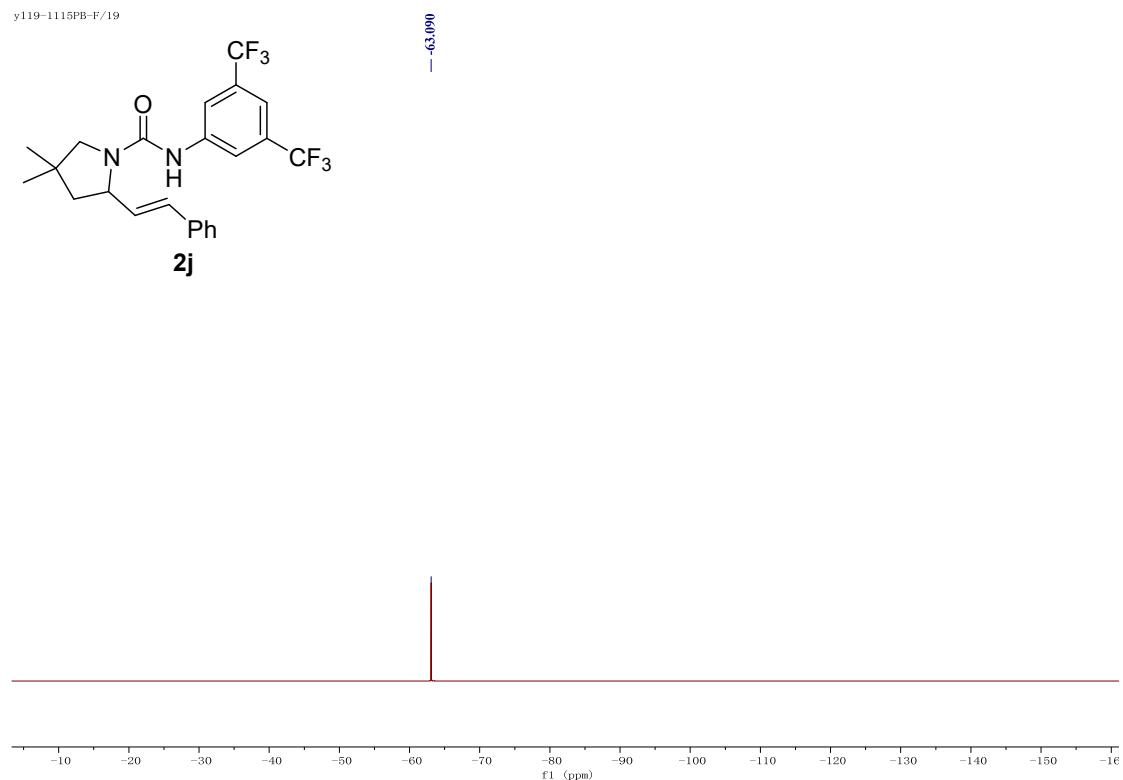
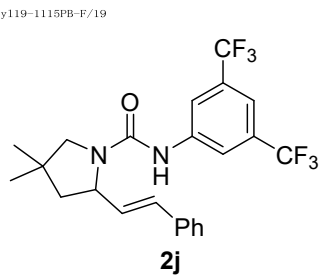


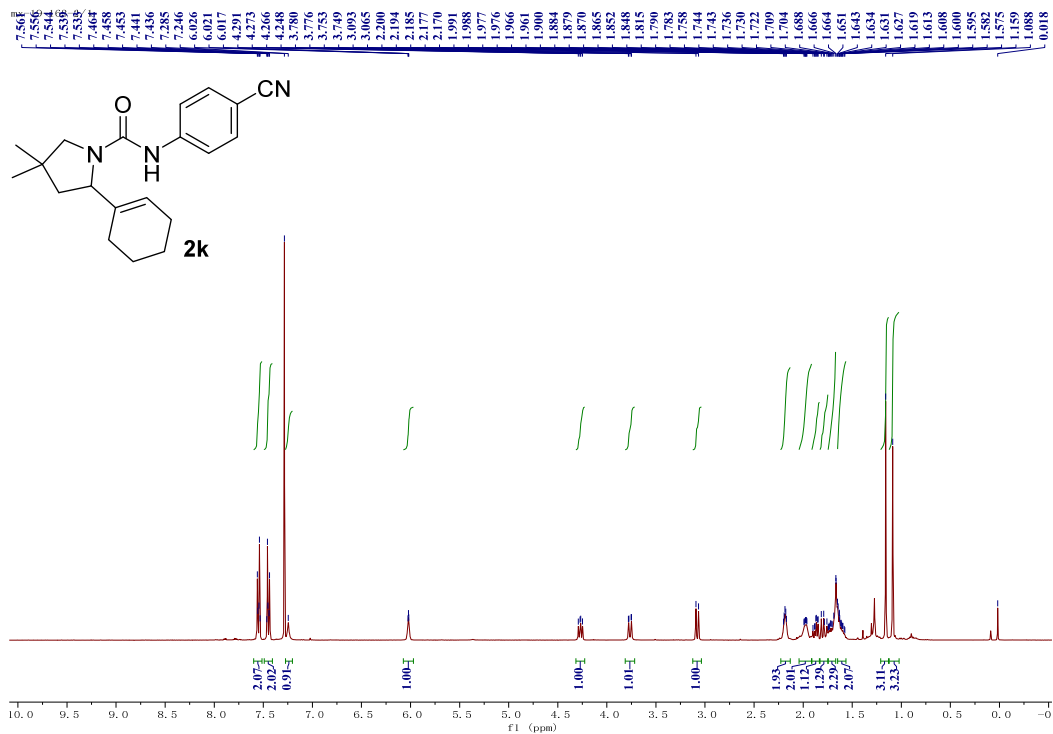
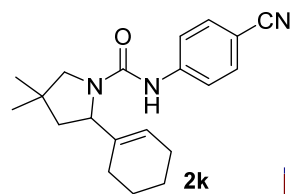


y119-1134pef3-c, 1, 1, 1r



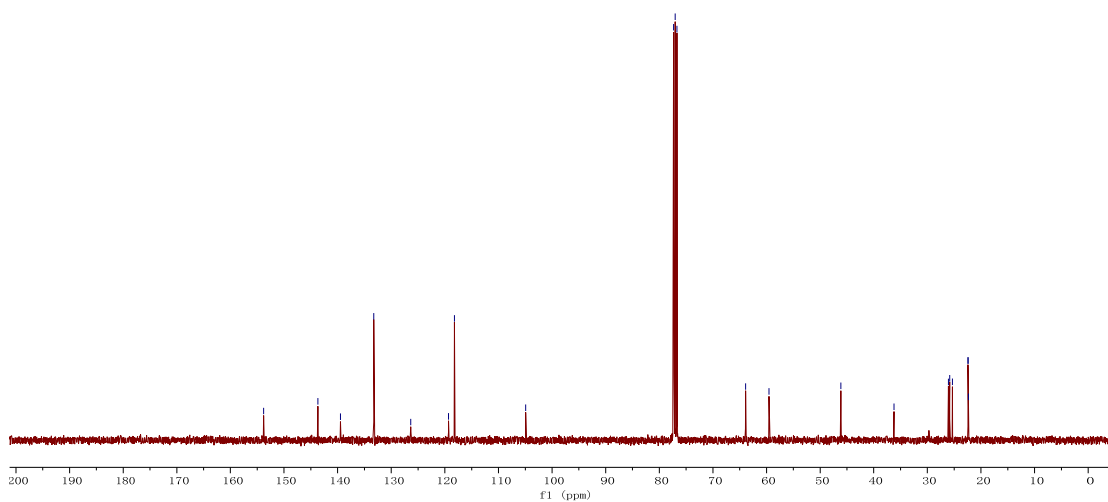
y119-1115PB-F/19



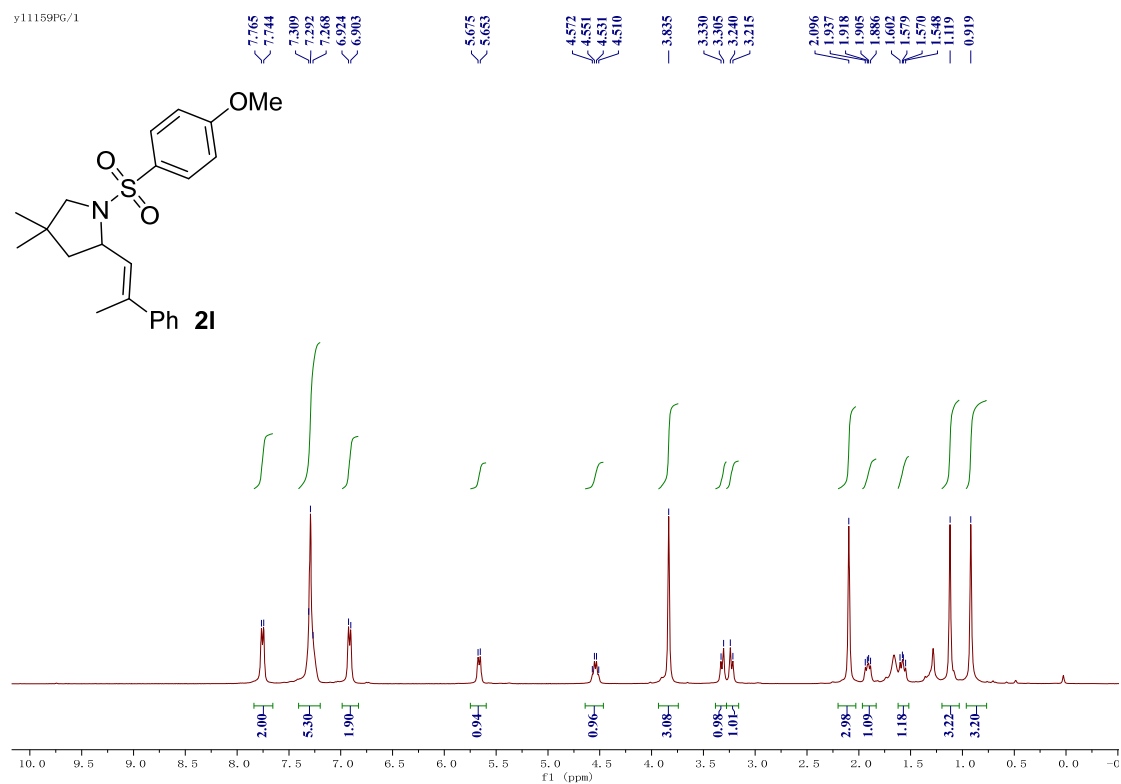


mx-19-163-2/2

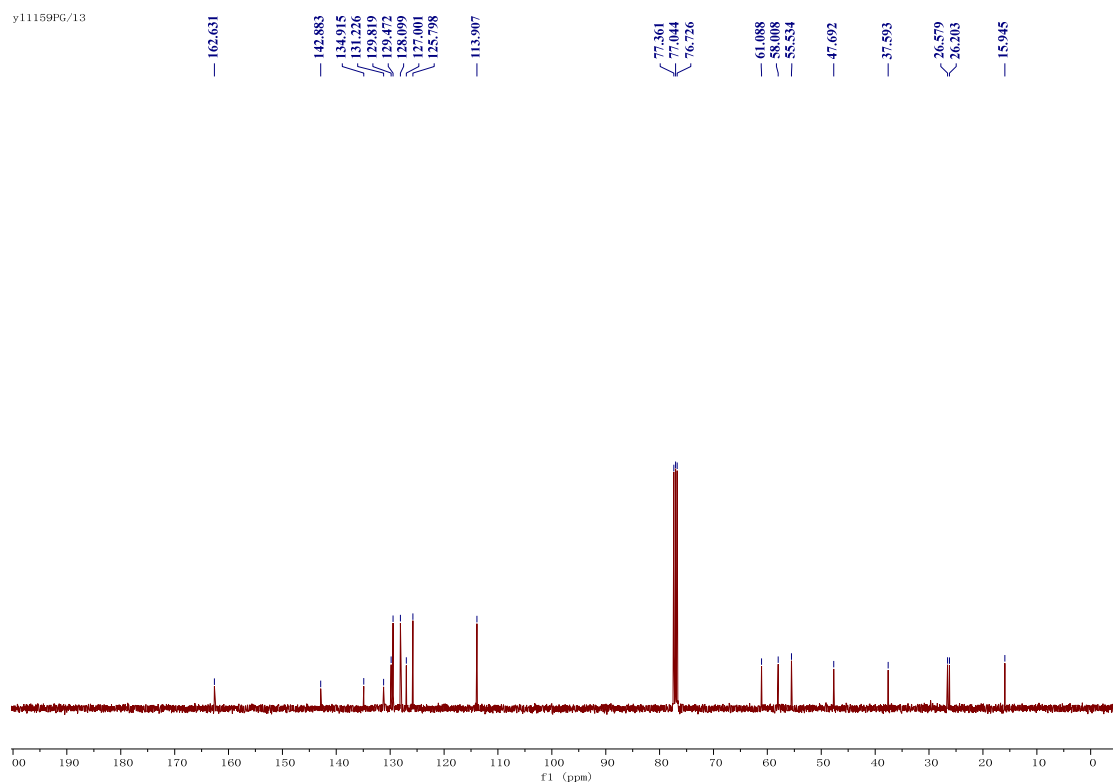
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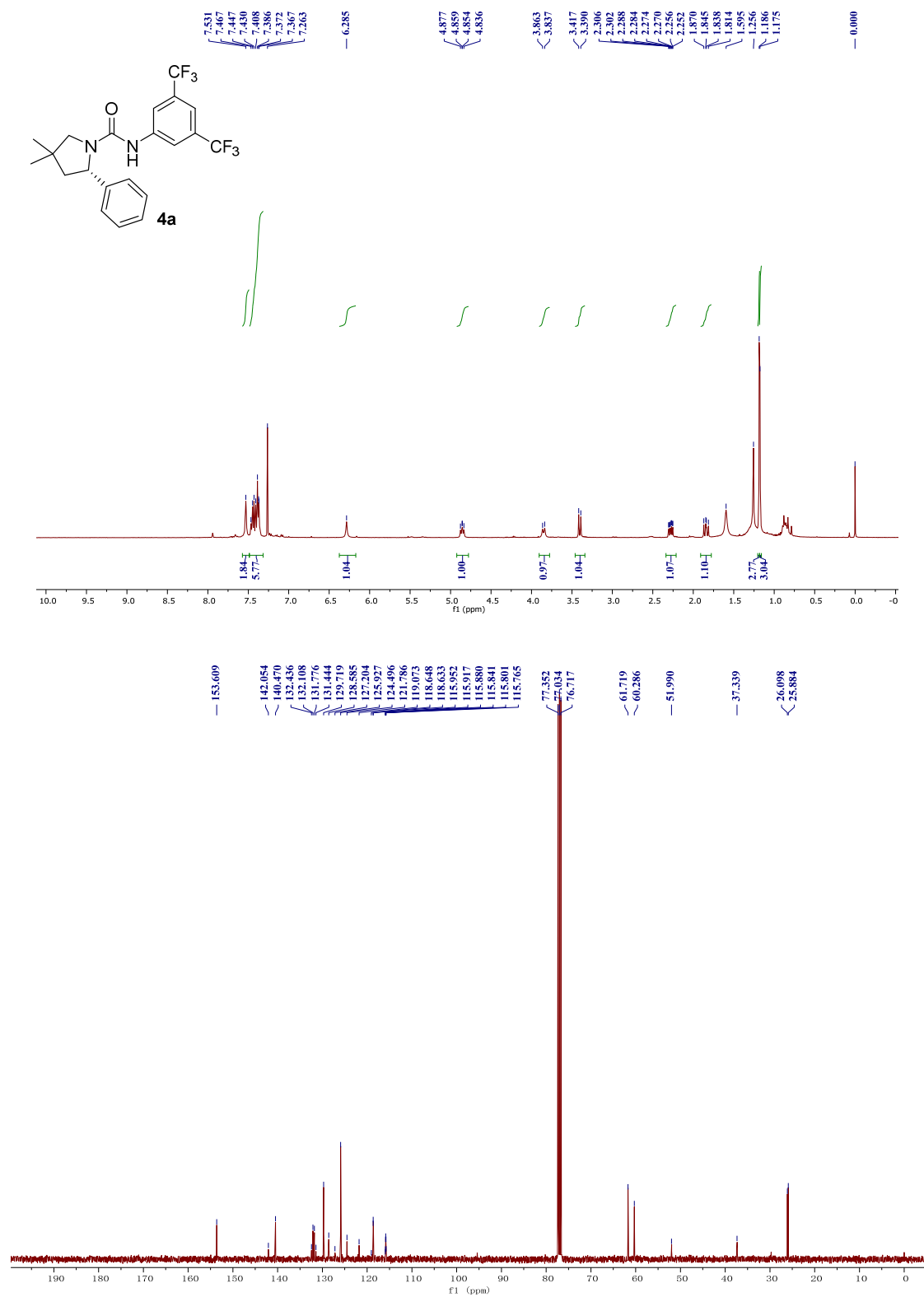


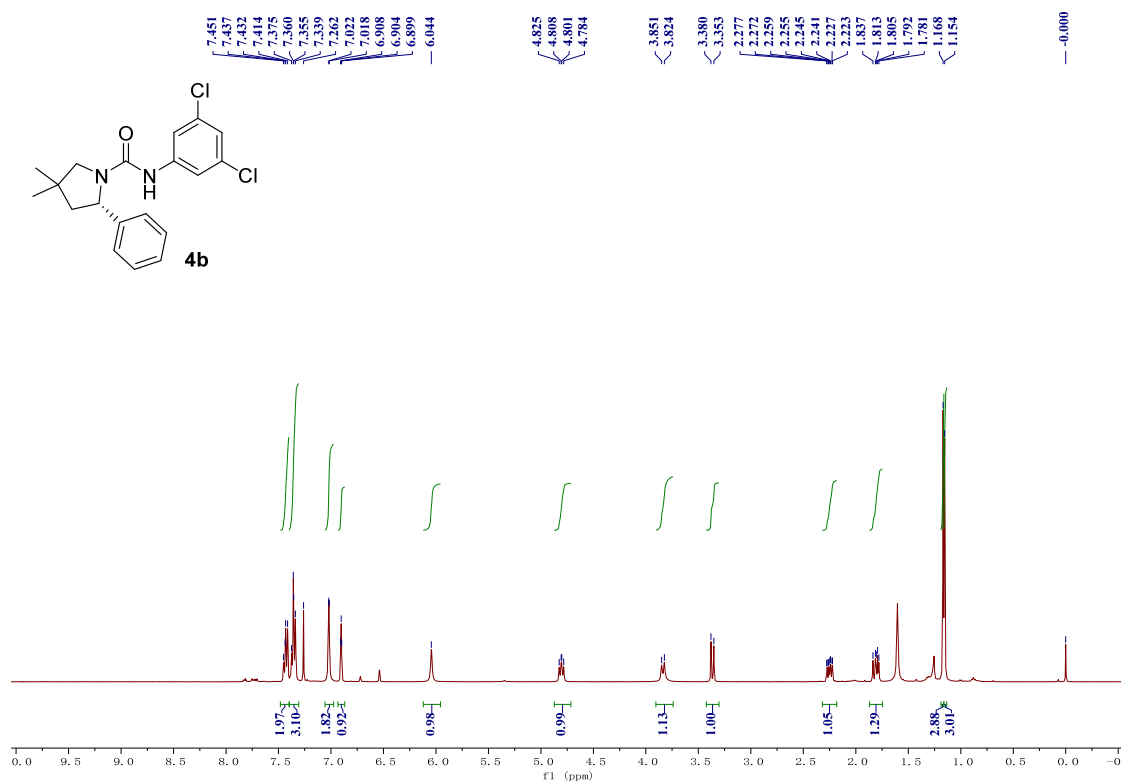
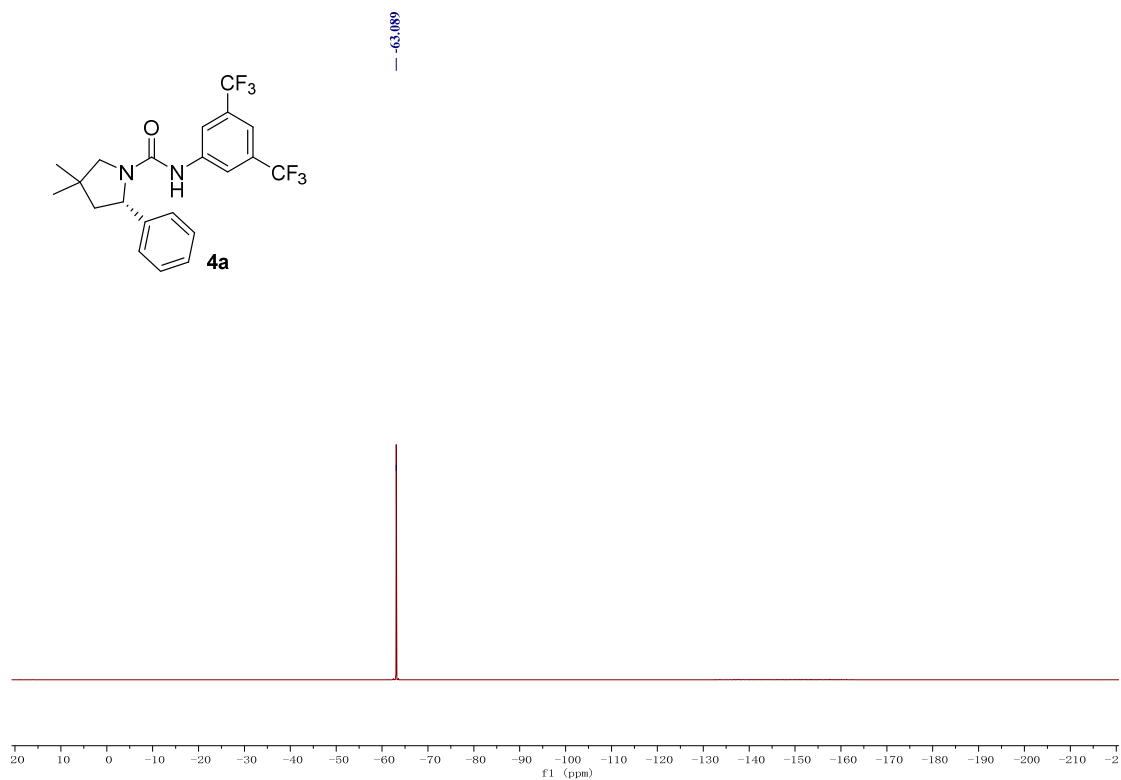
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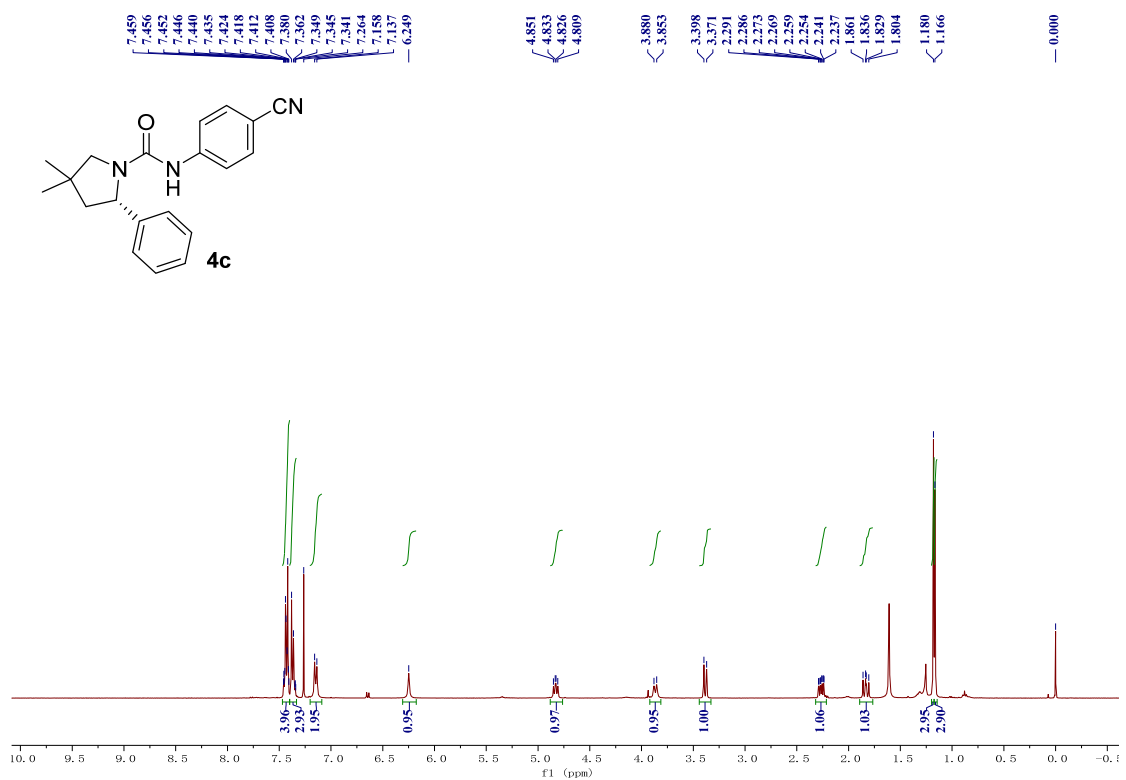
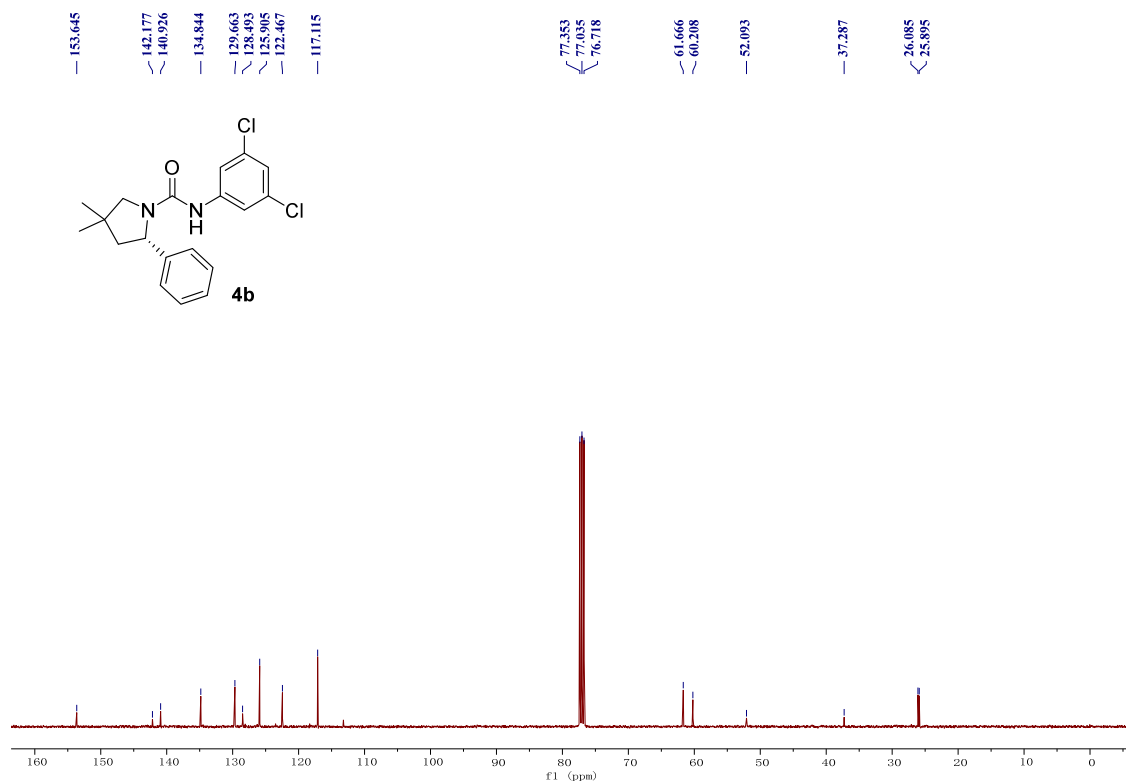


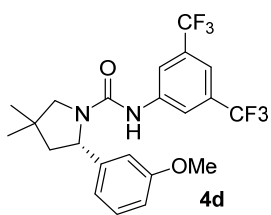
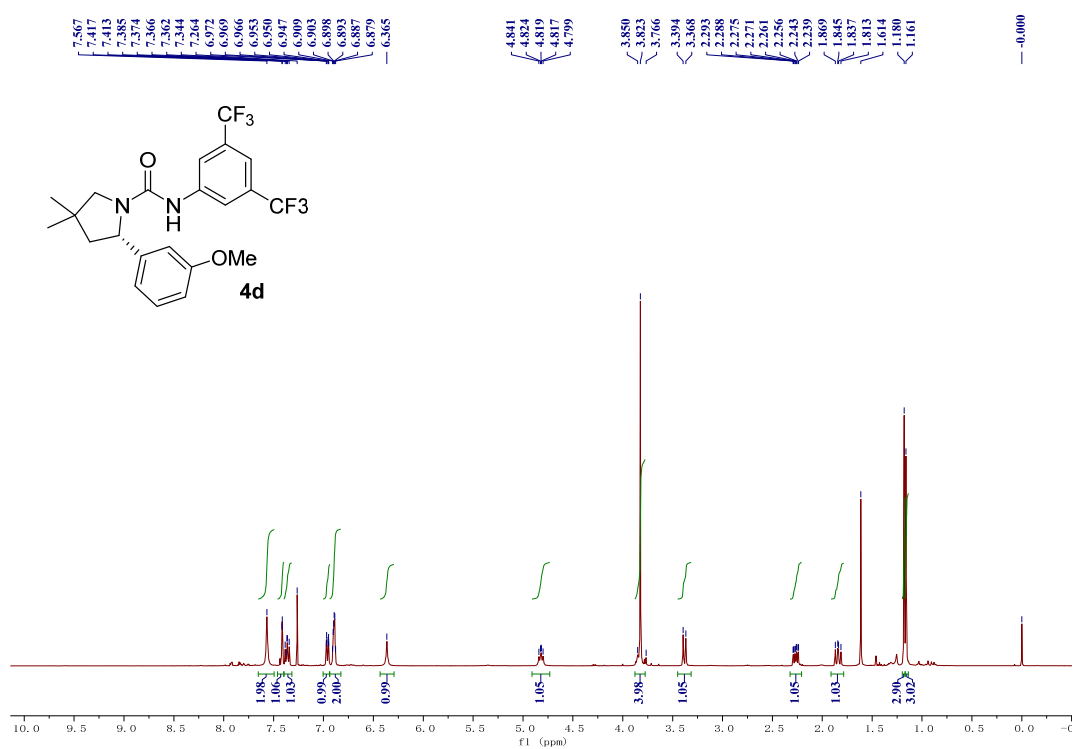
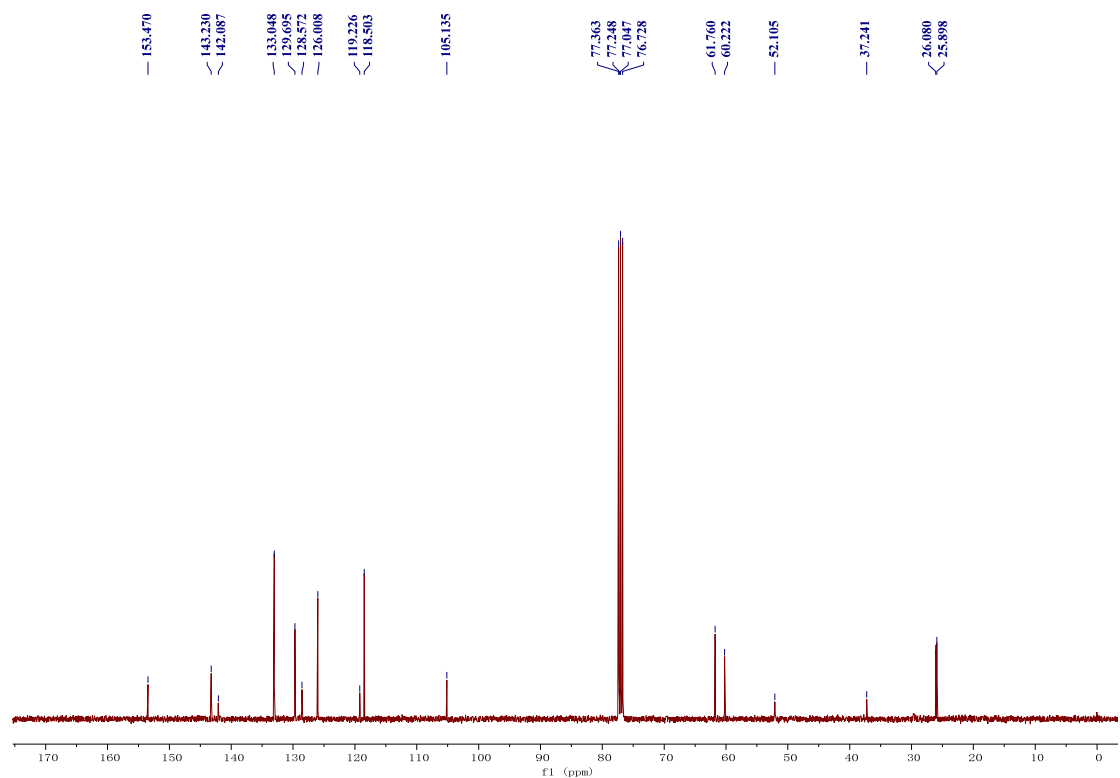
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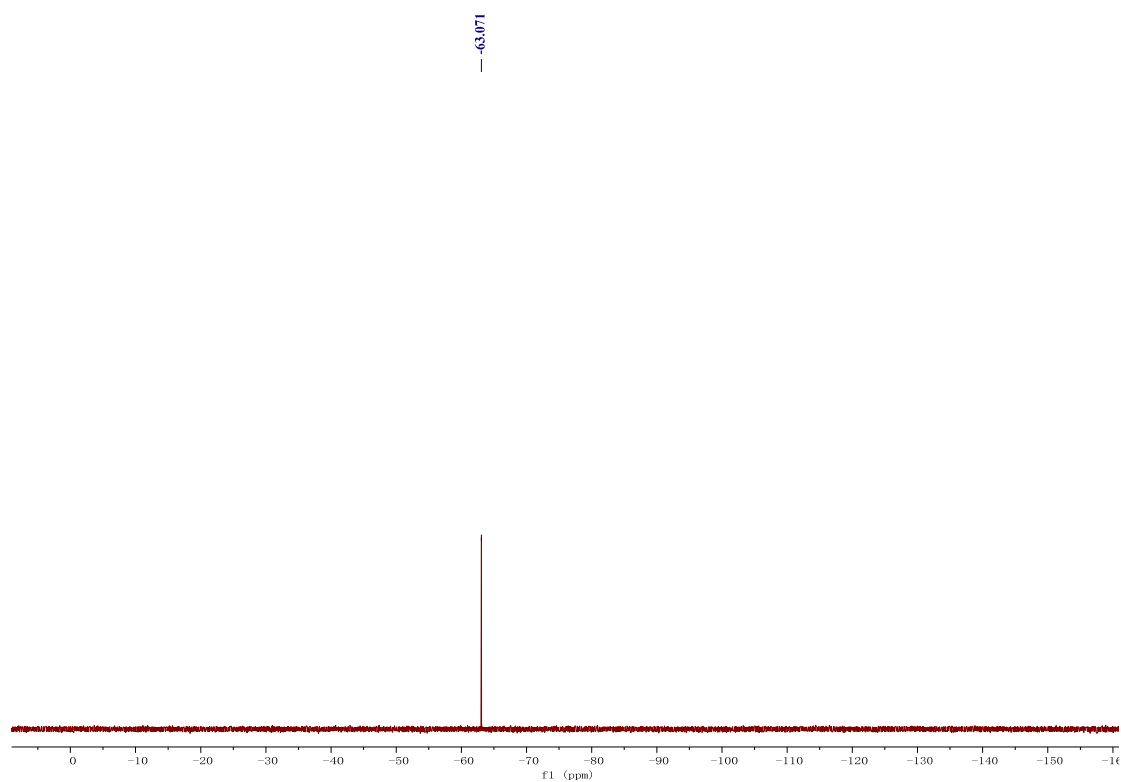
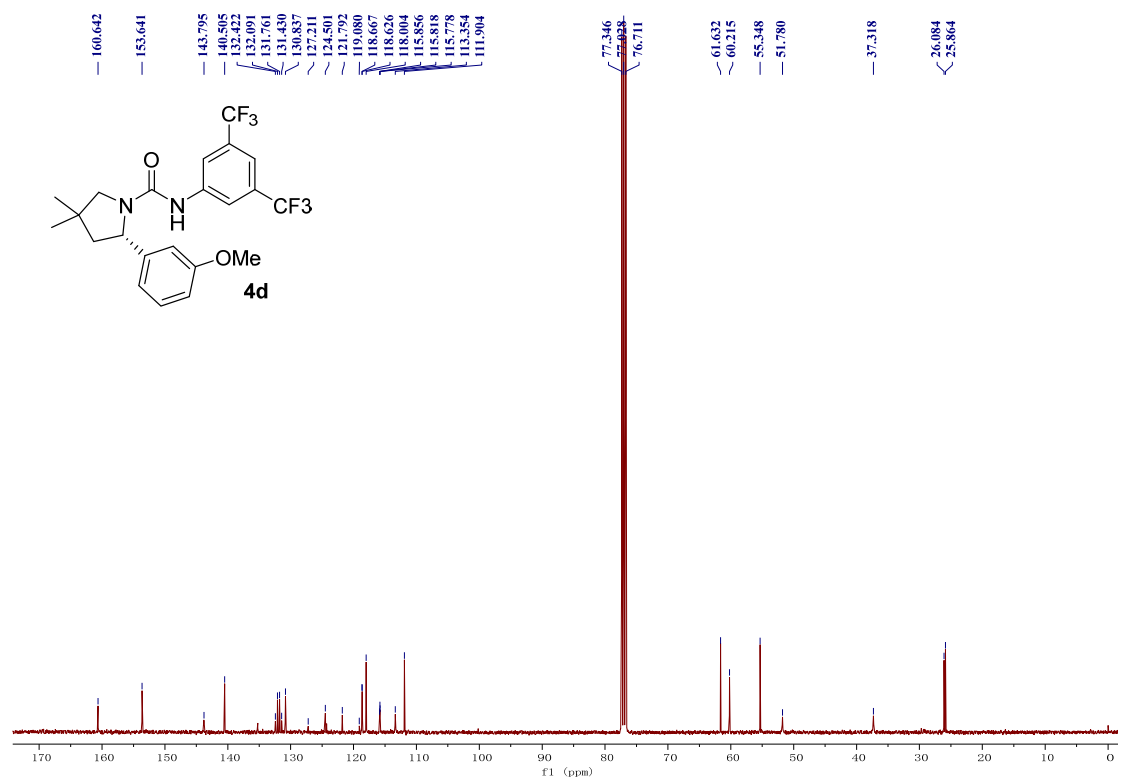


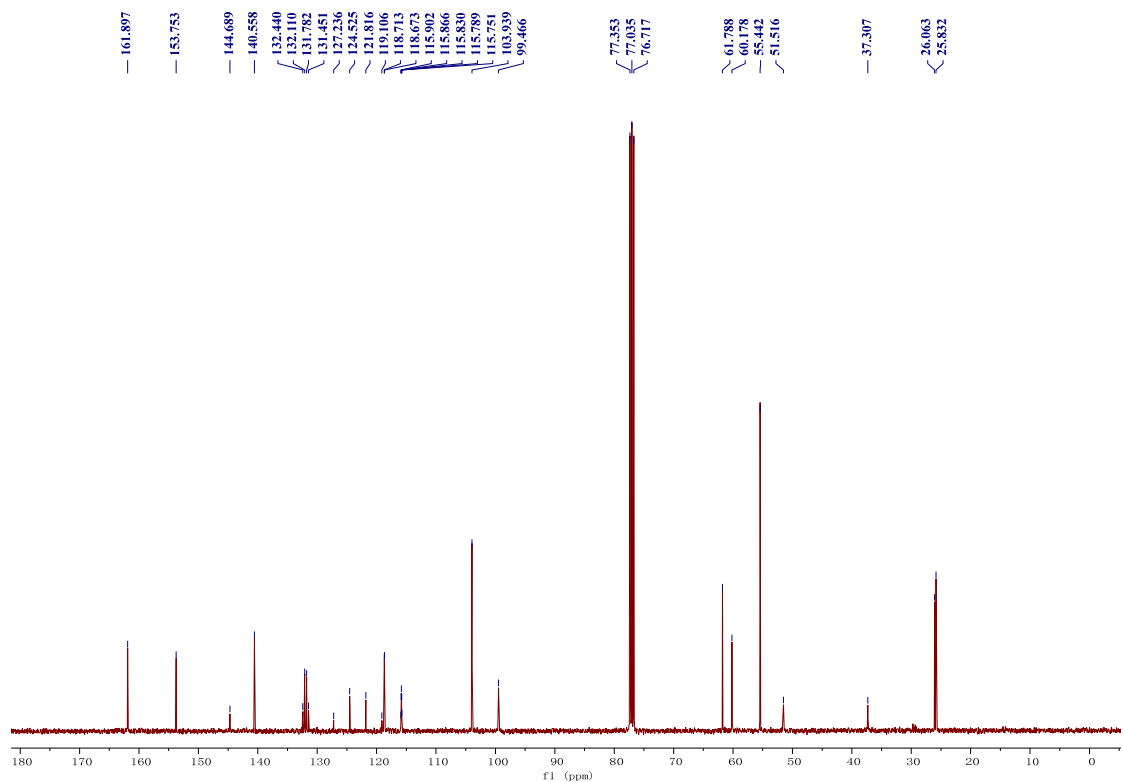
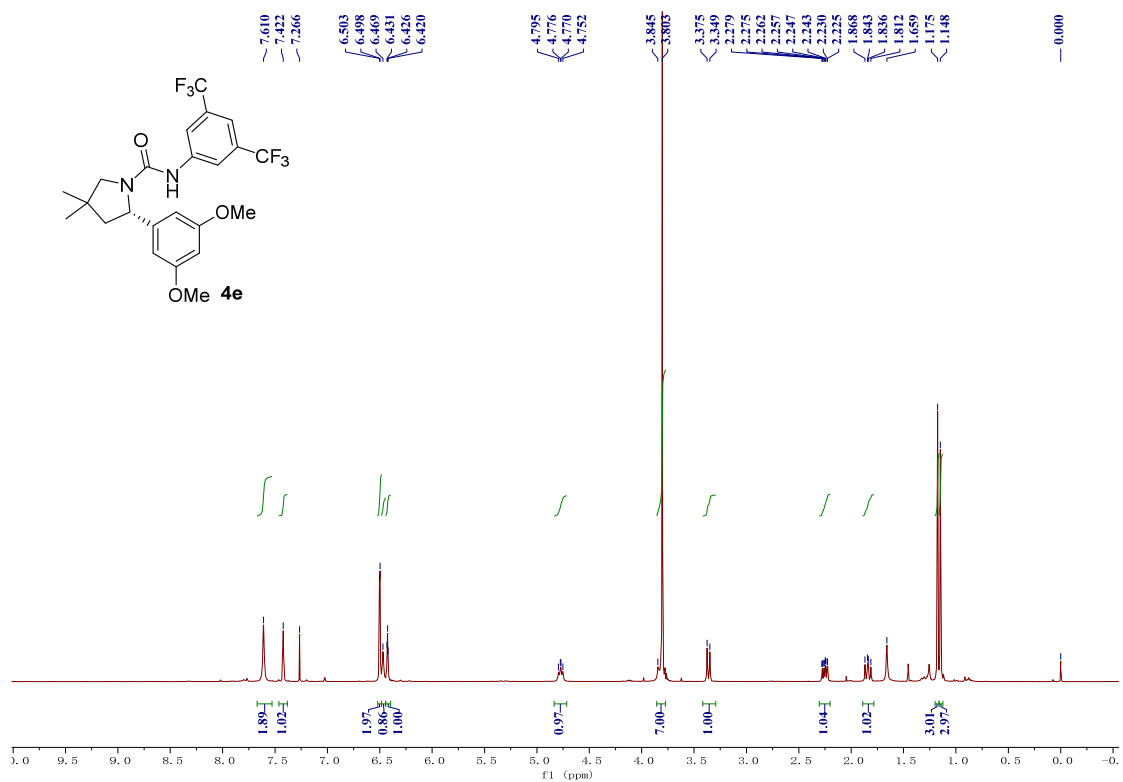


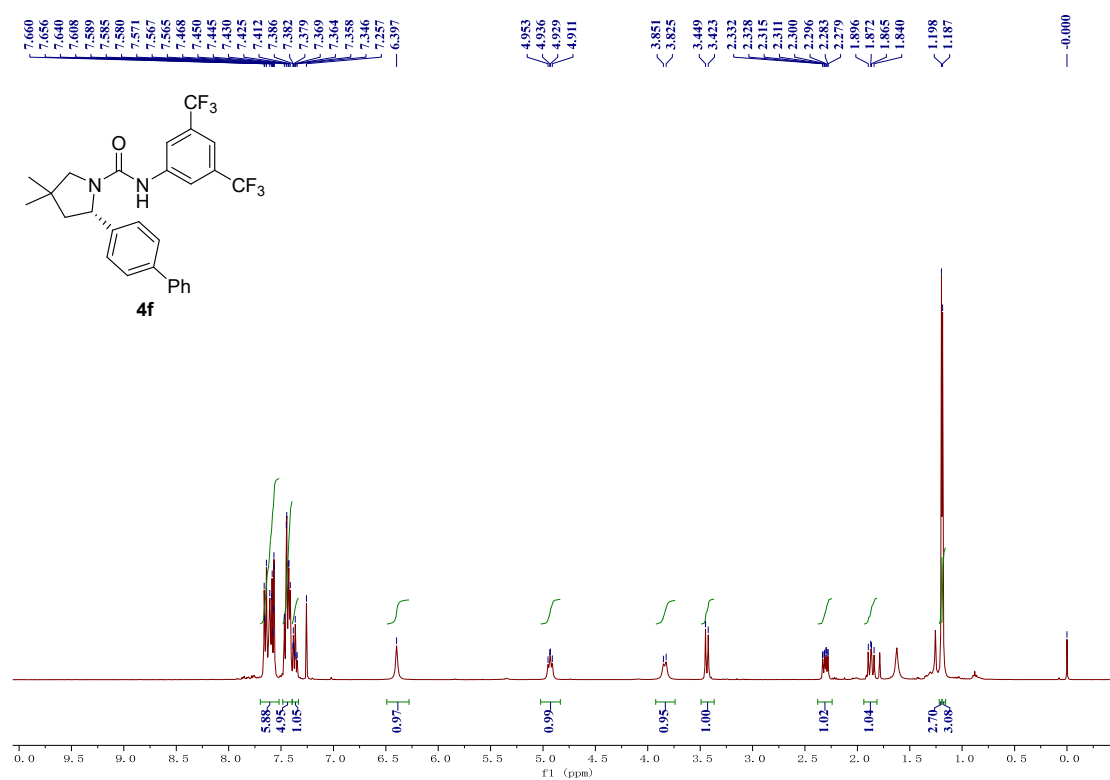
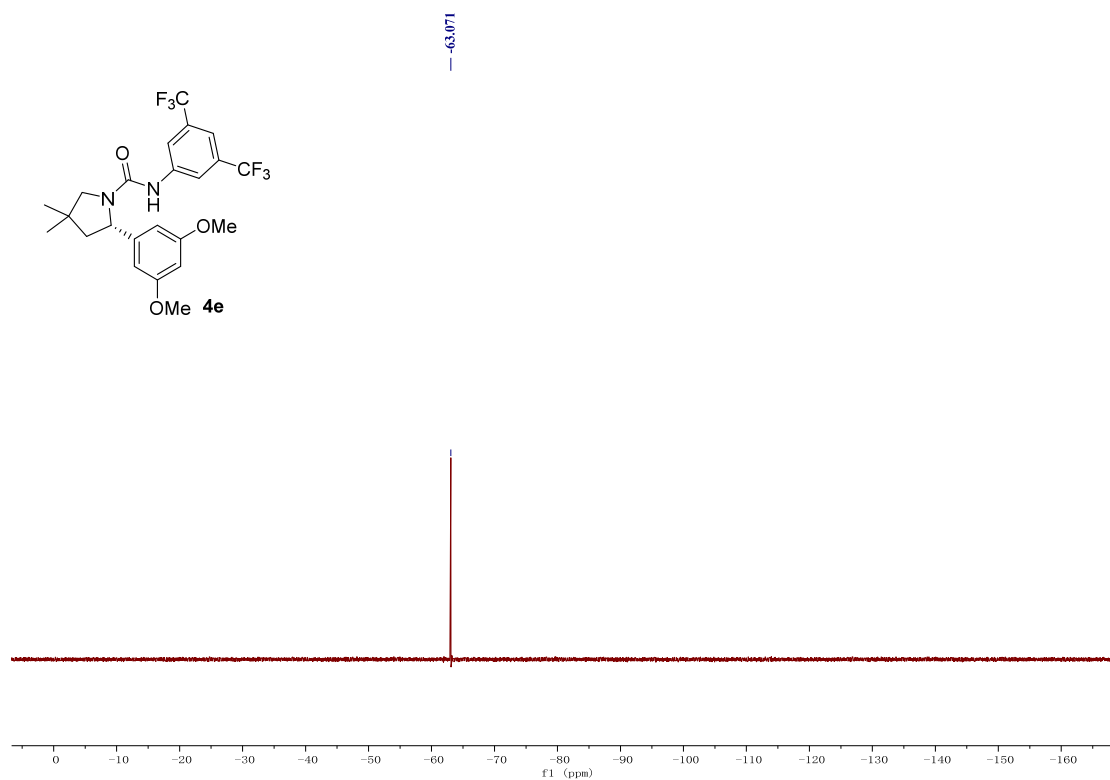


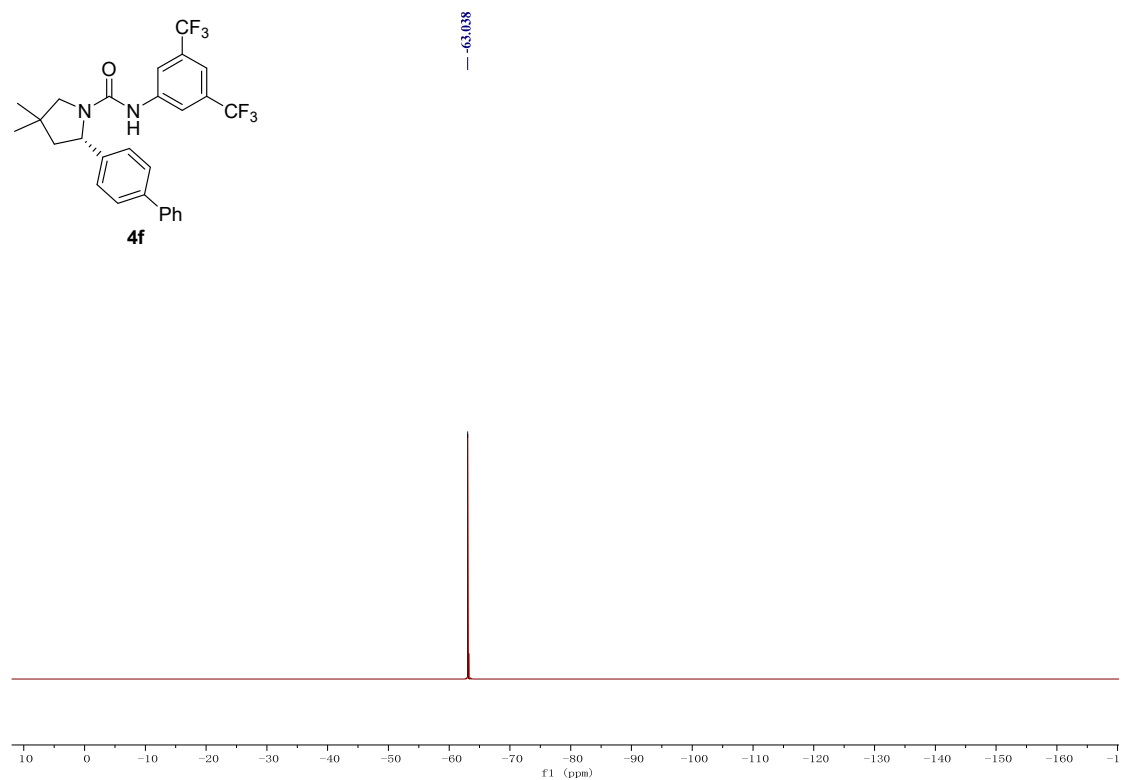
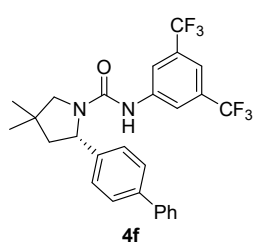
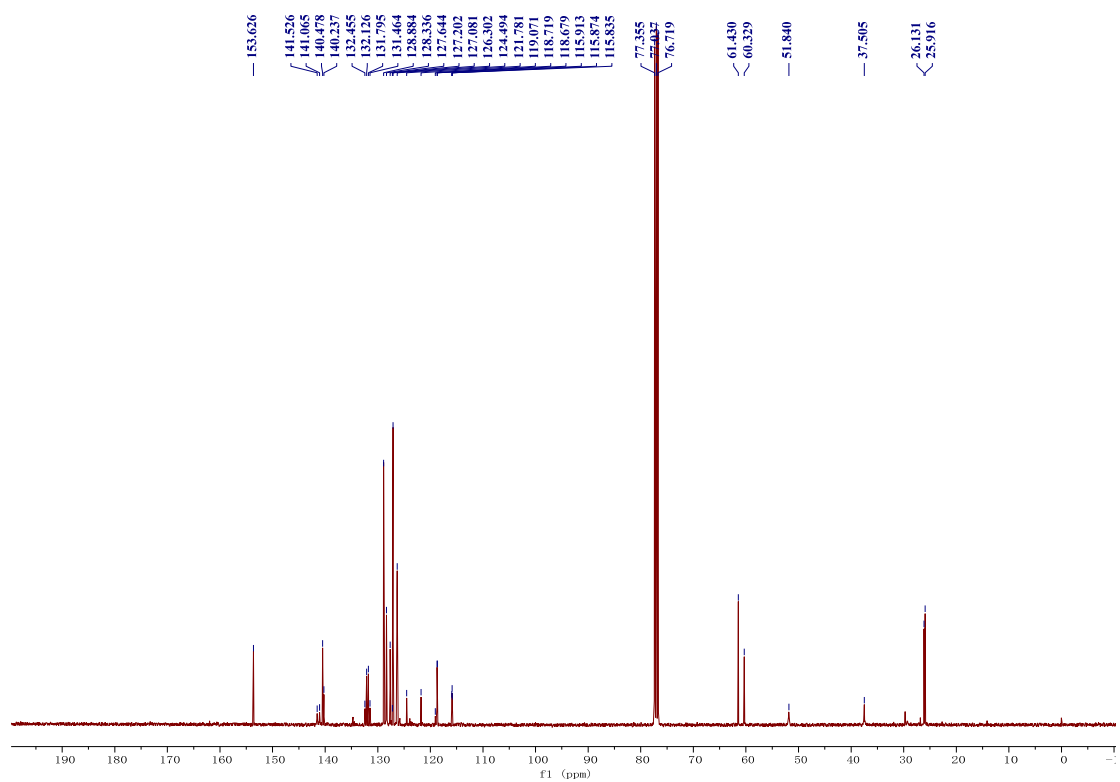


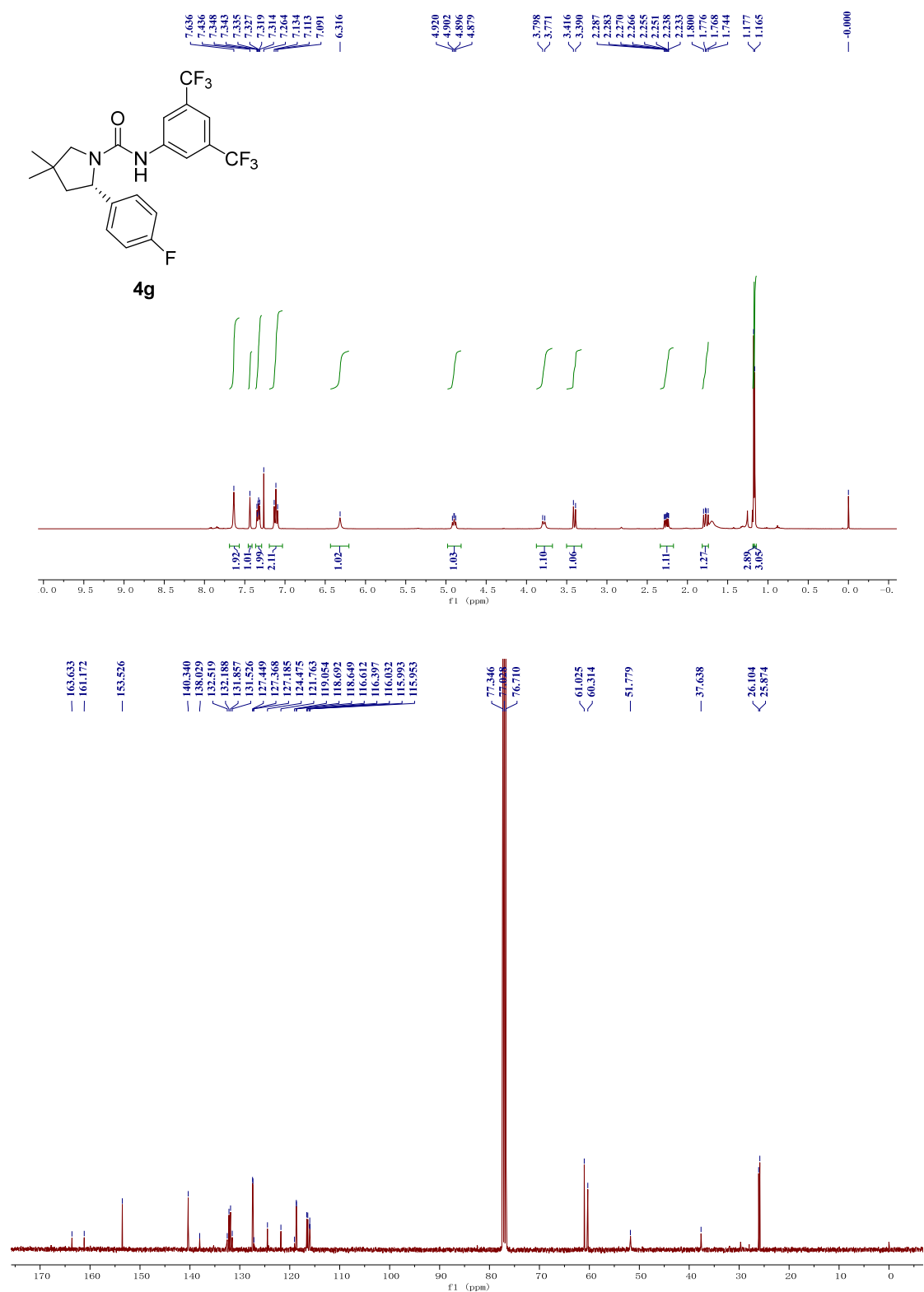




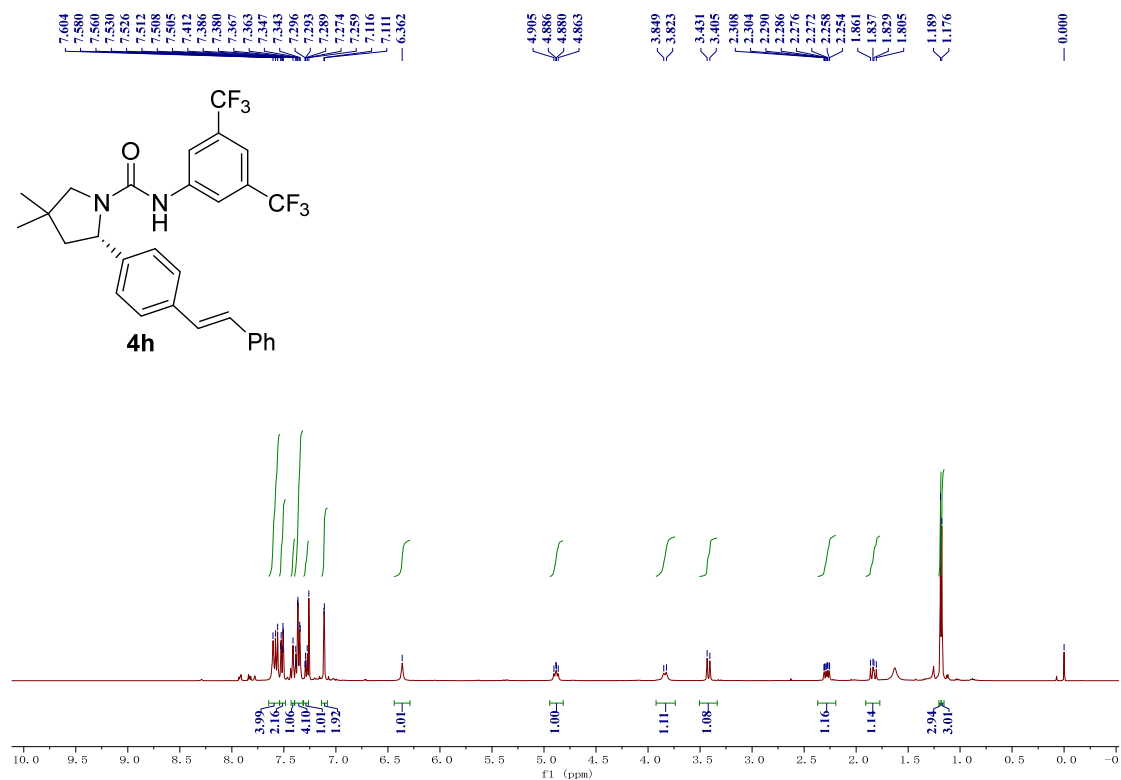
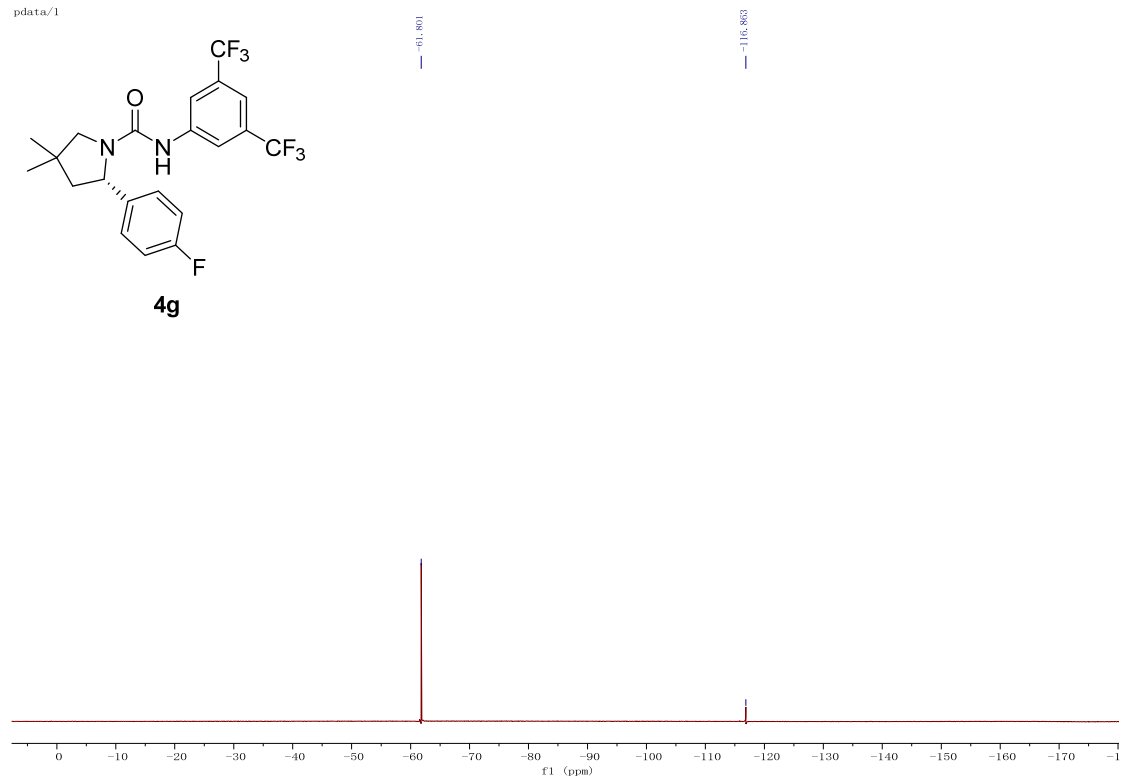
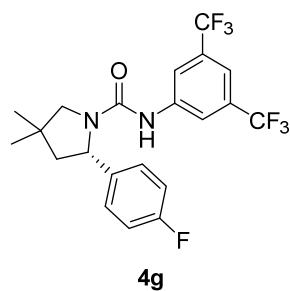


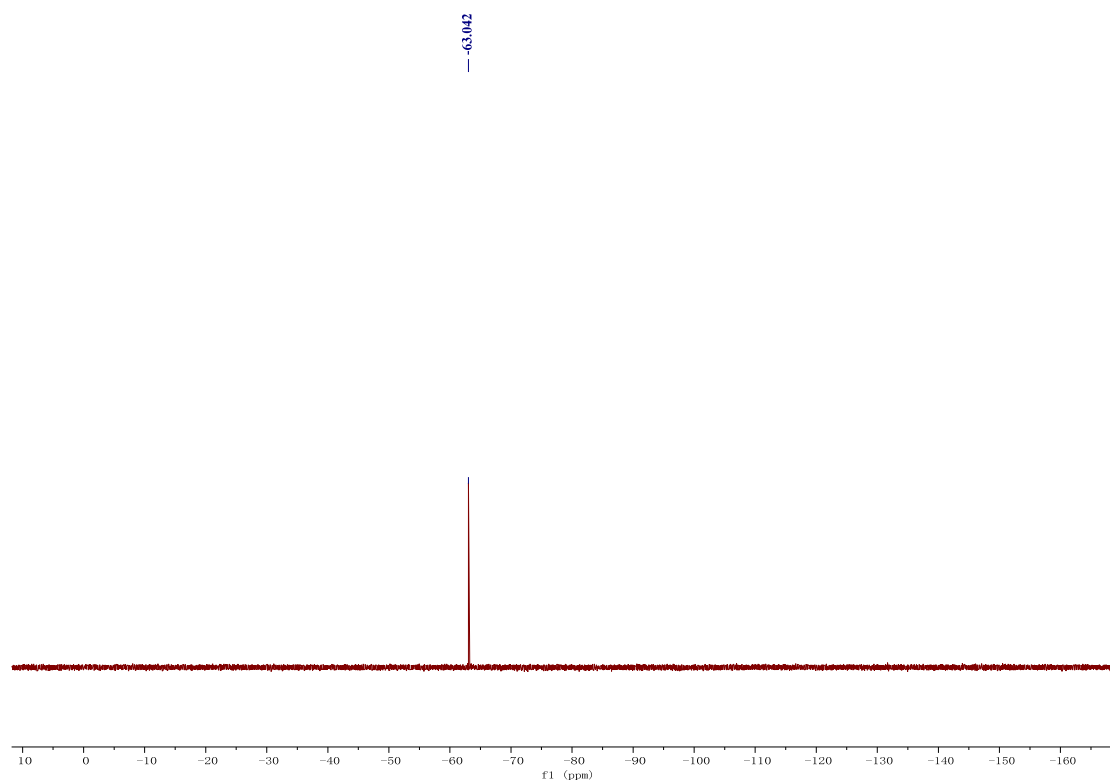
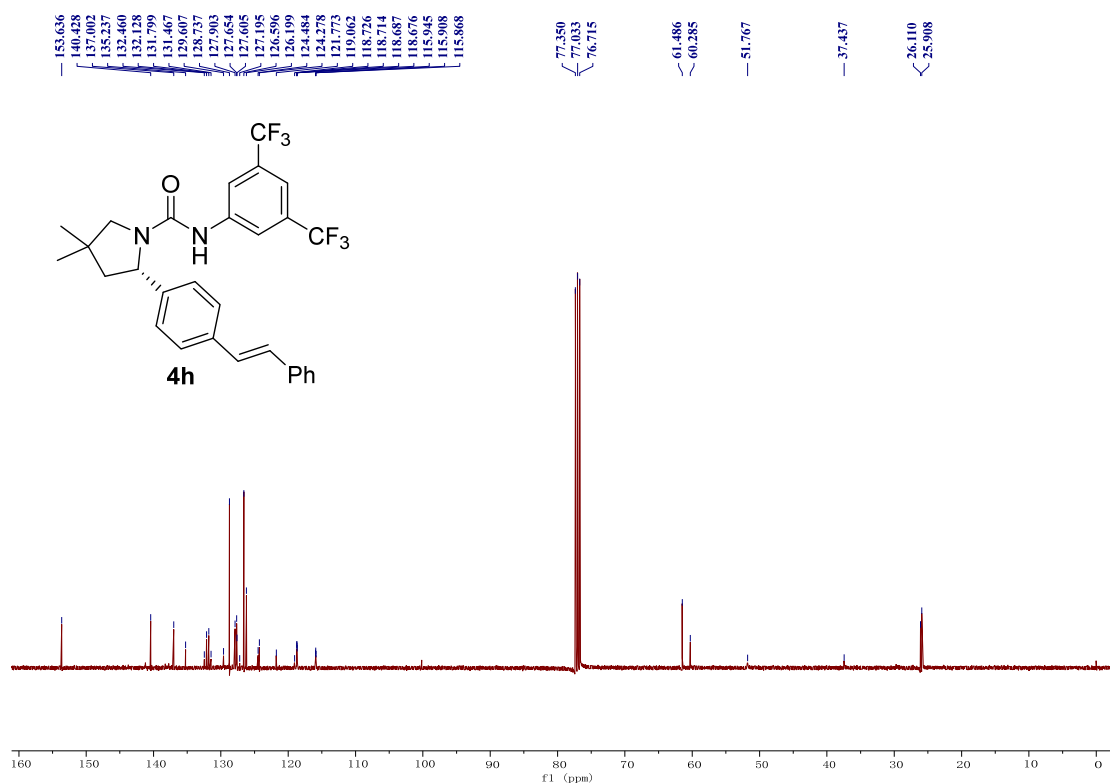


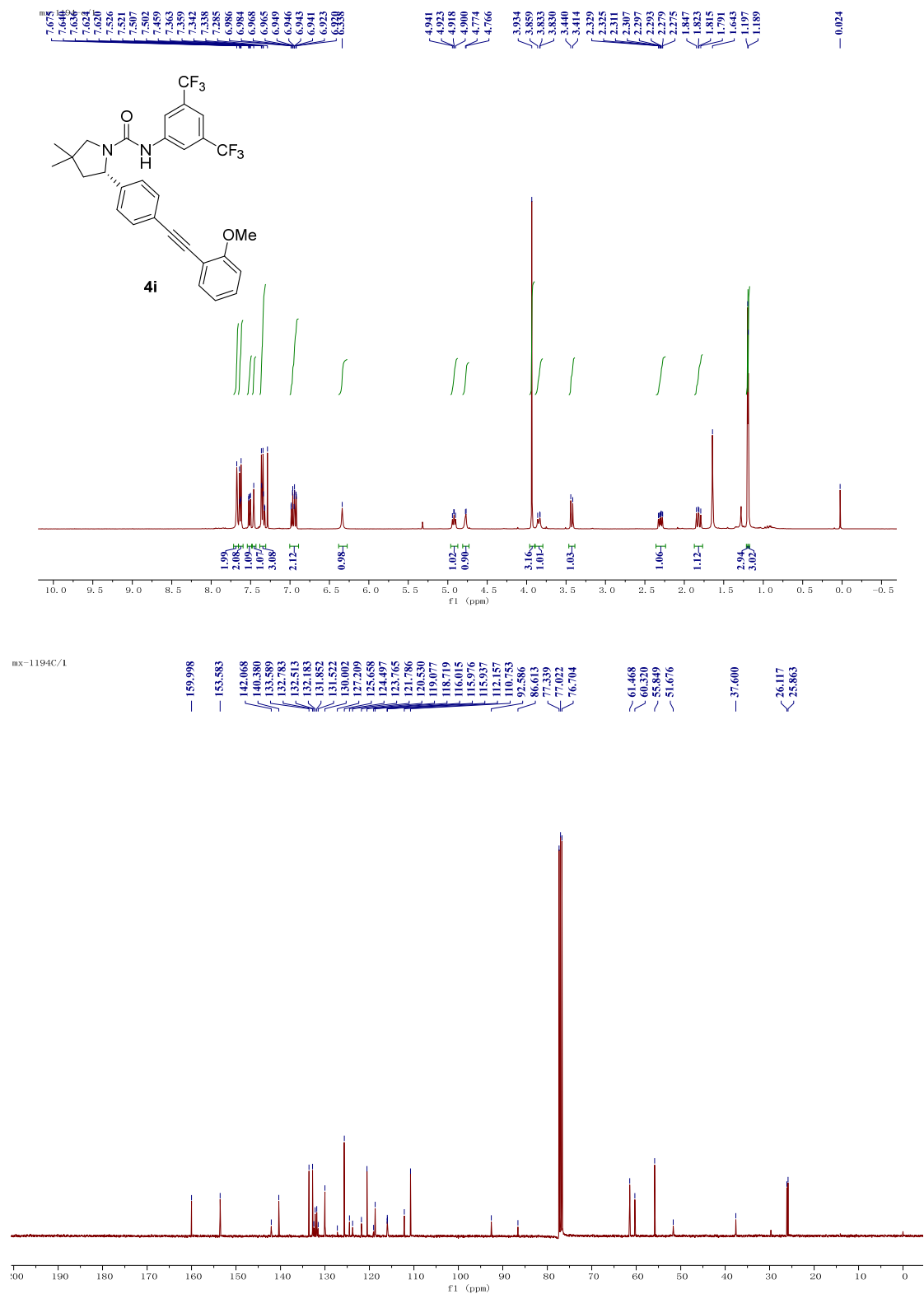


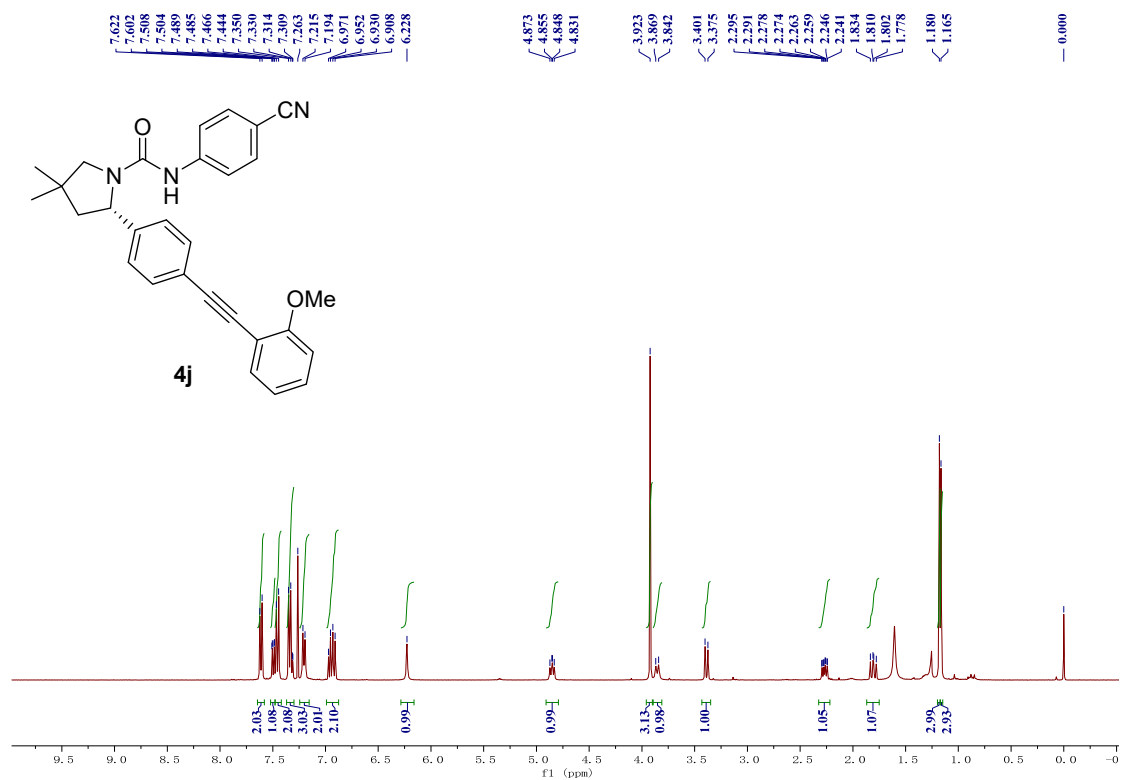
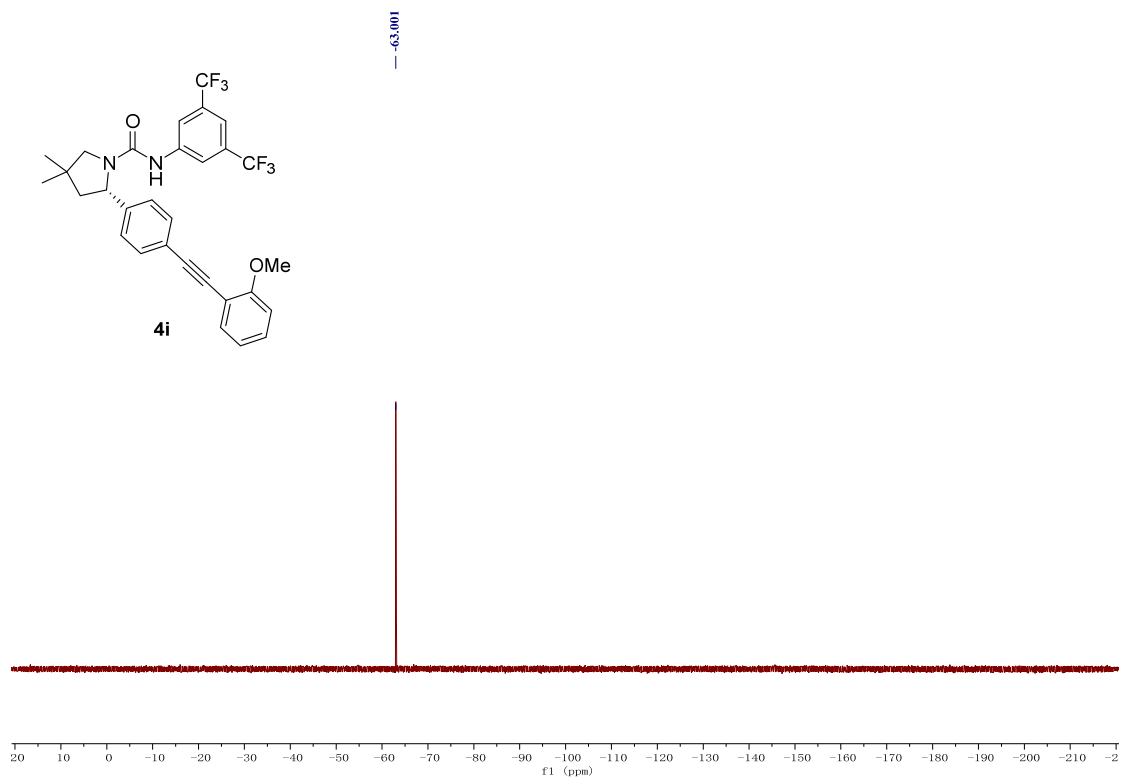


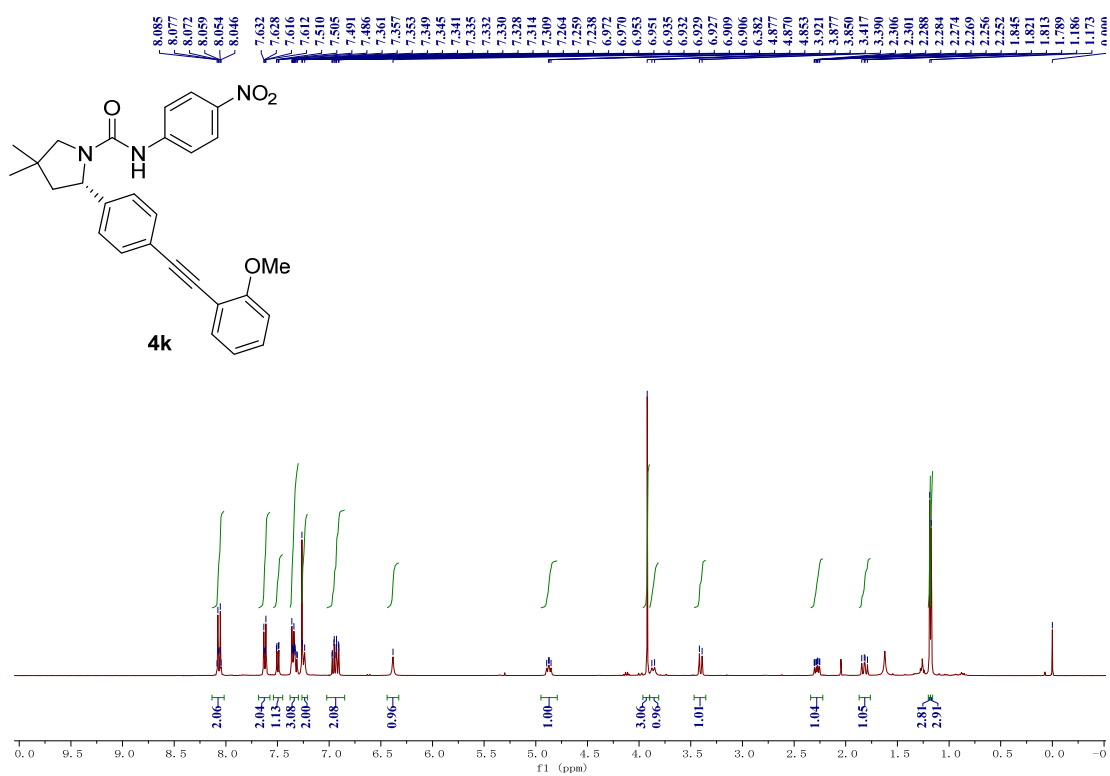
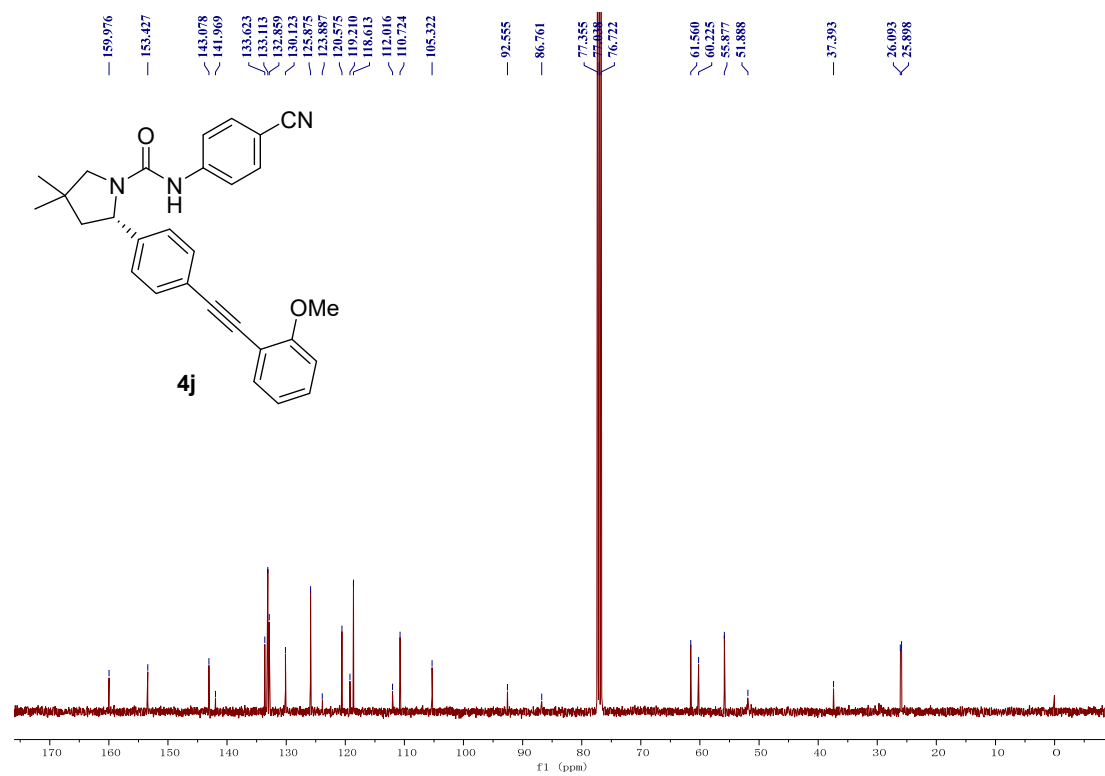
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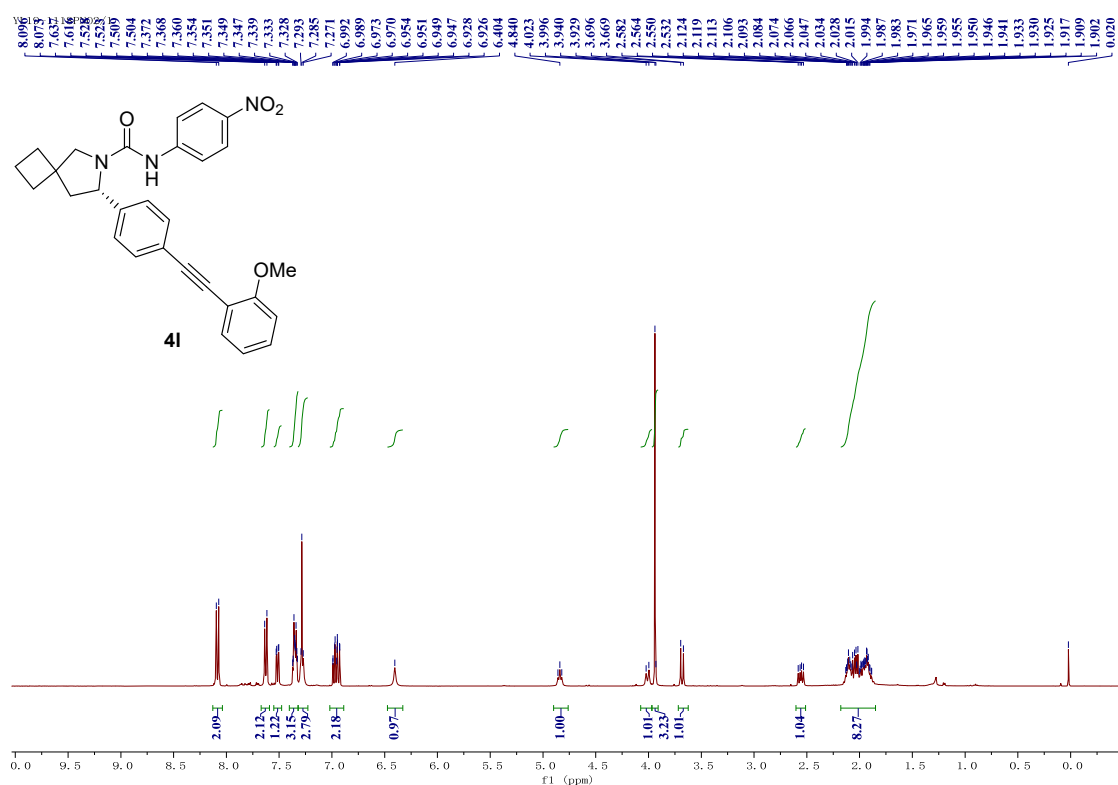
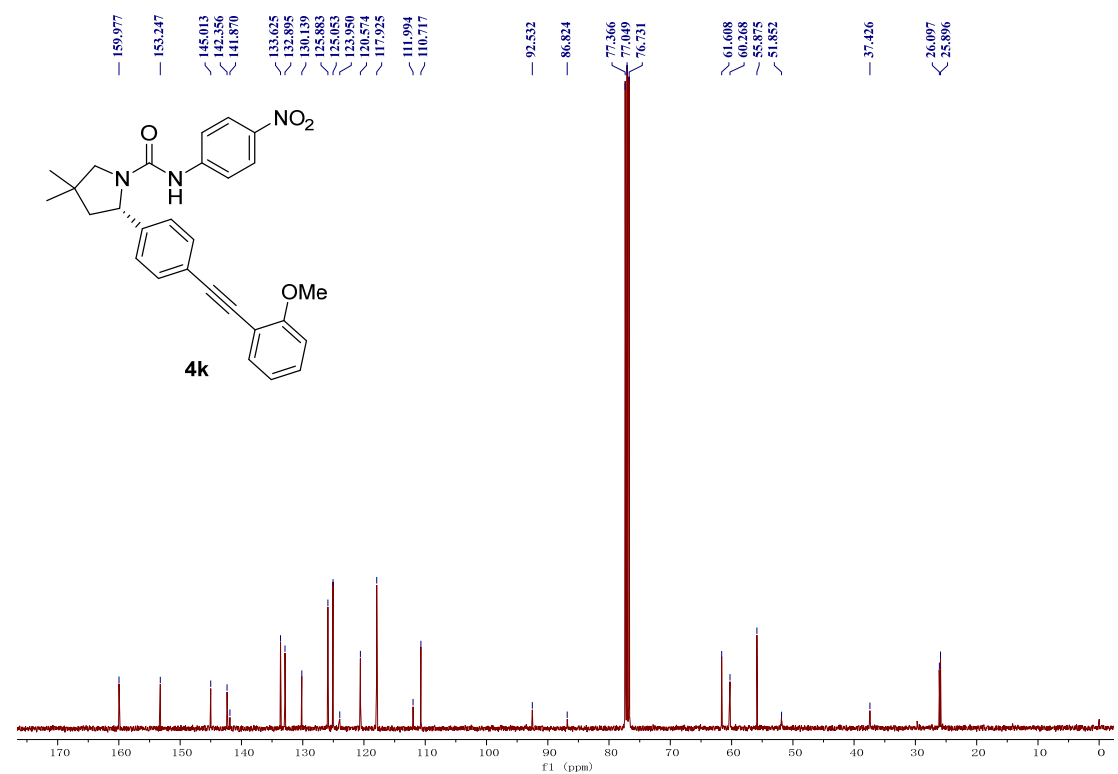


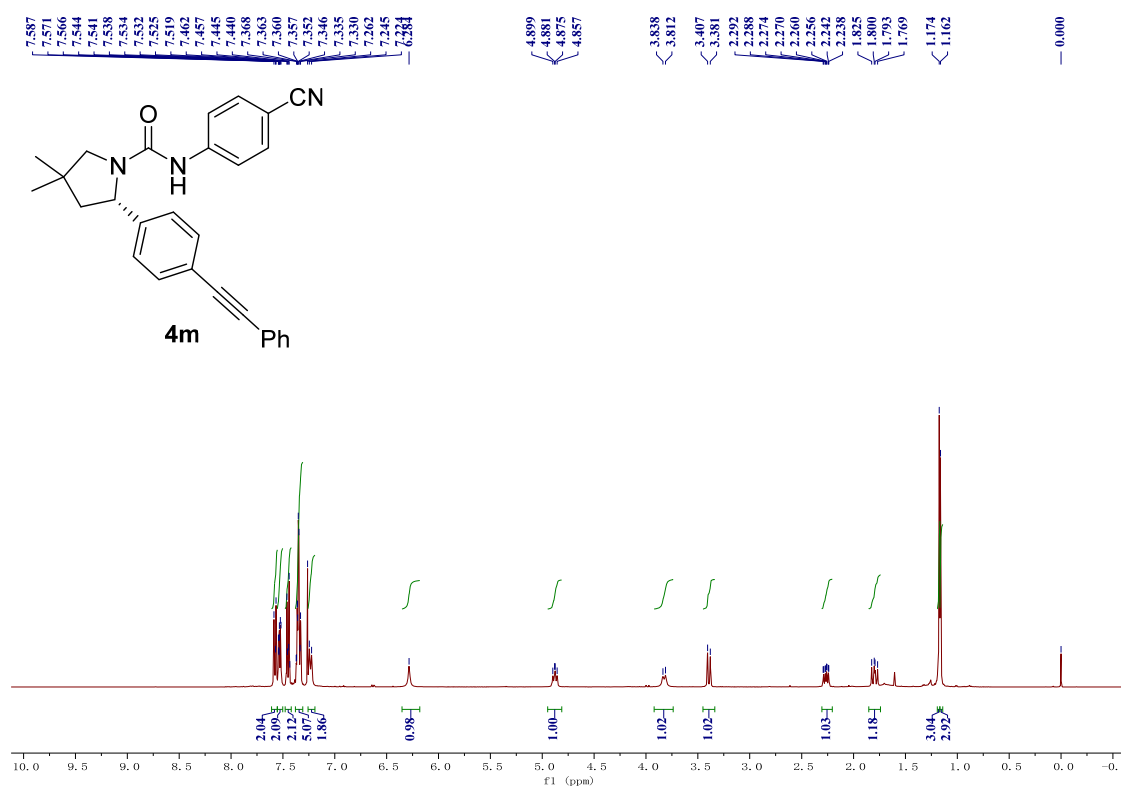
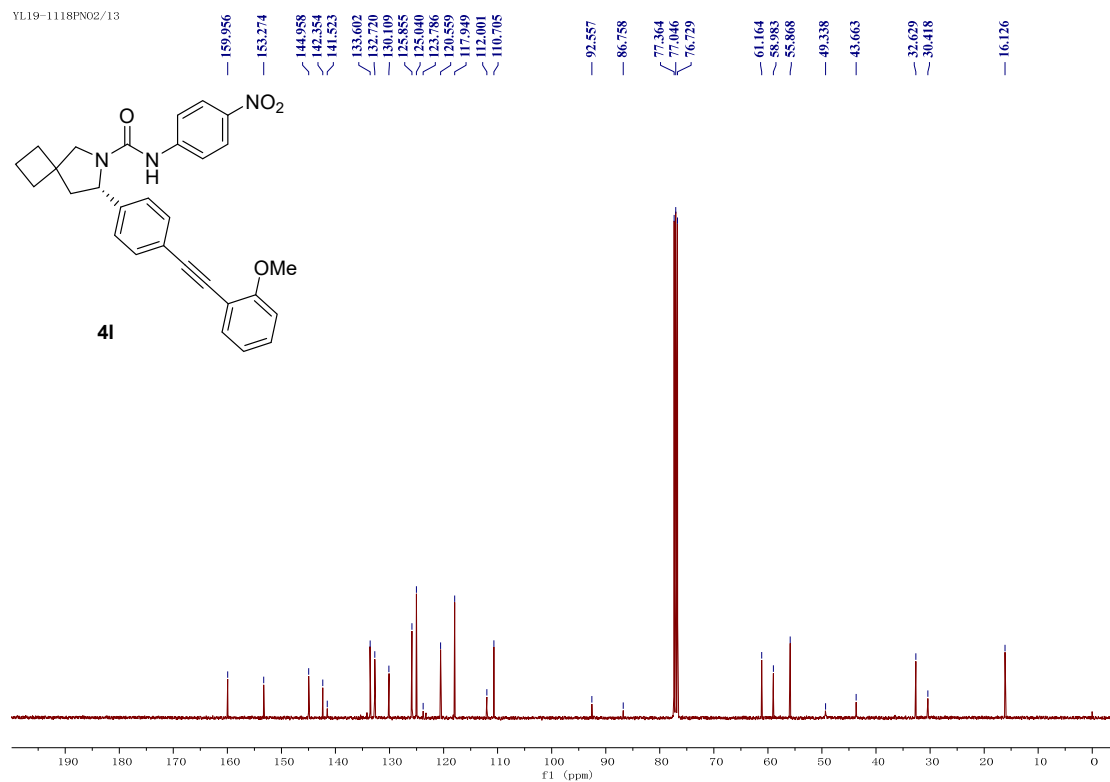


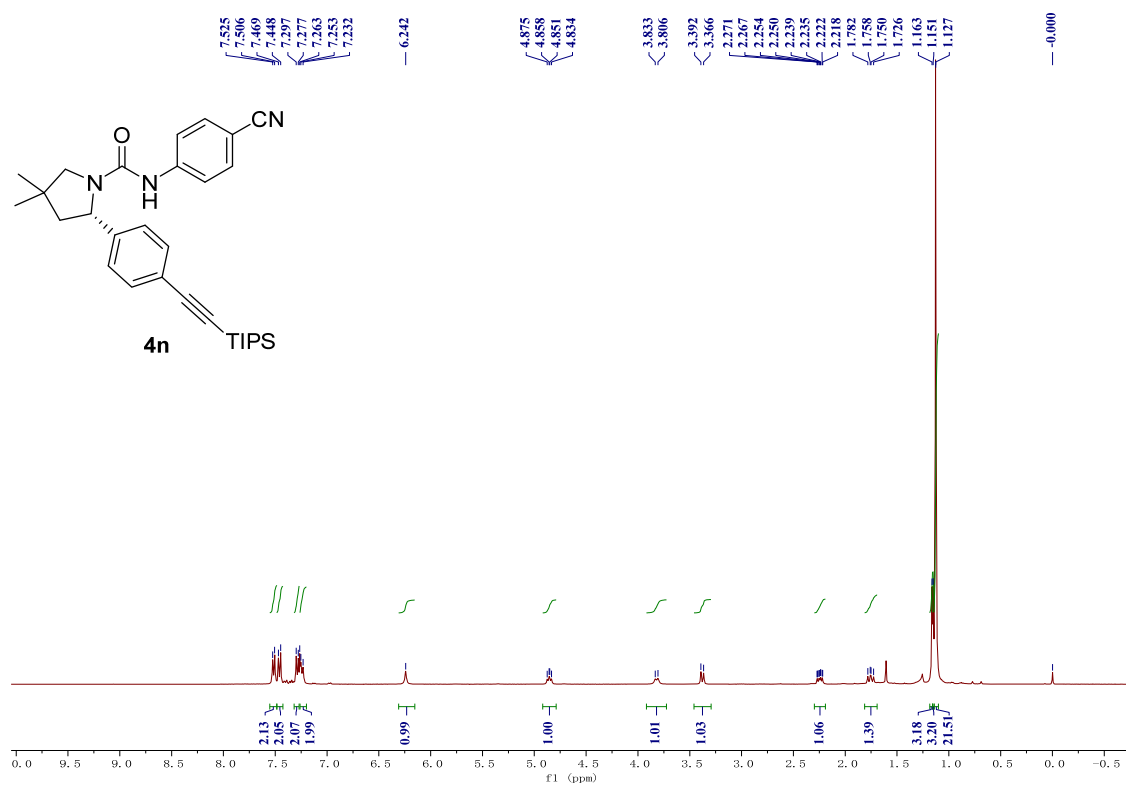
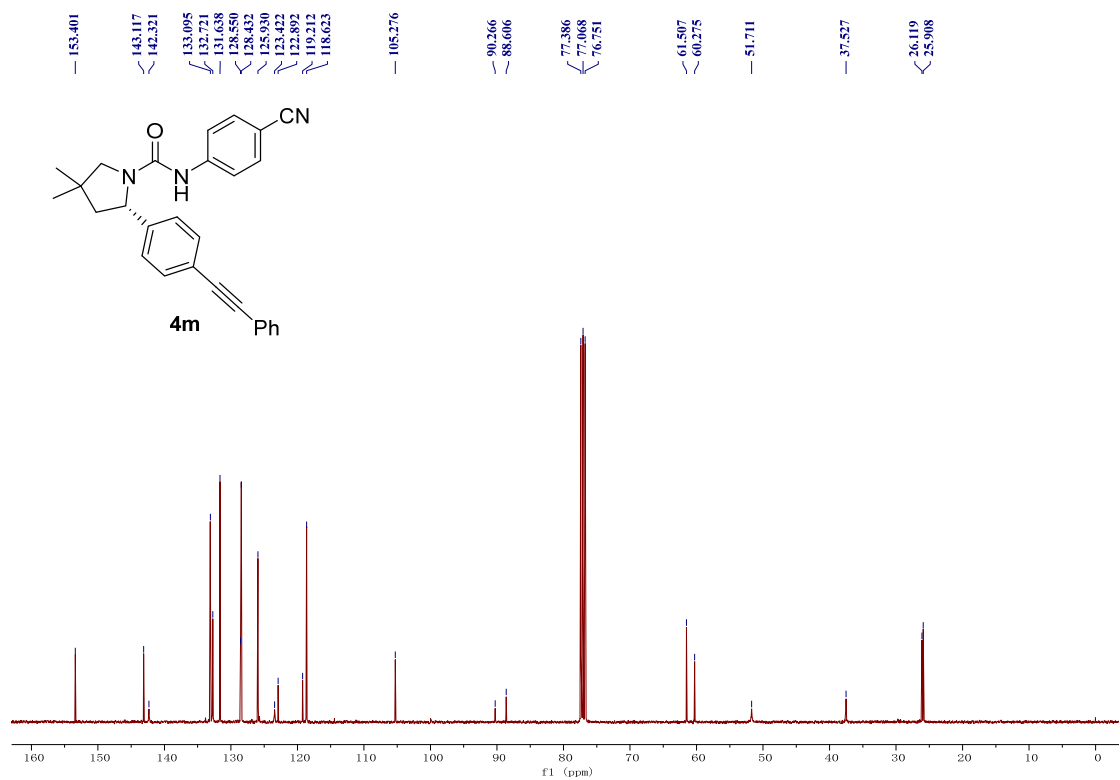


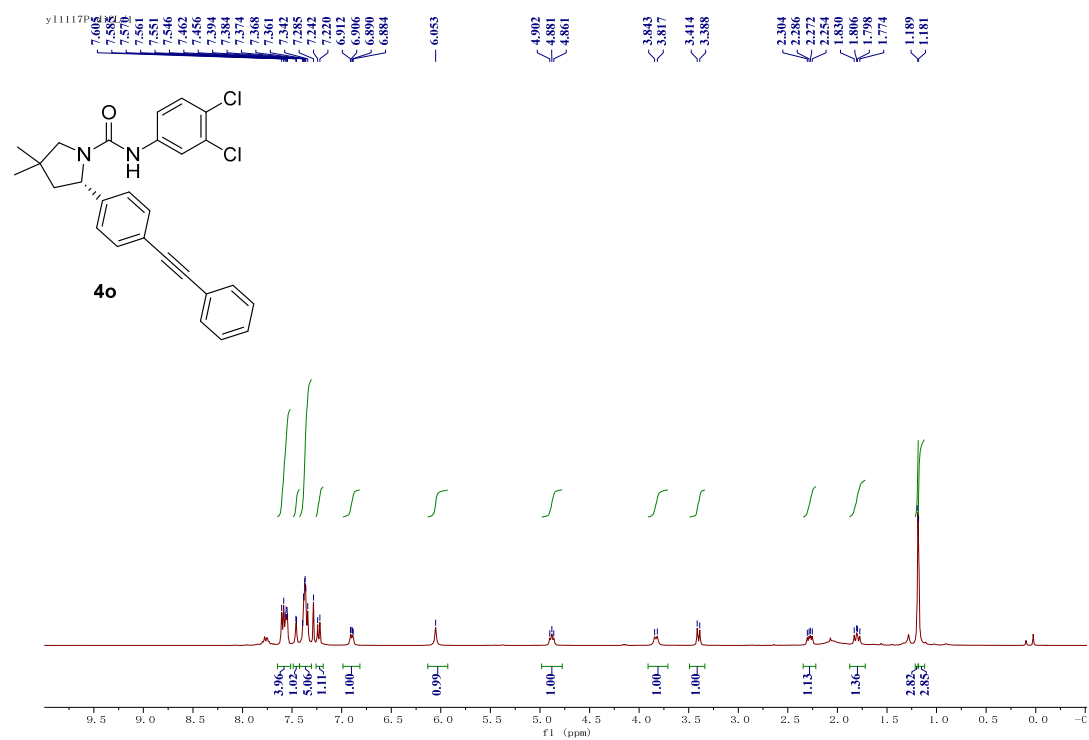
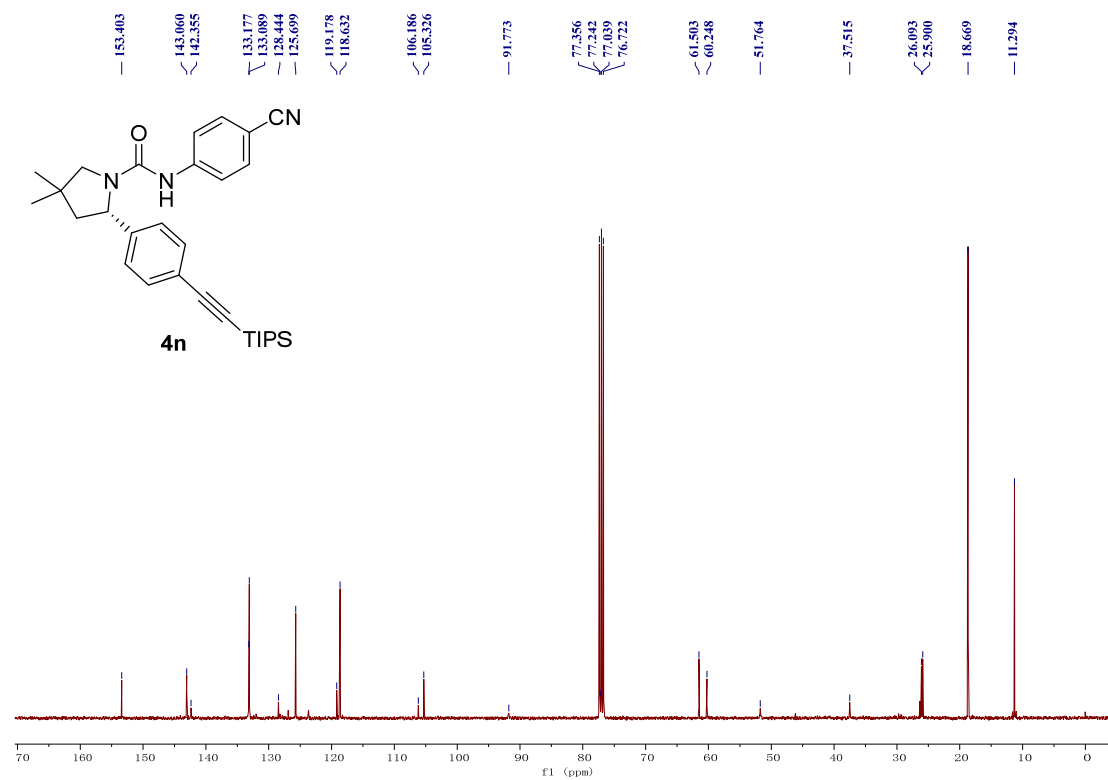


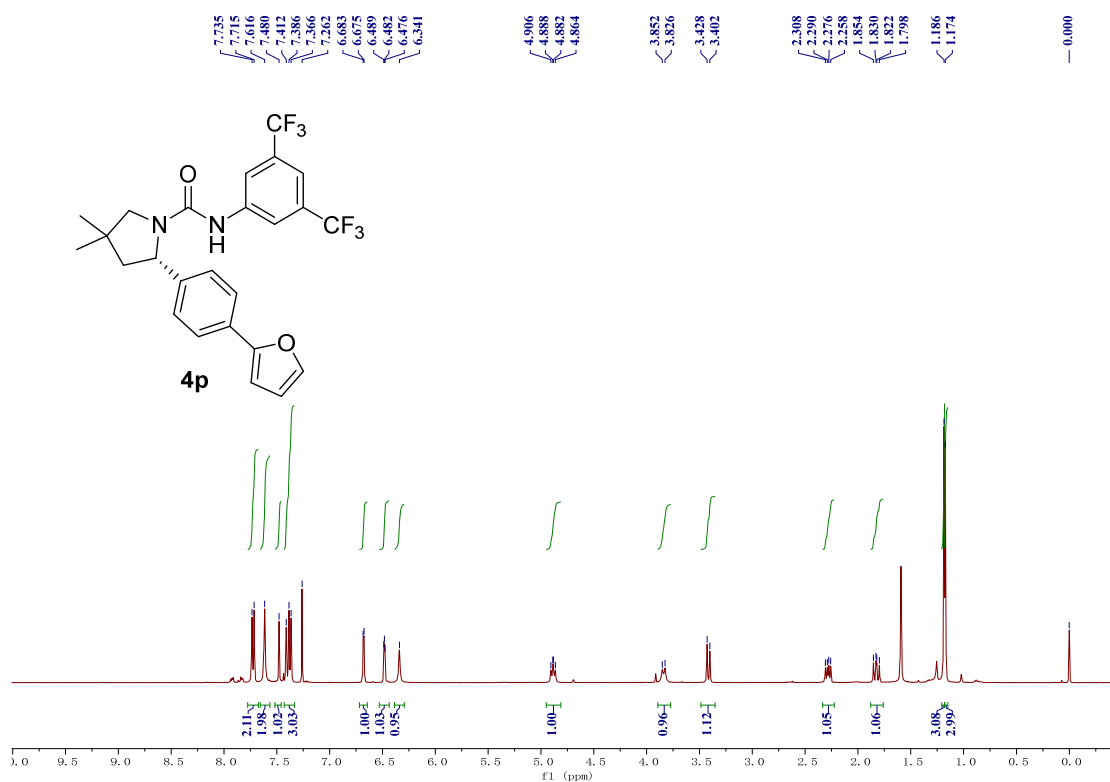
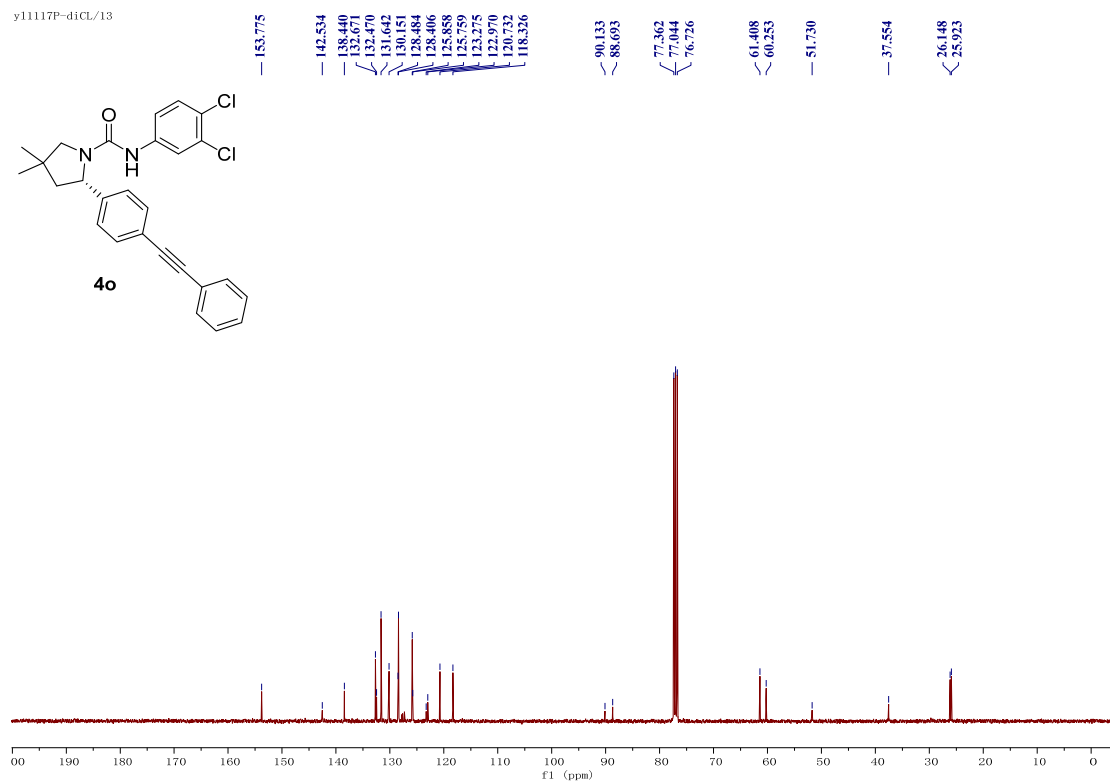


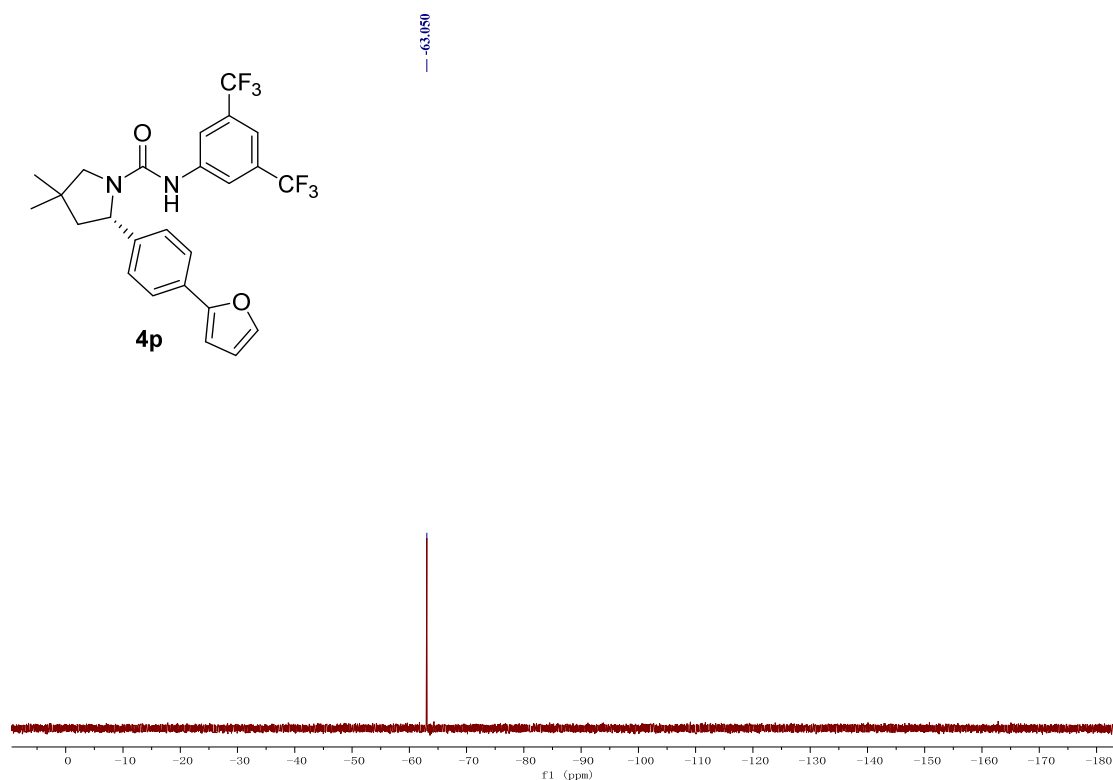
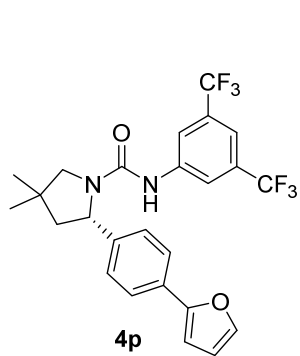
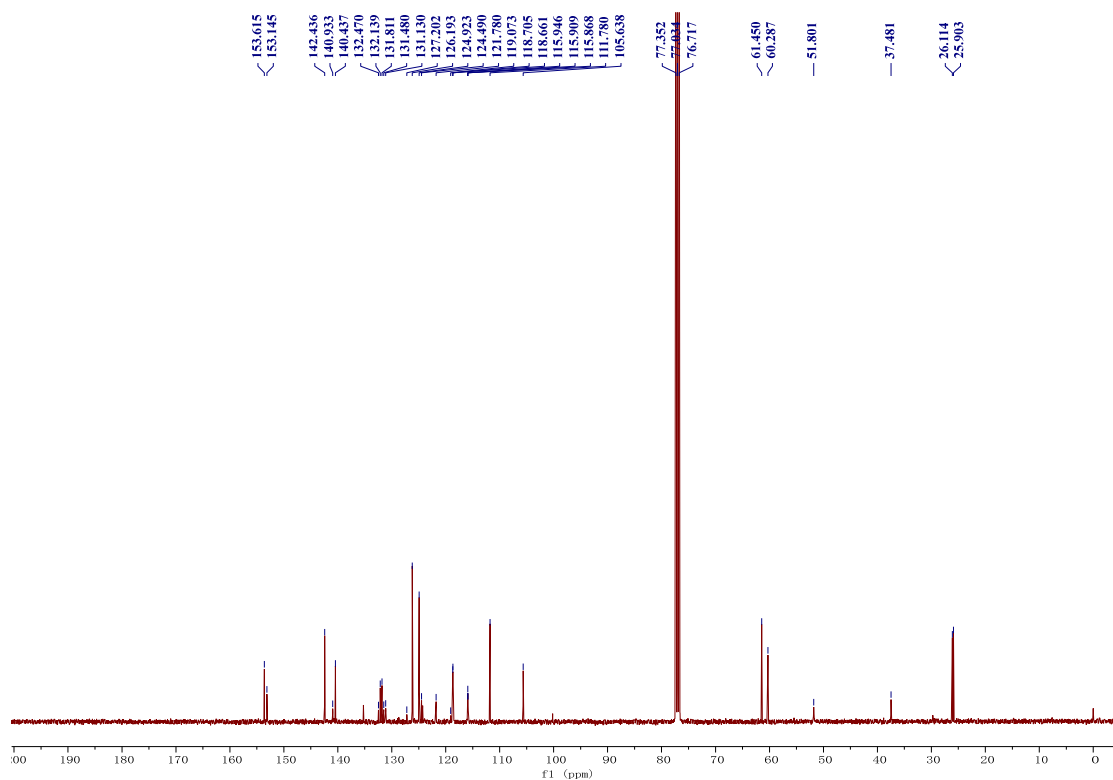


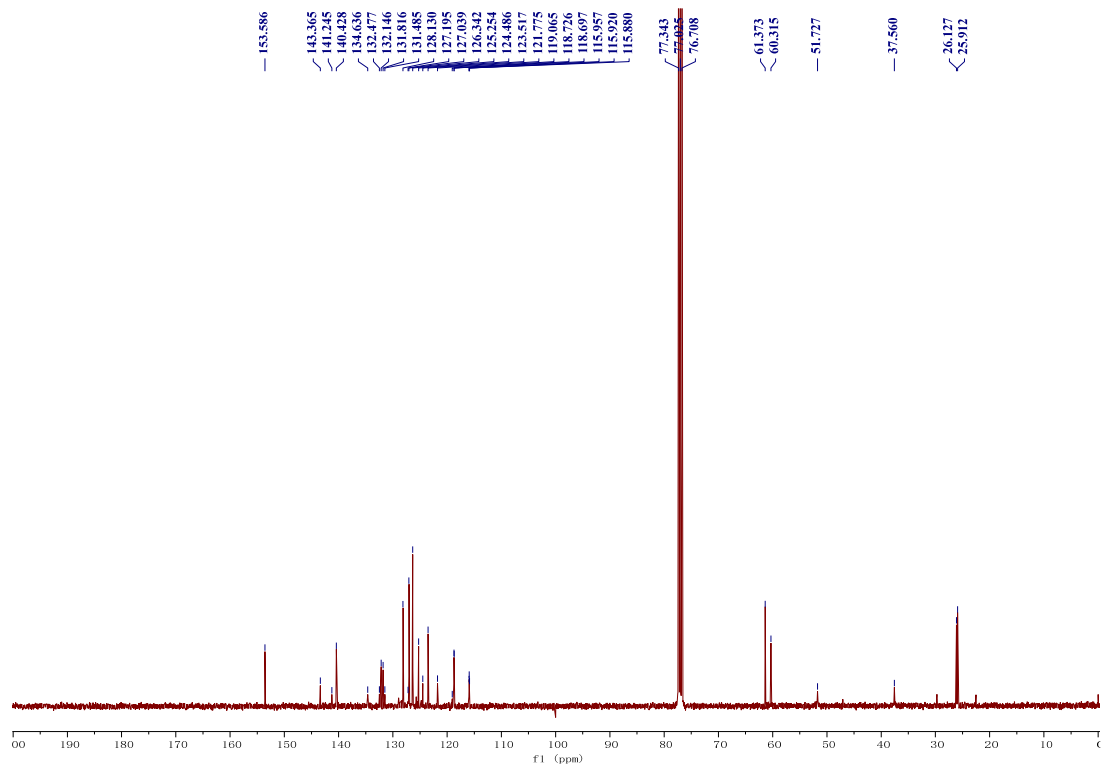
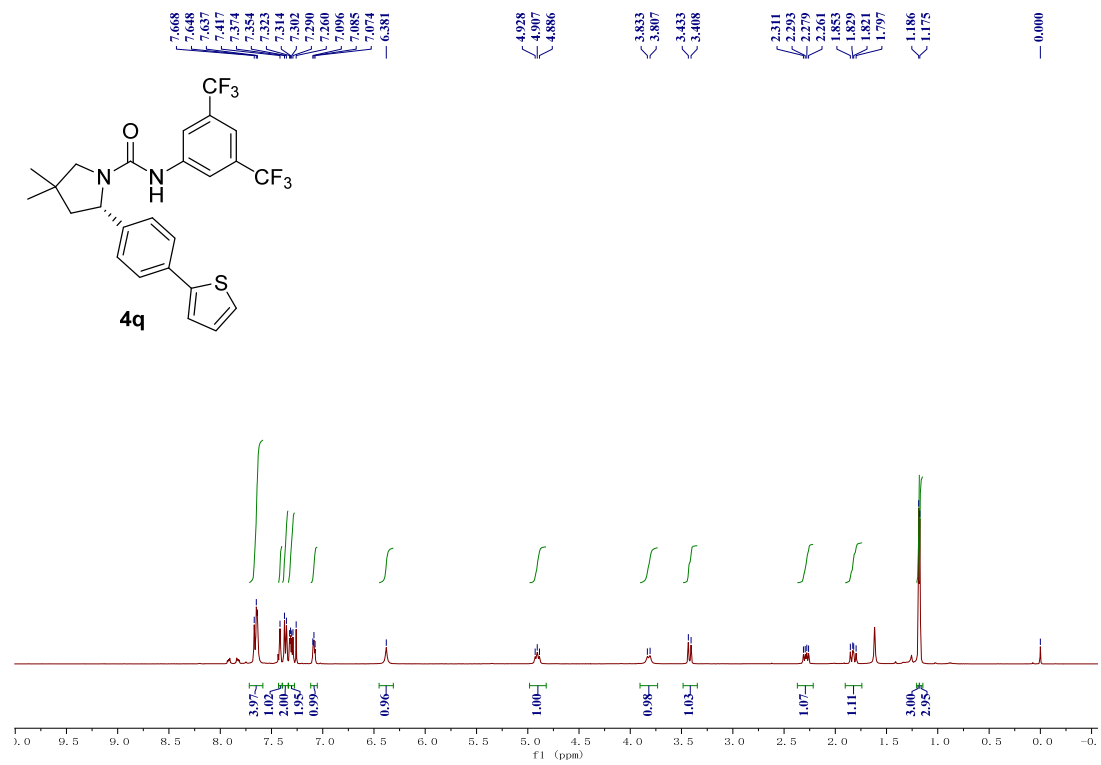


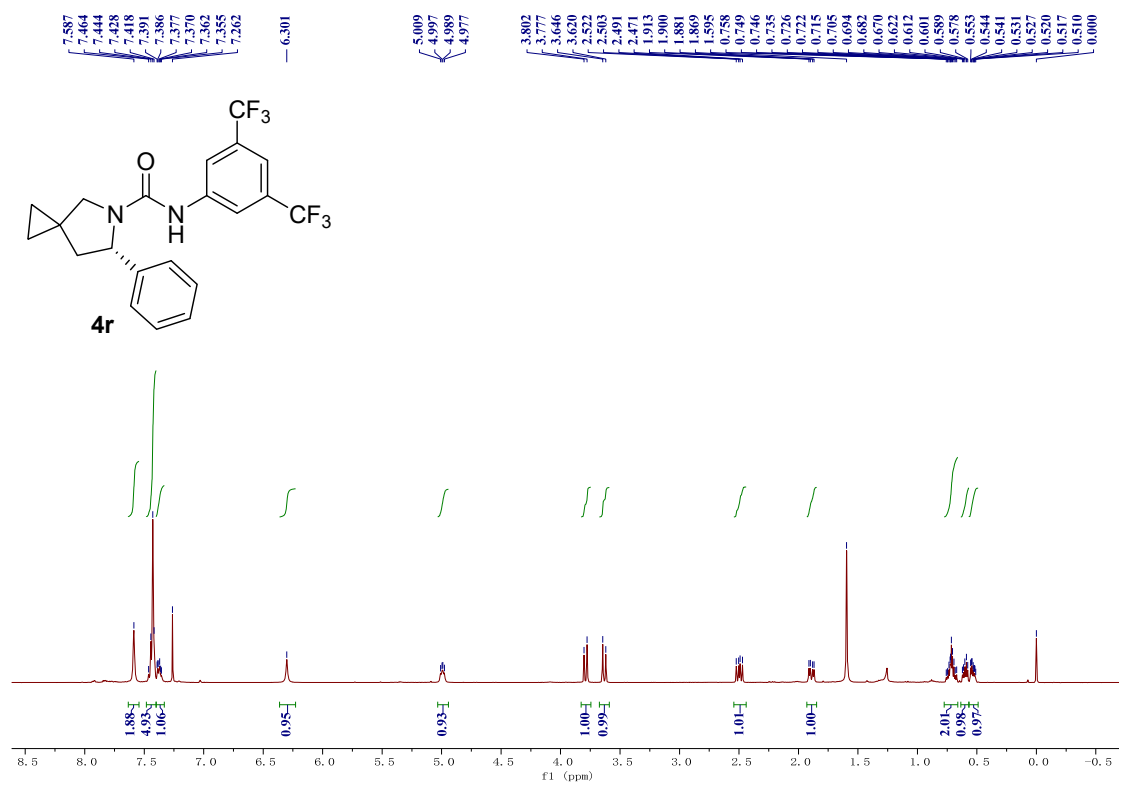
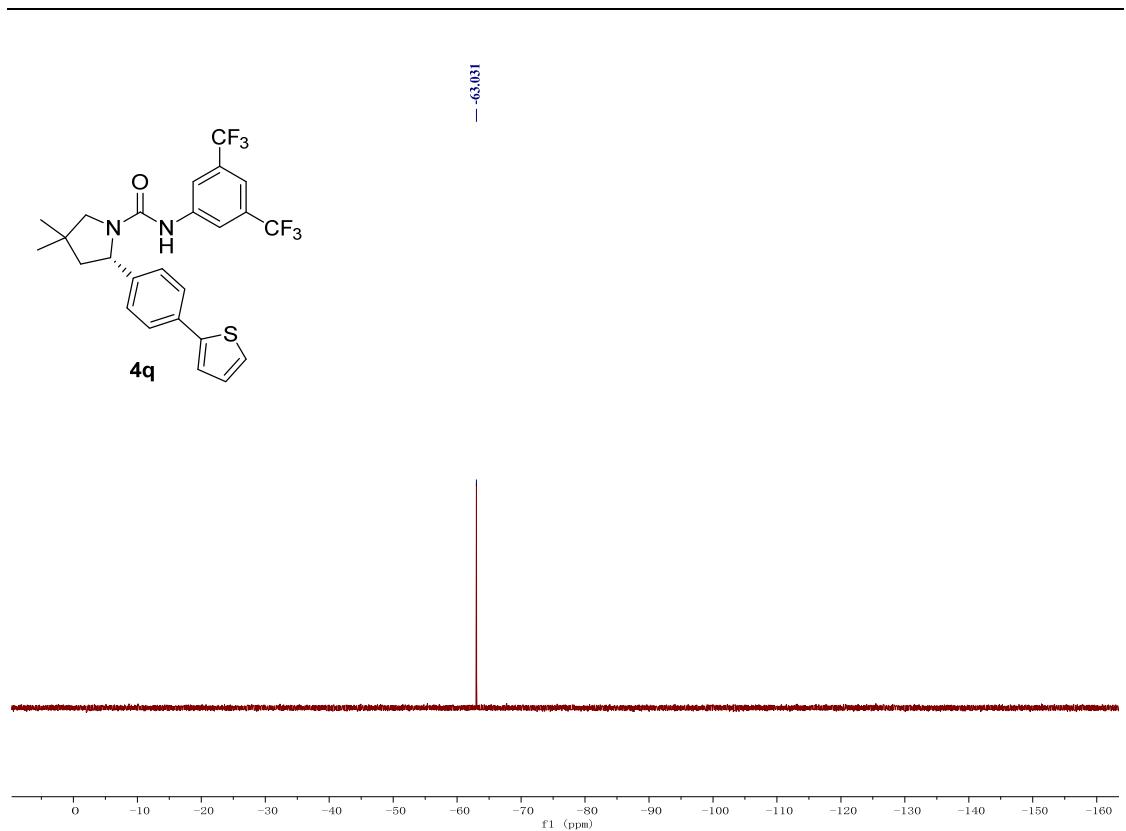


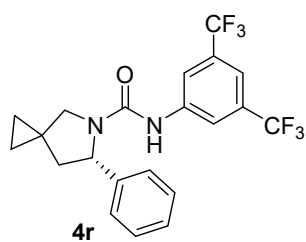
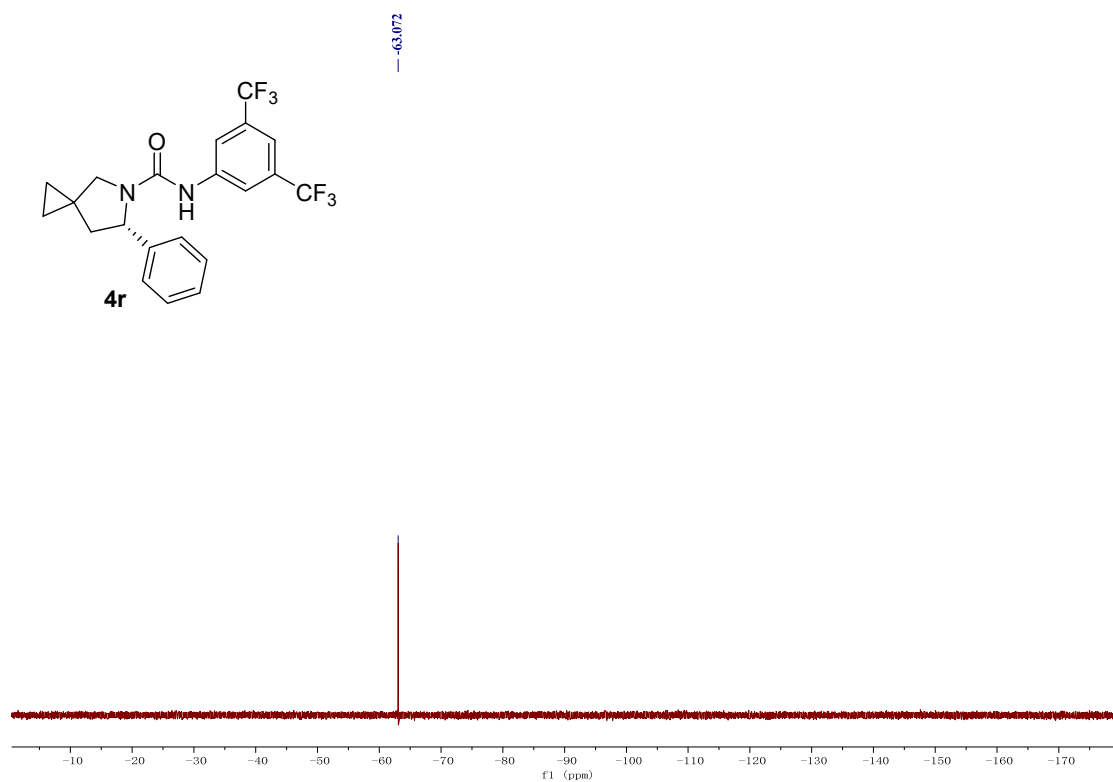
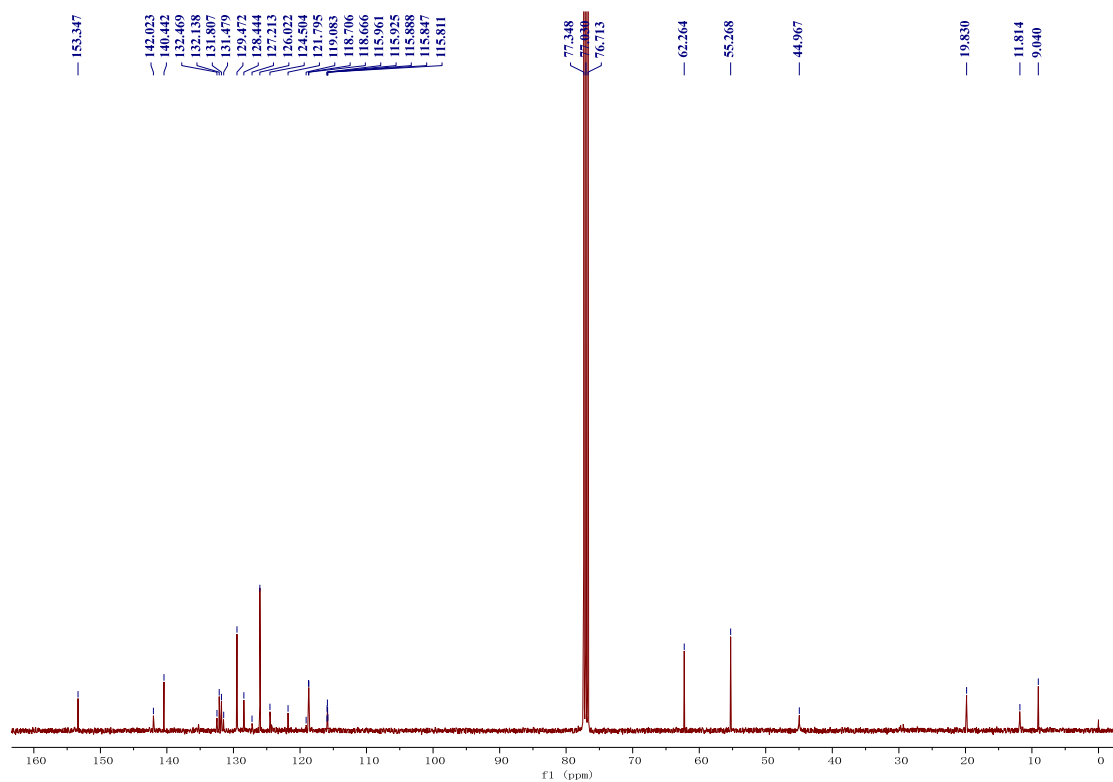


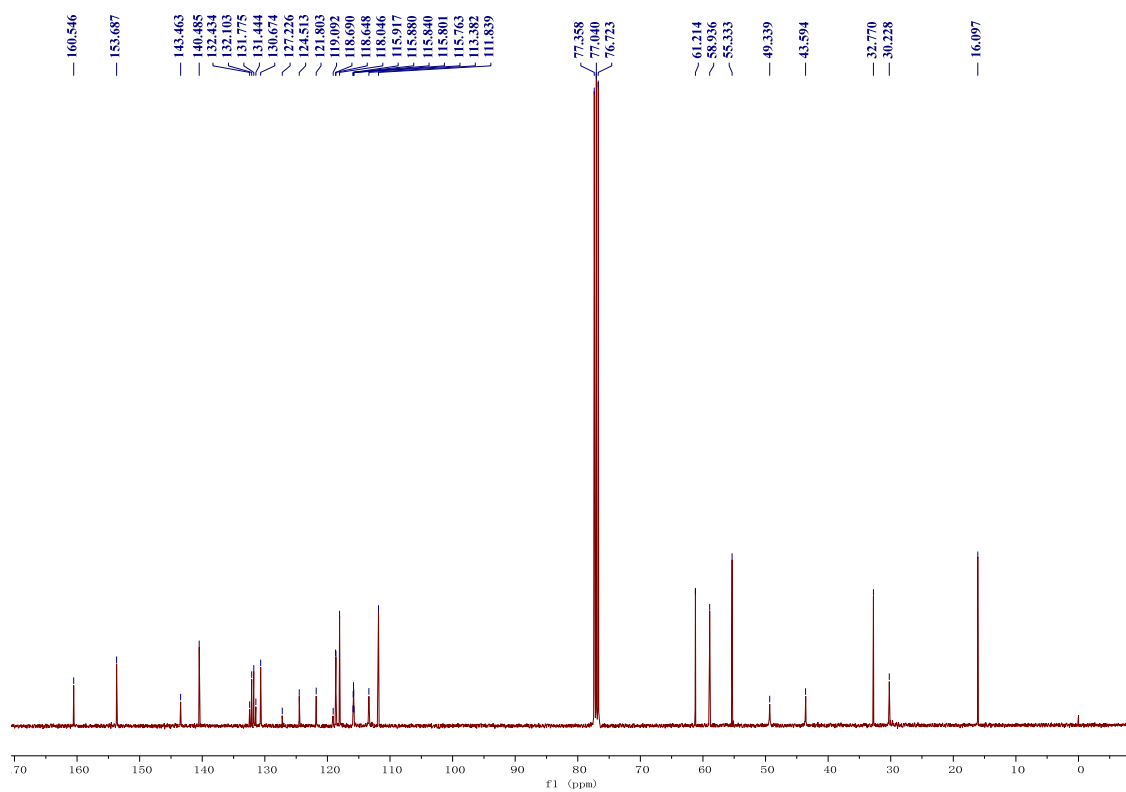
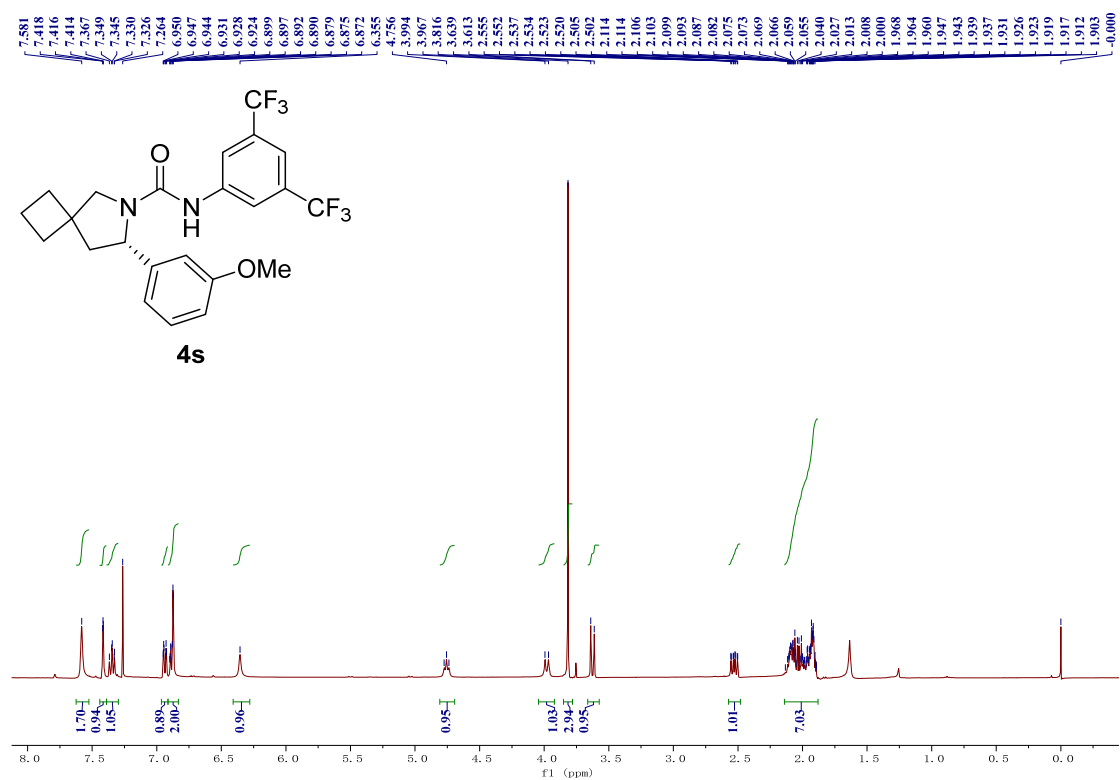


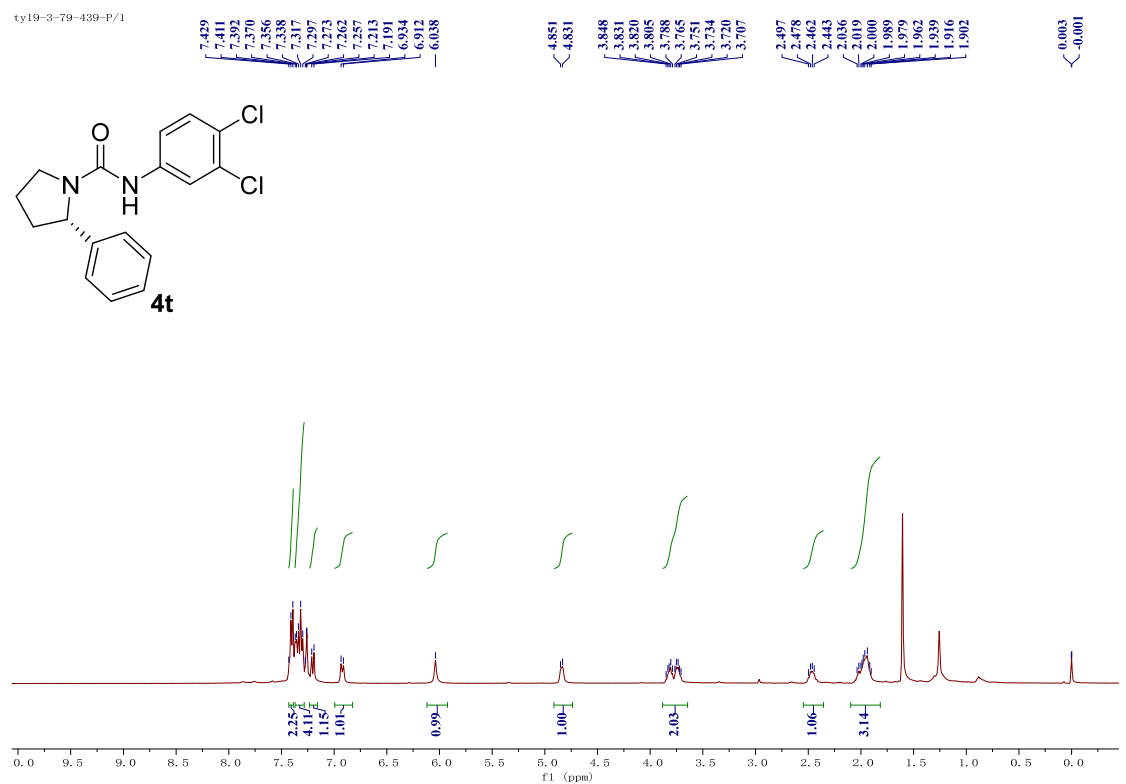
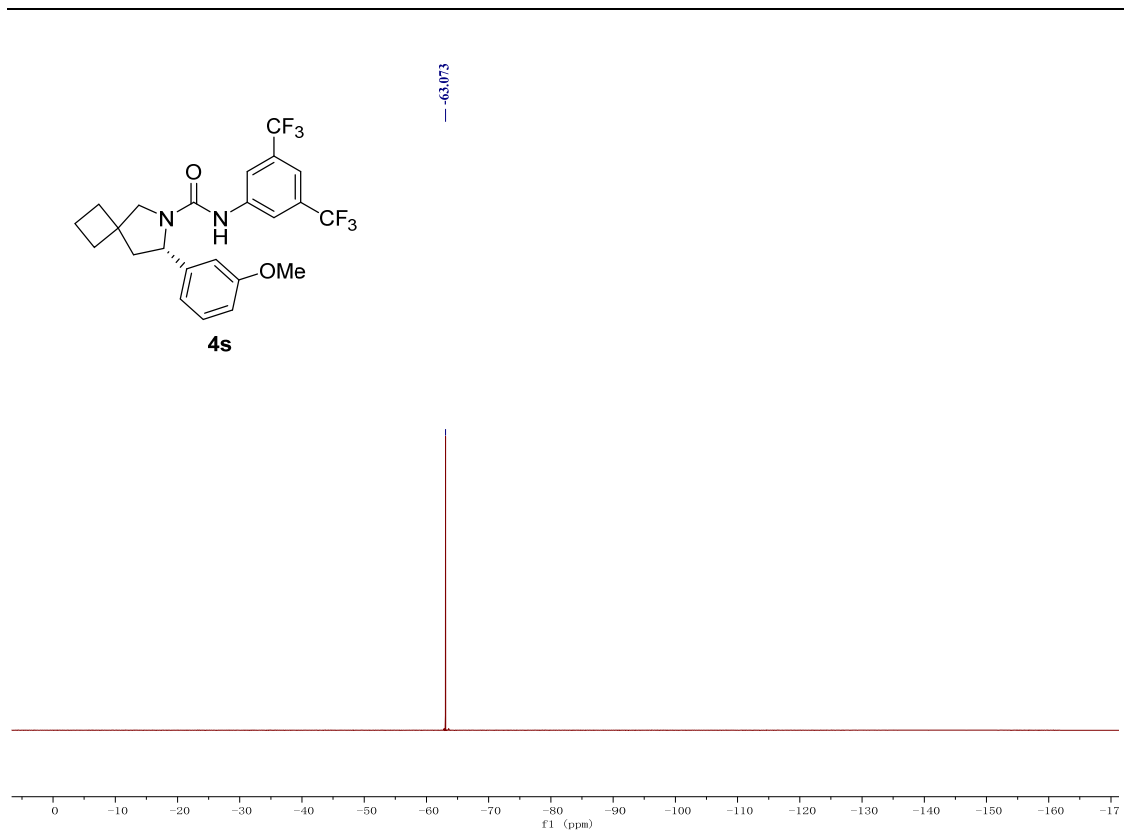




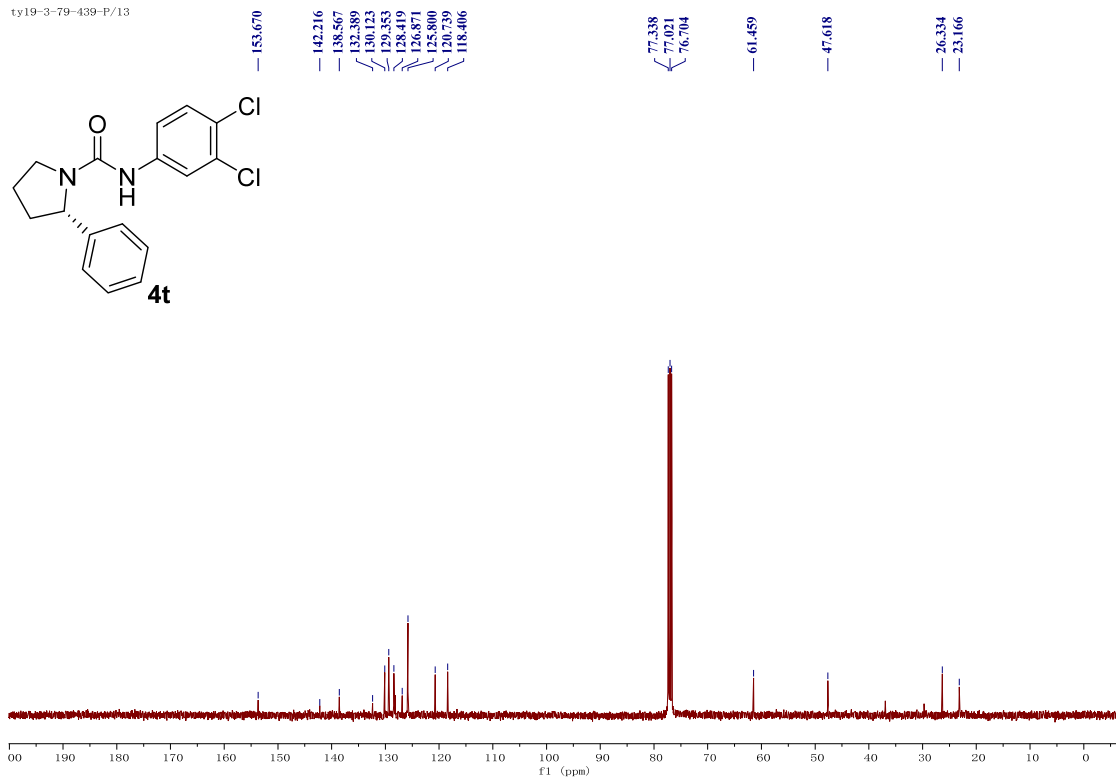




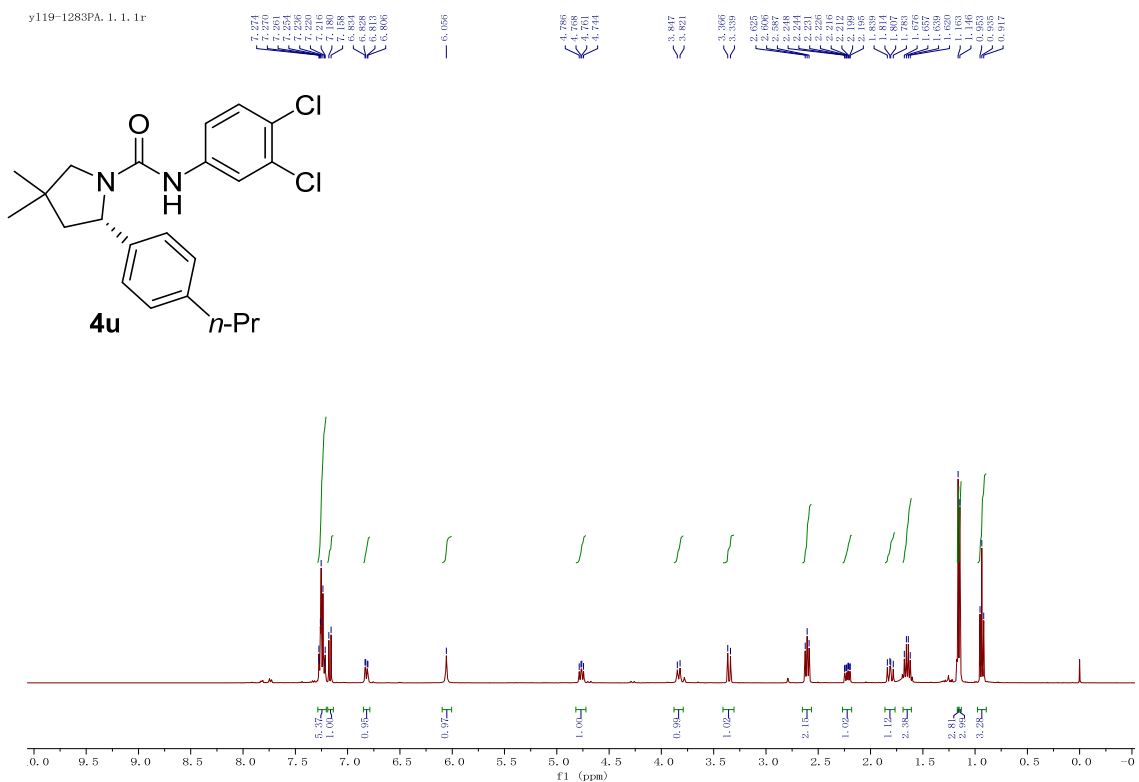




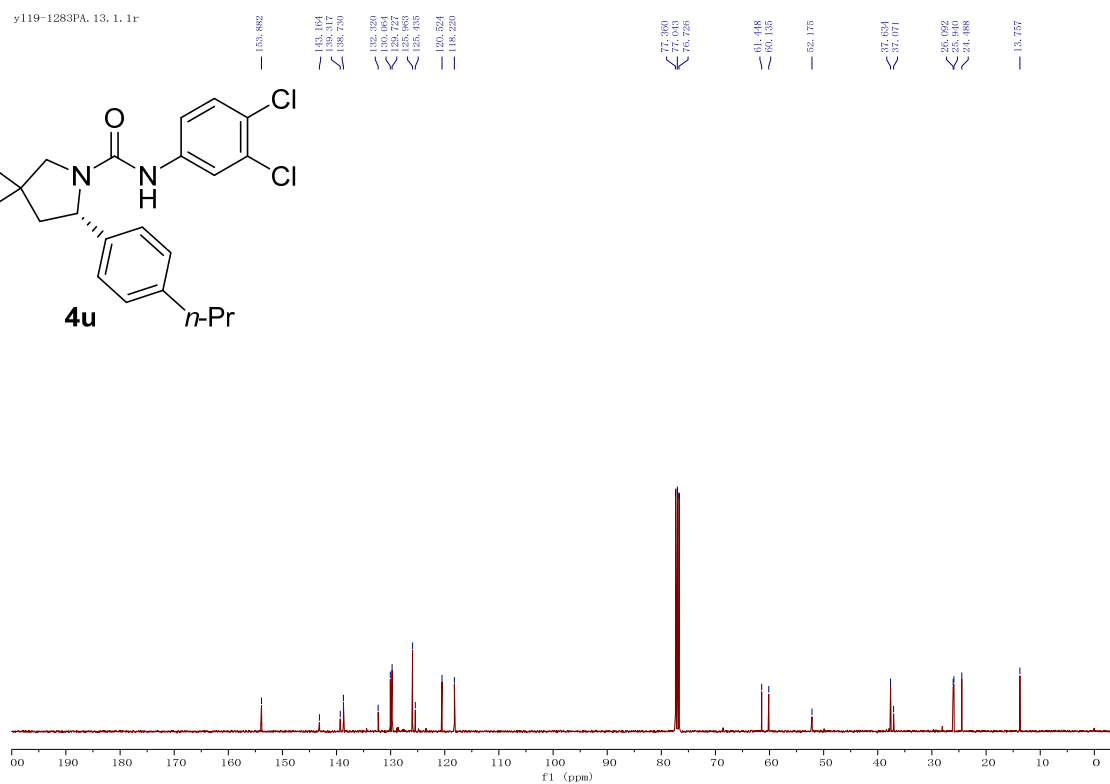
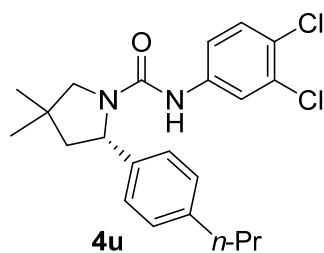
ty19-3-79-439-P/13



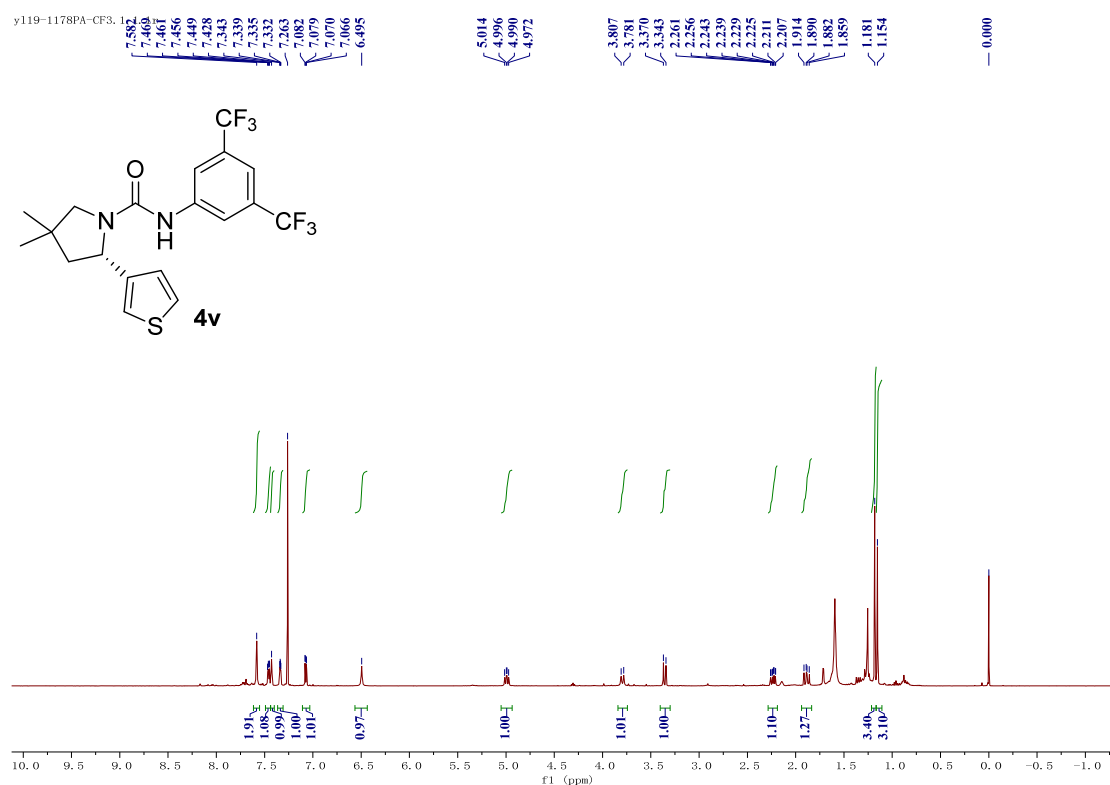
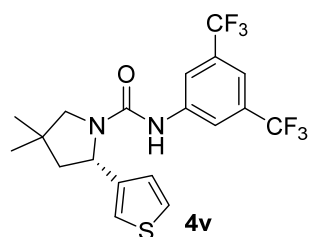
y119-1283PA. 1. 1. 1r



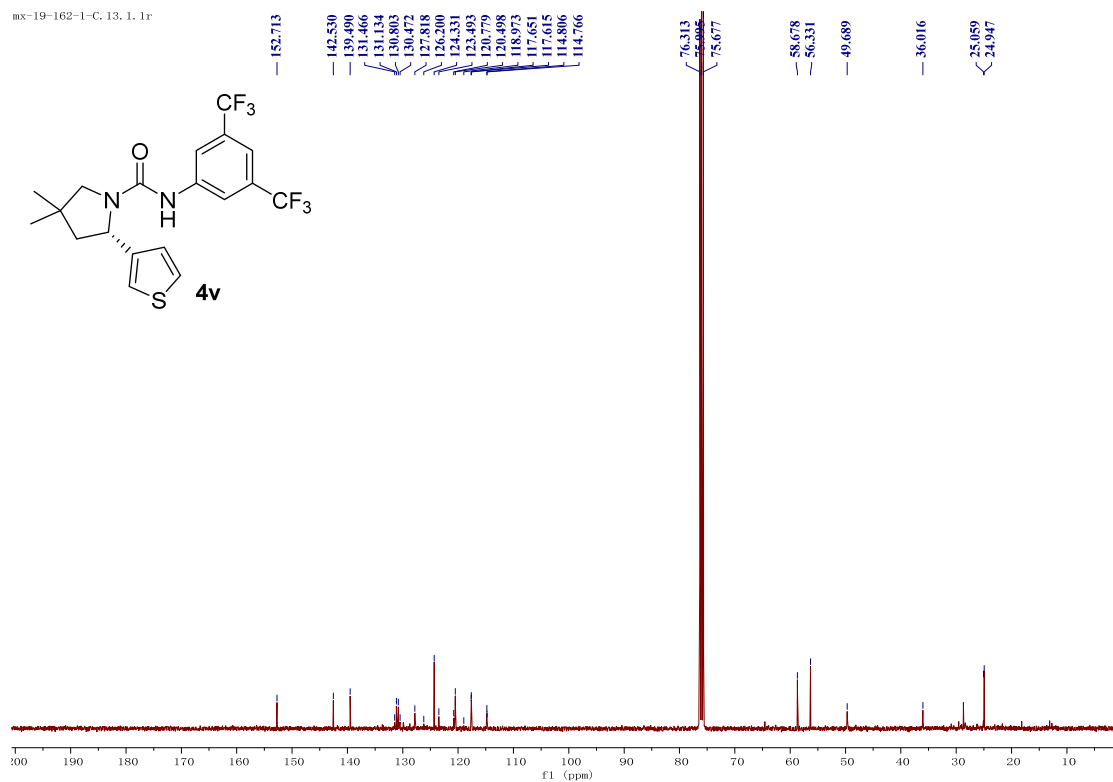
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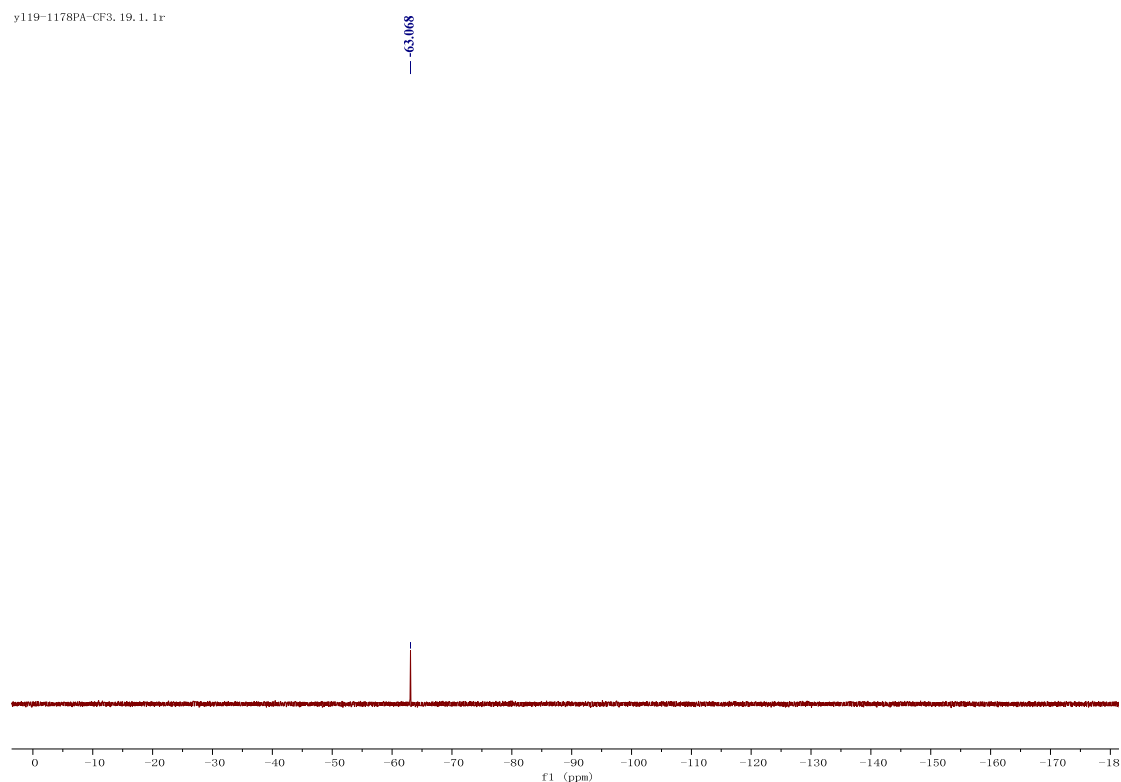
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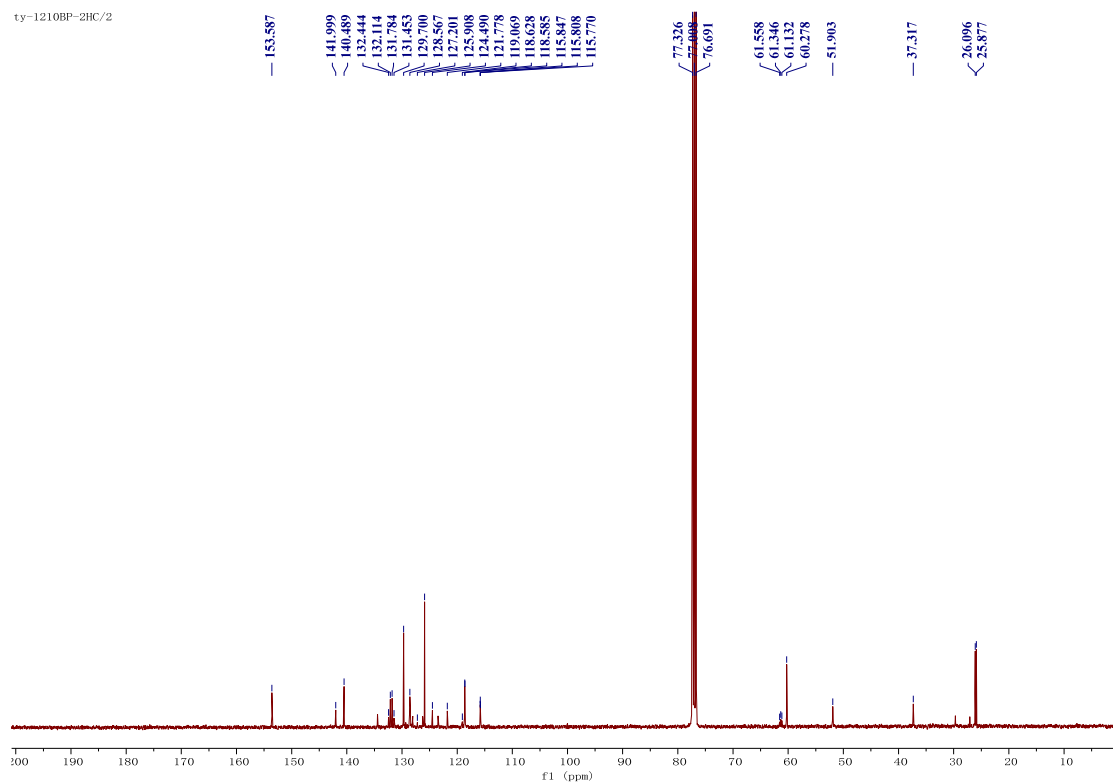
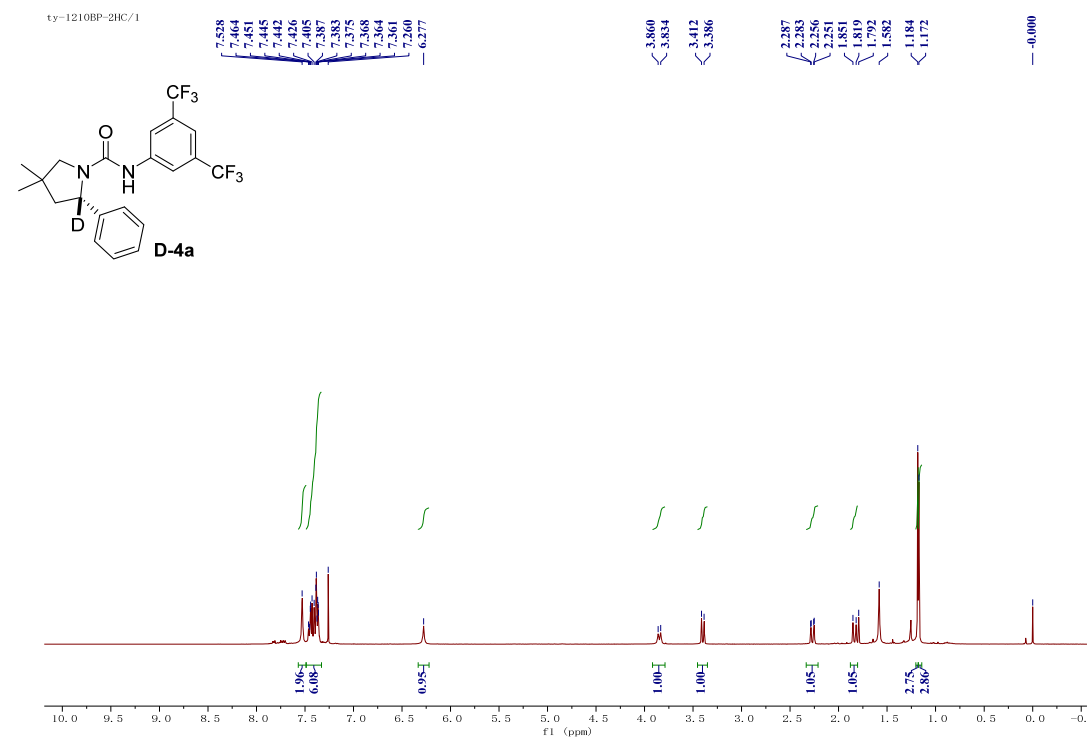


mx-19-162-1-C, 13, 1, 1r

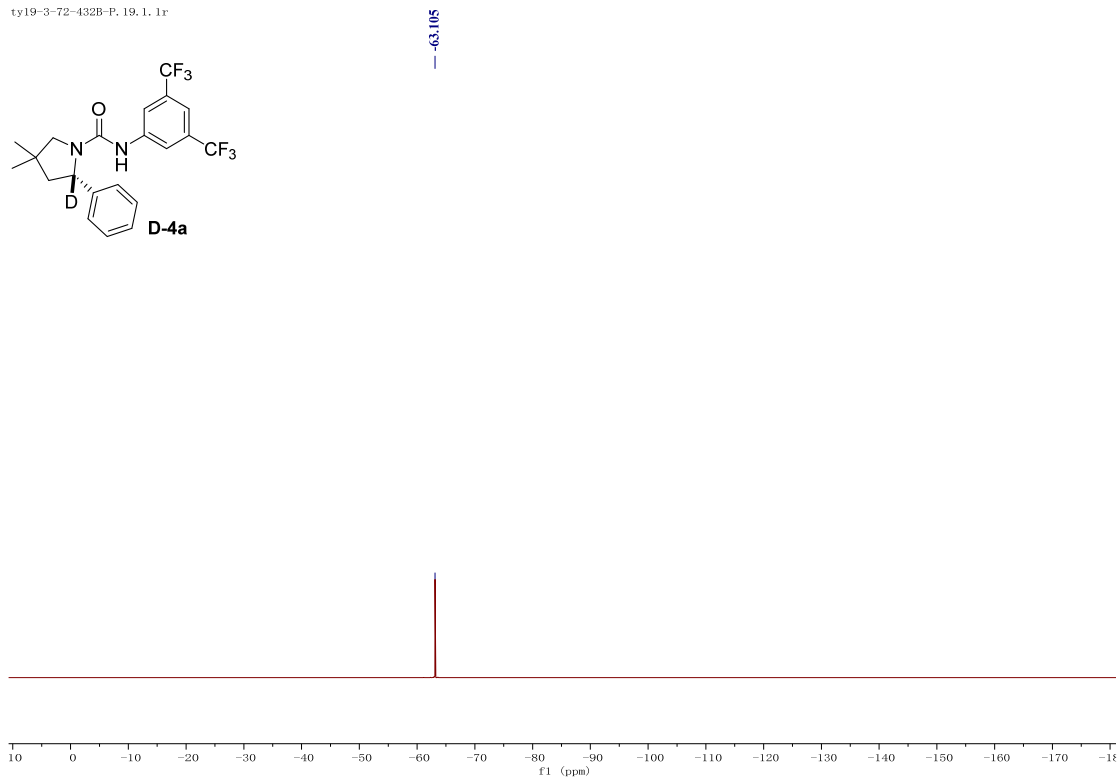
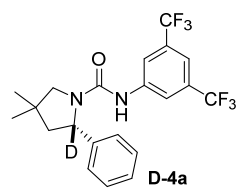


y119-1178PA-CF3, 19.1, 1r

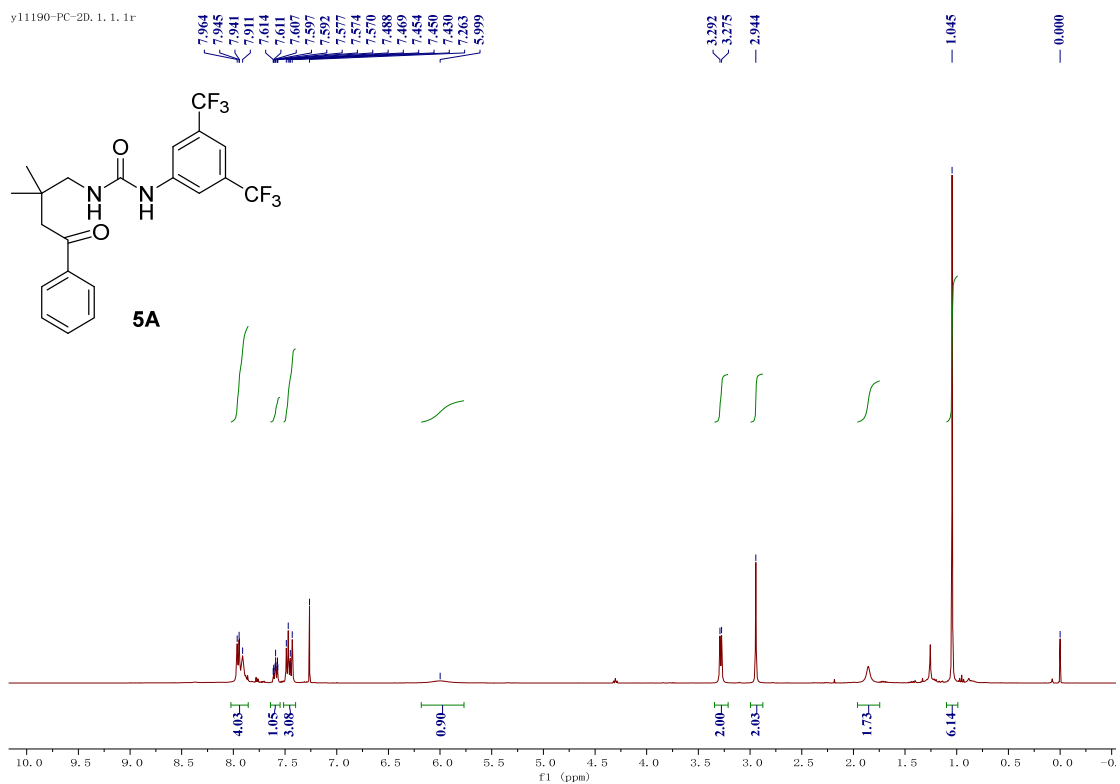
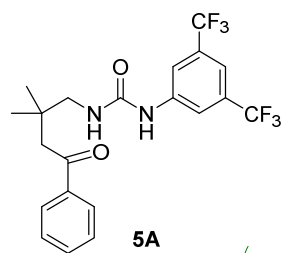


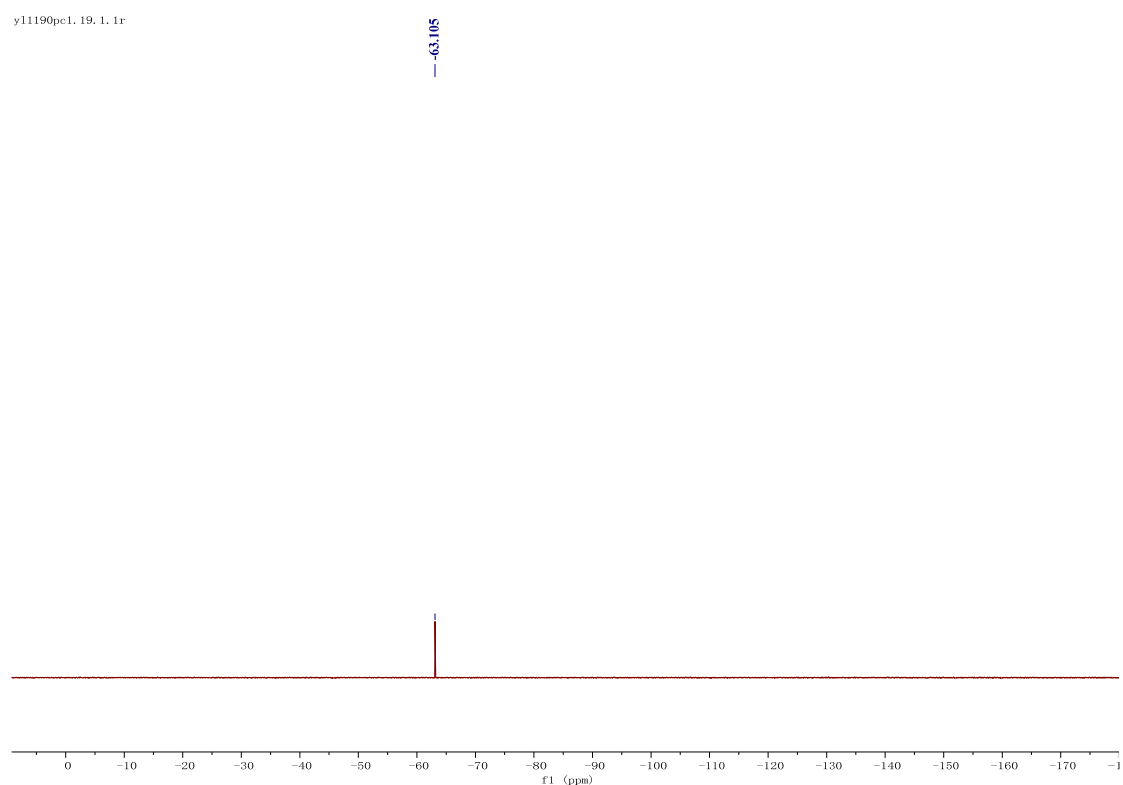
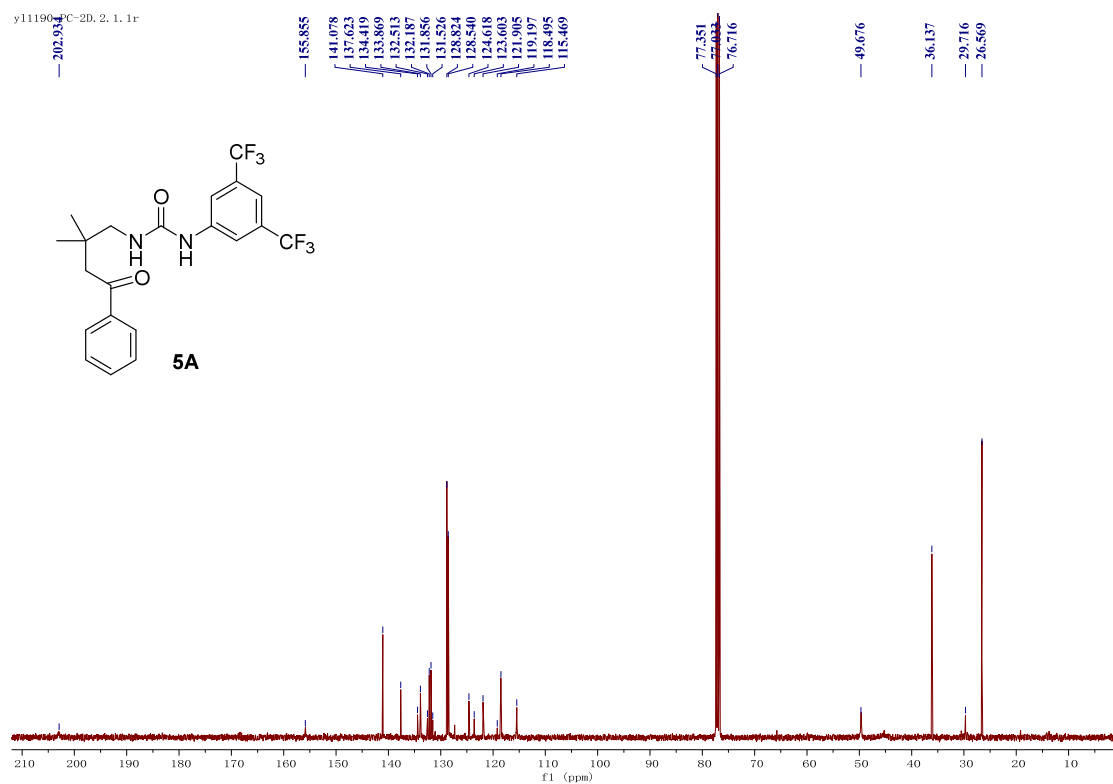


ty19-3-72-432B-P, 19, 1, 1r

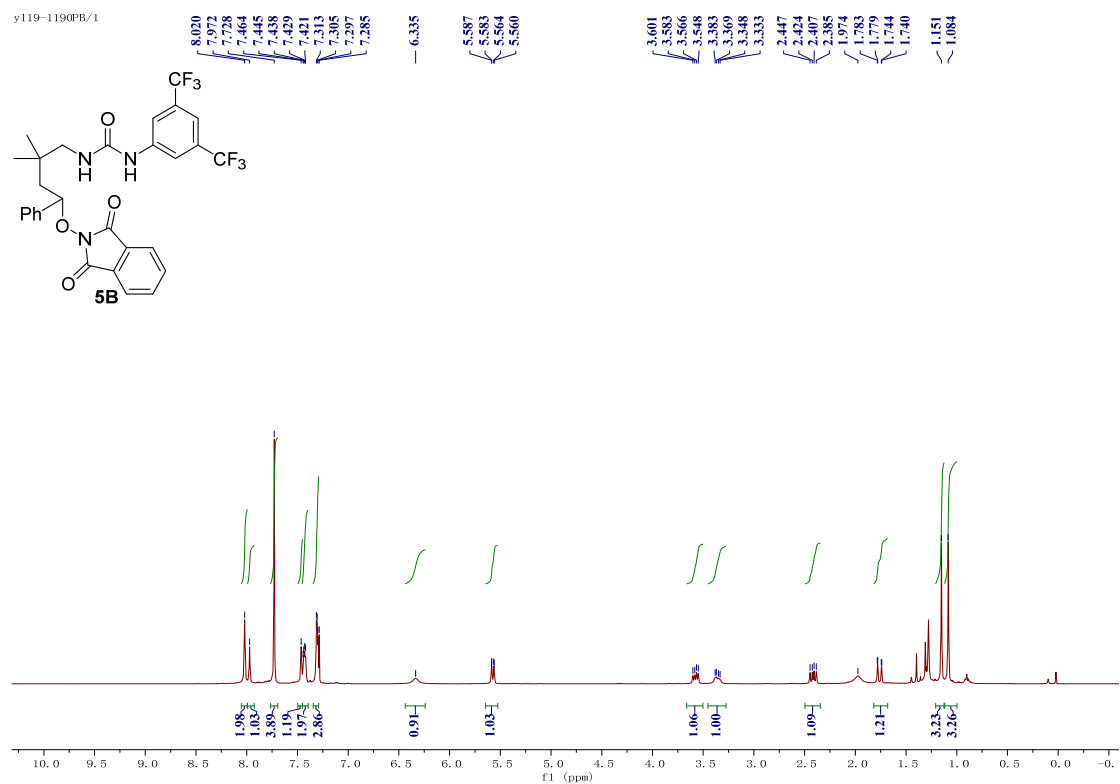


y11190-PC-2D, 1, 1, 1r

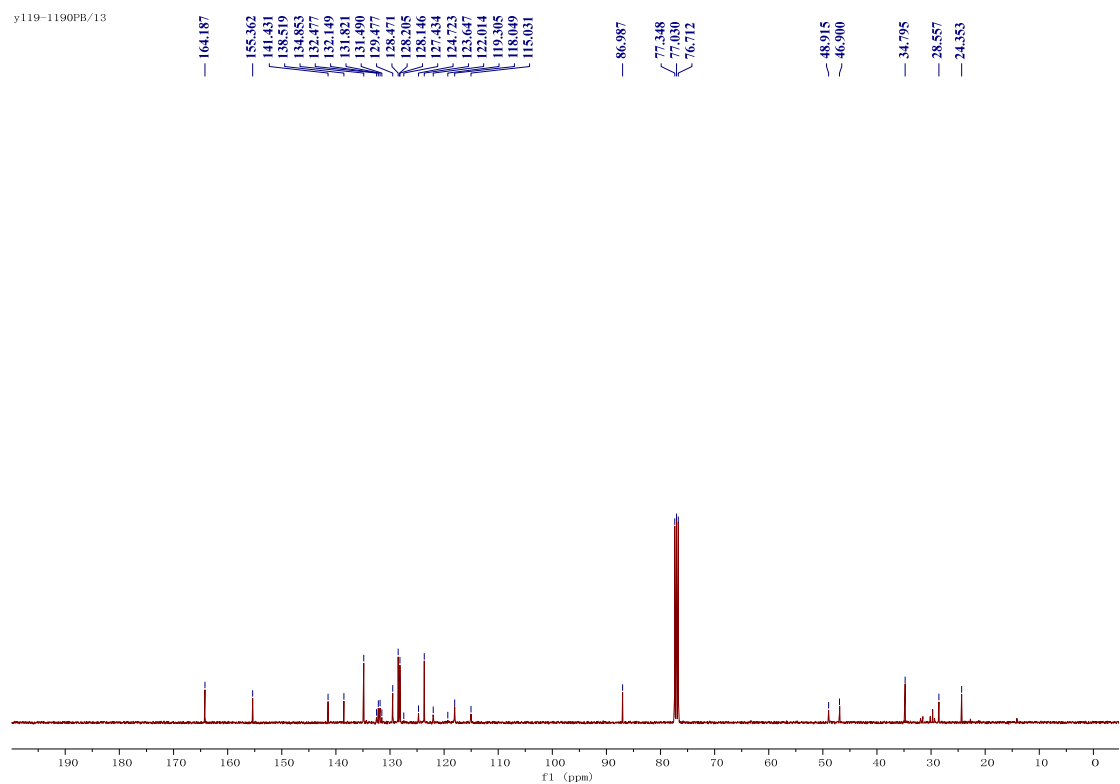




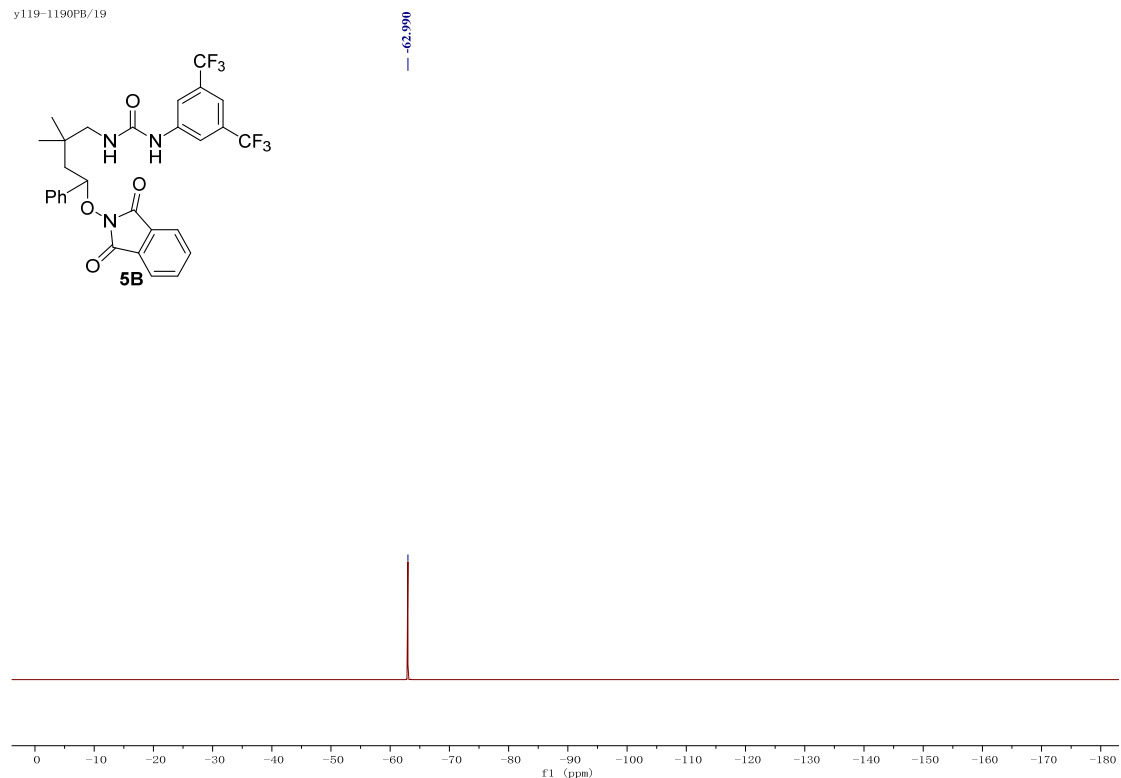
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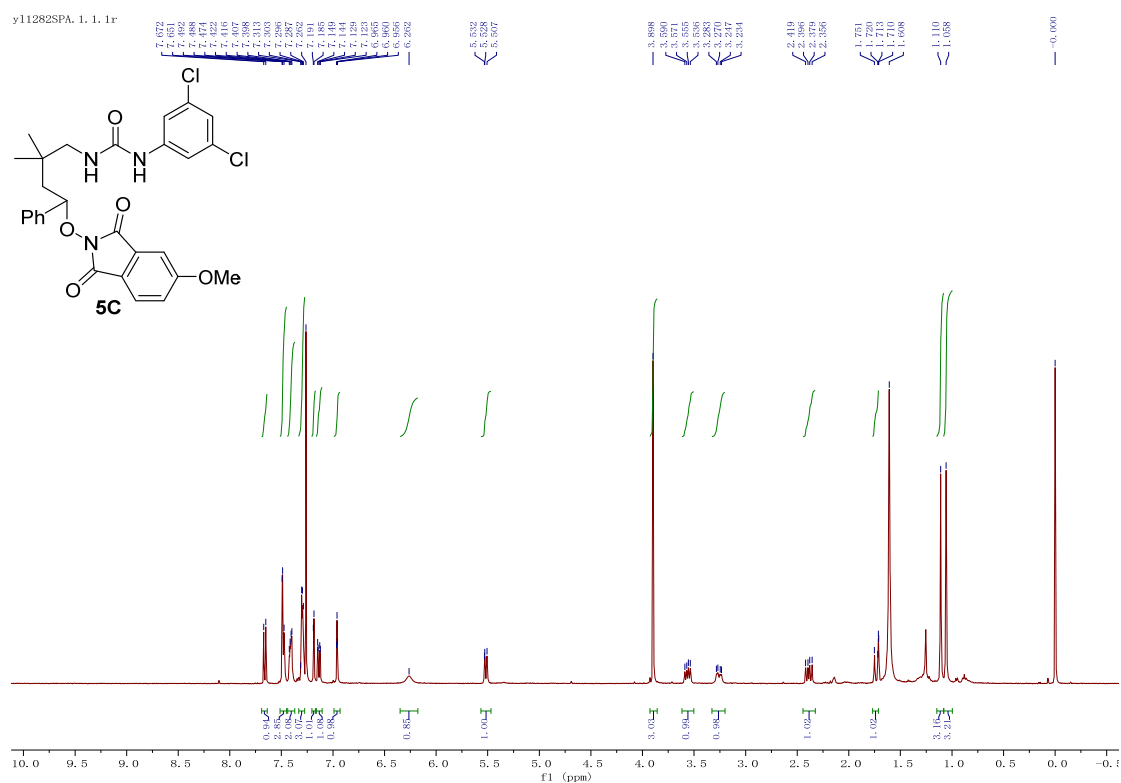
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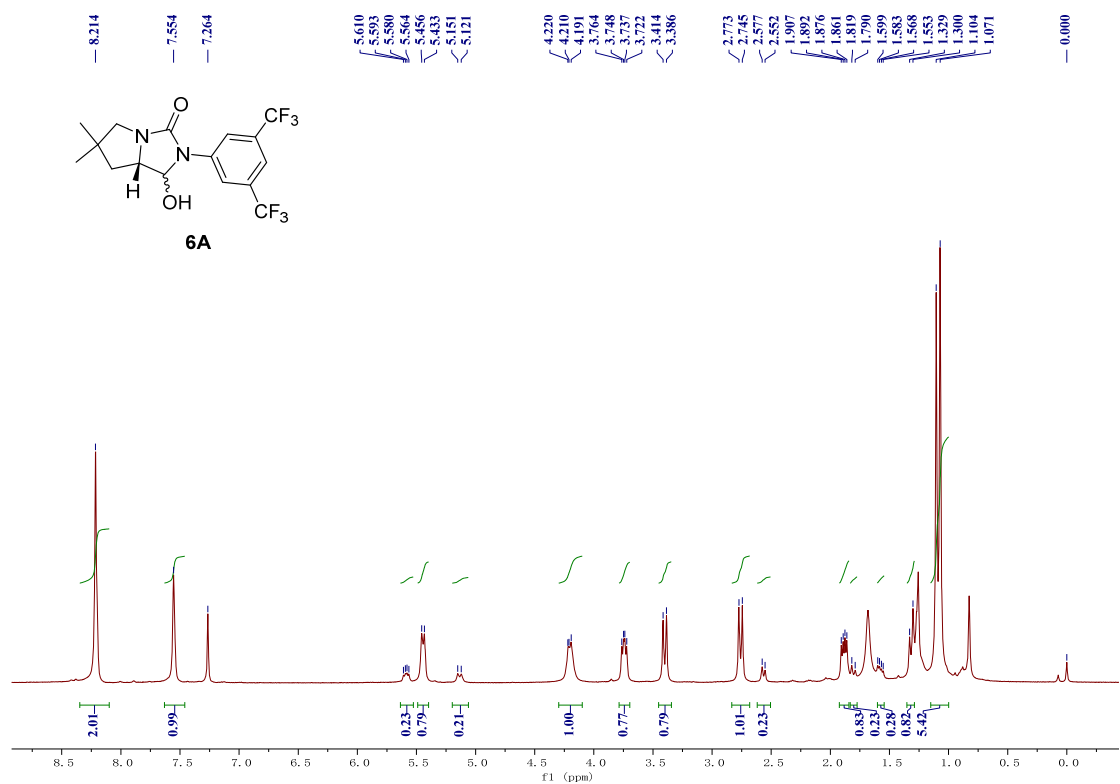
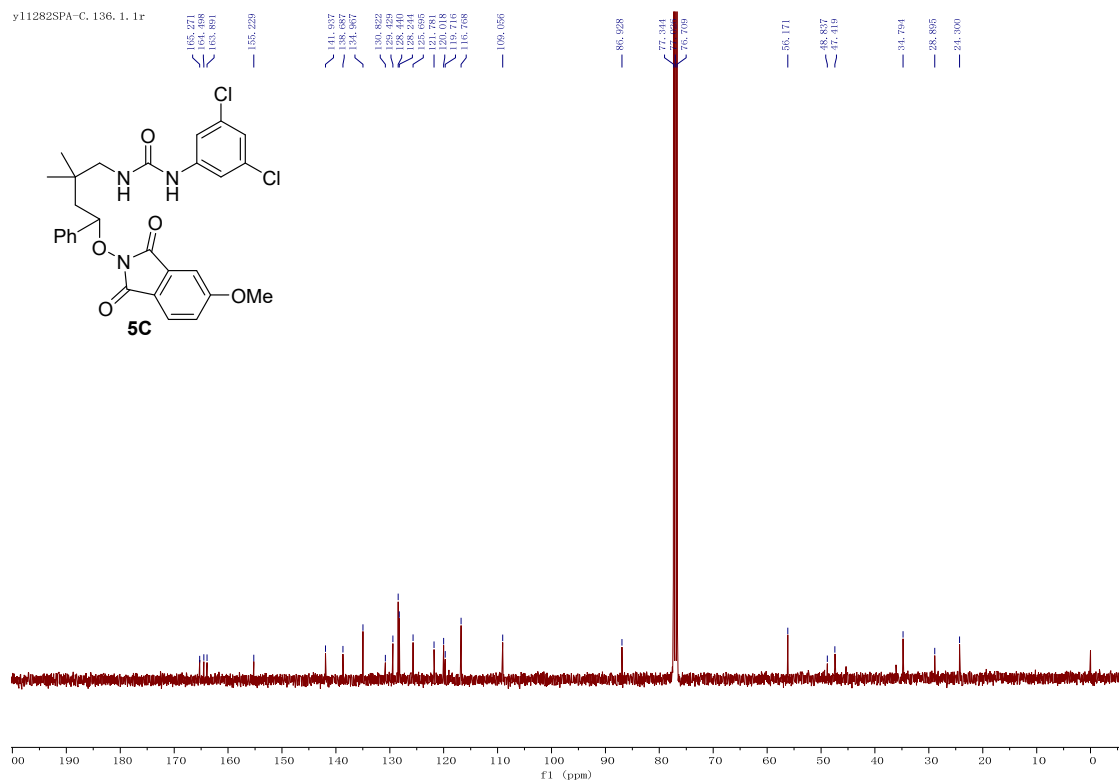
y119-1190PB/19

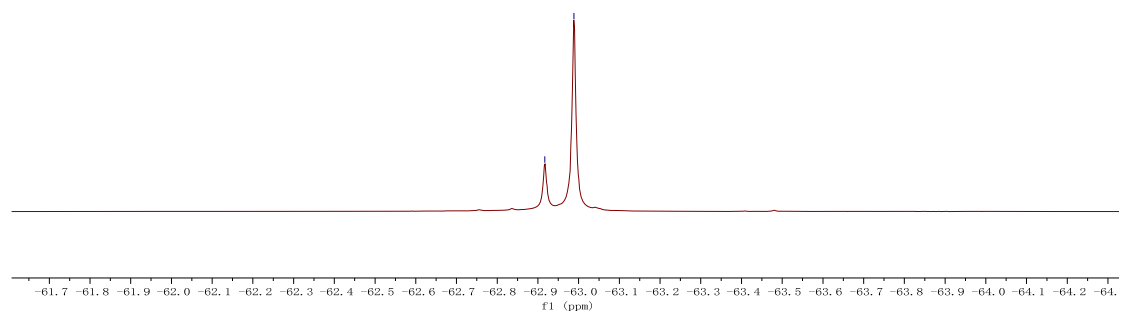
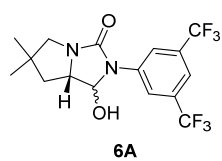
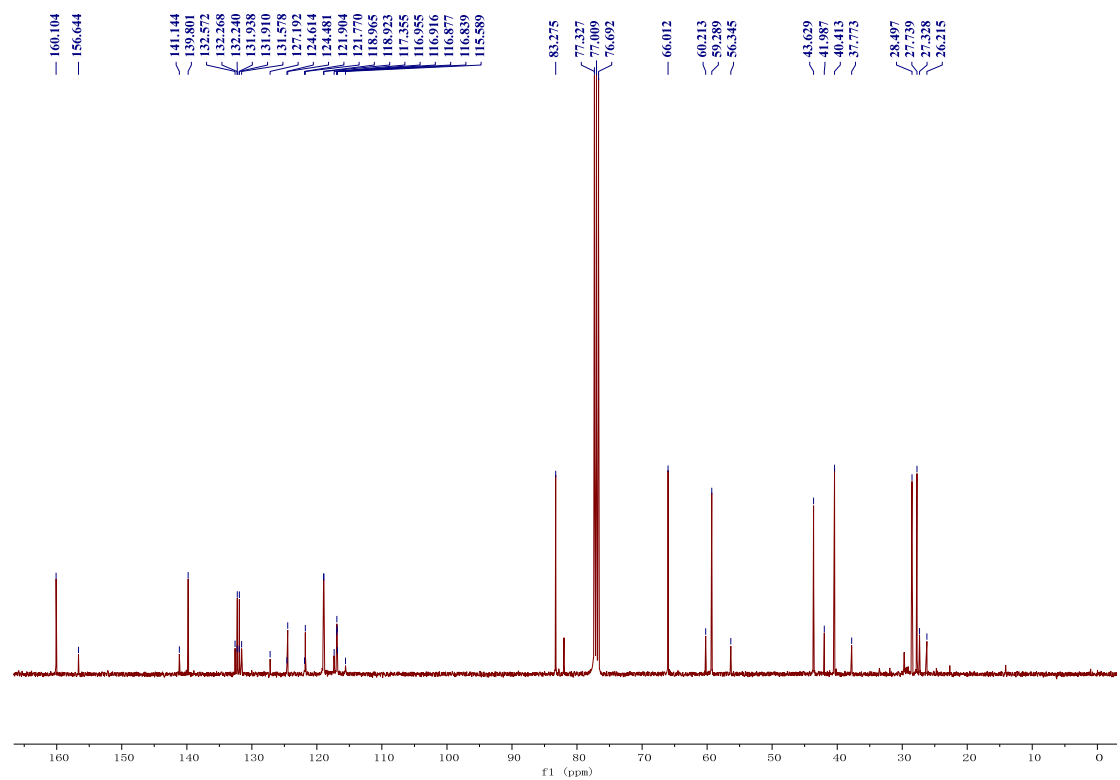


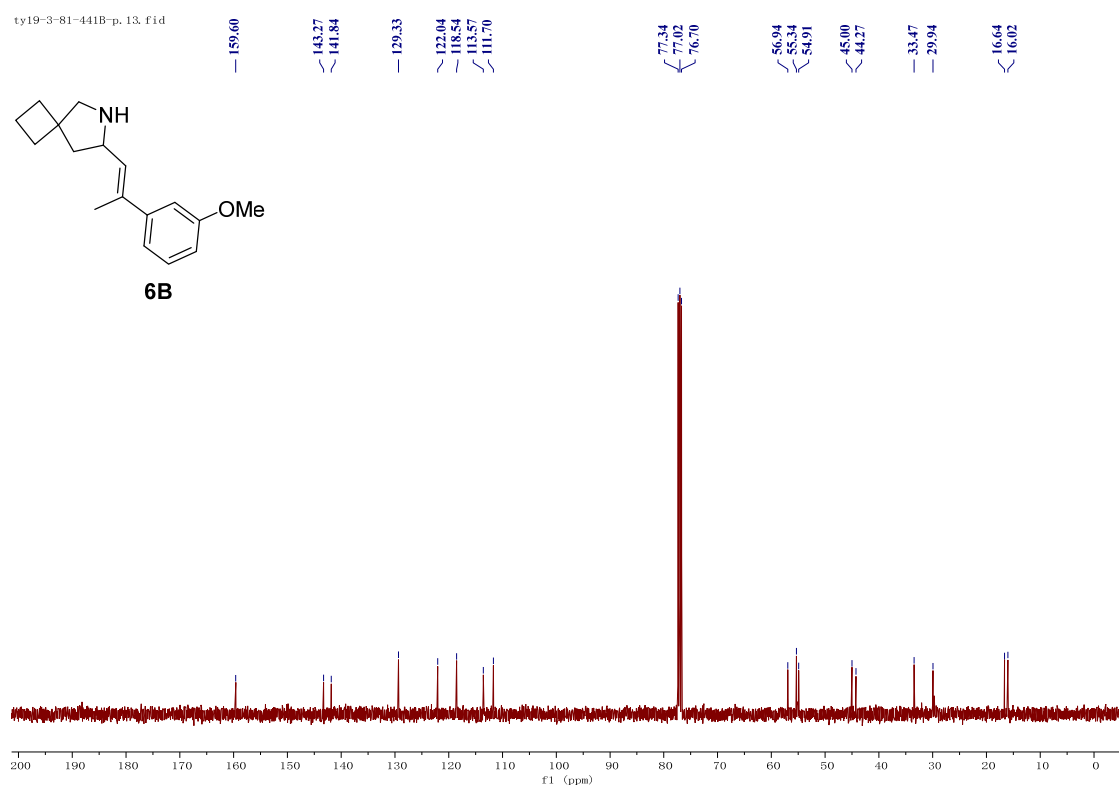
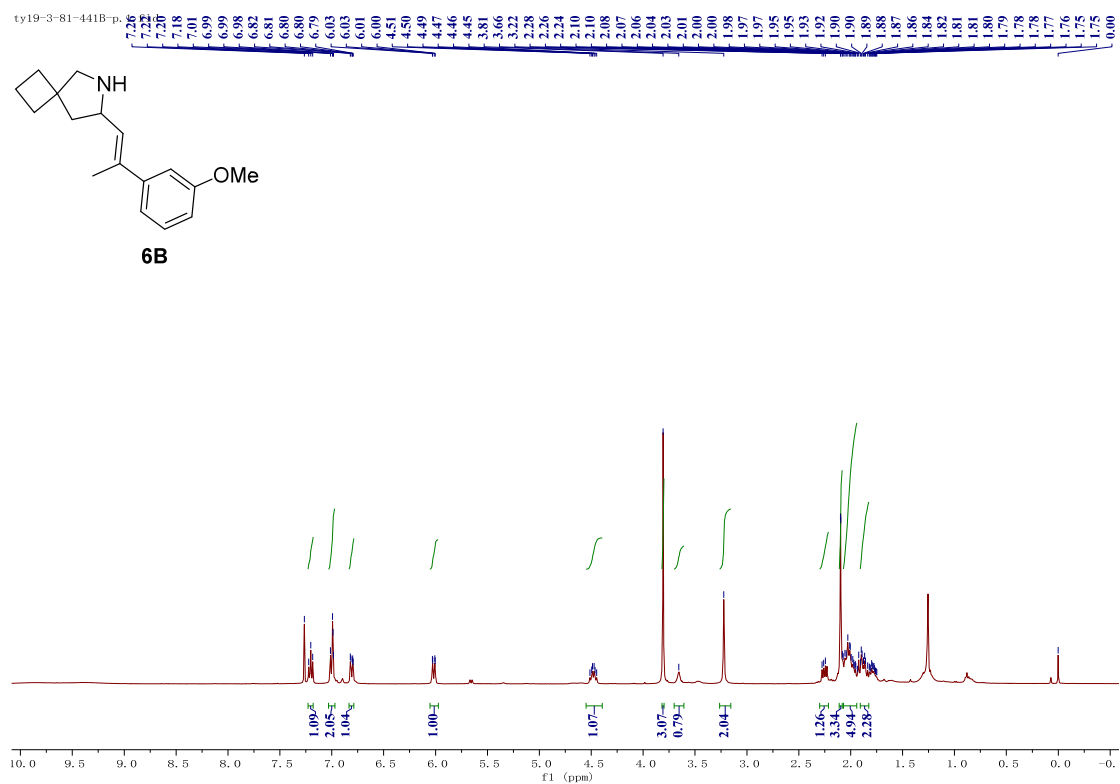
y11282SPA. 1. 1. 1r



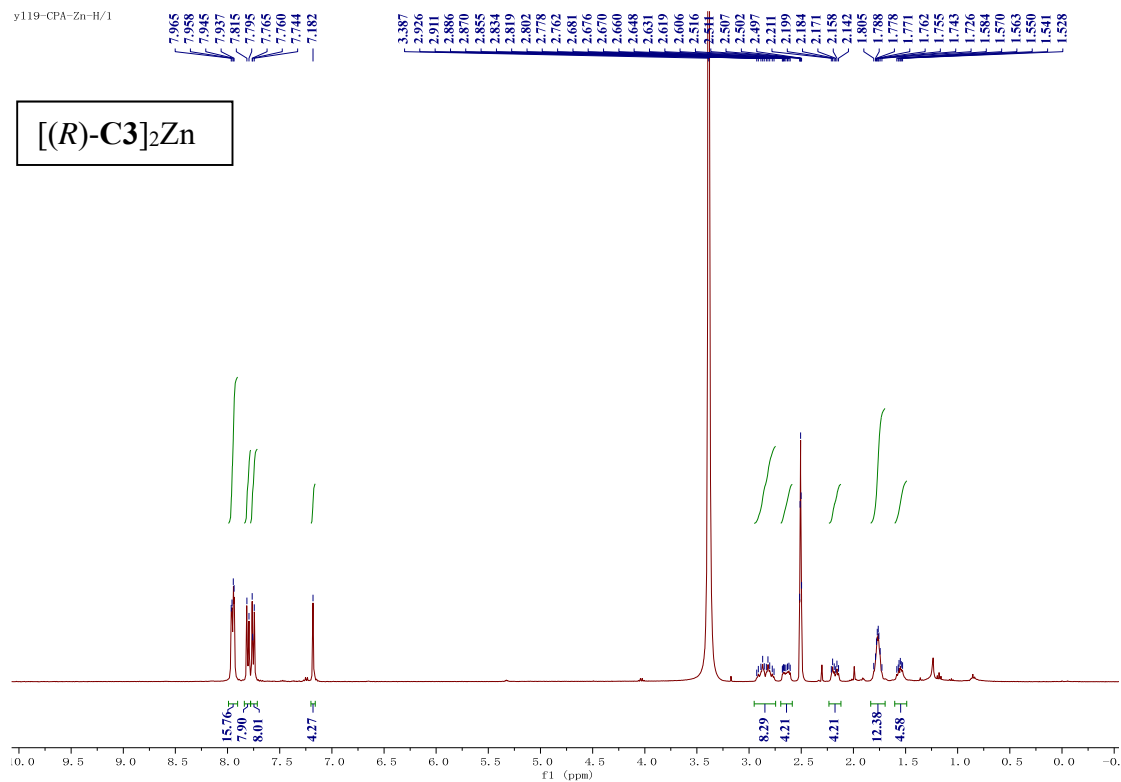
y11282SPA-C, 136, 1. 1r



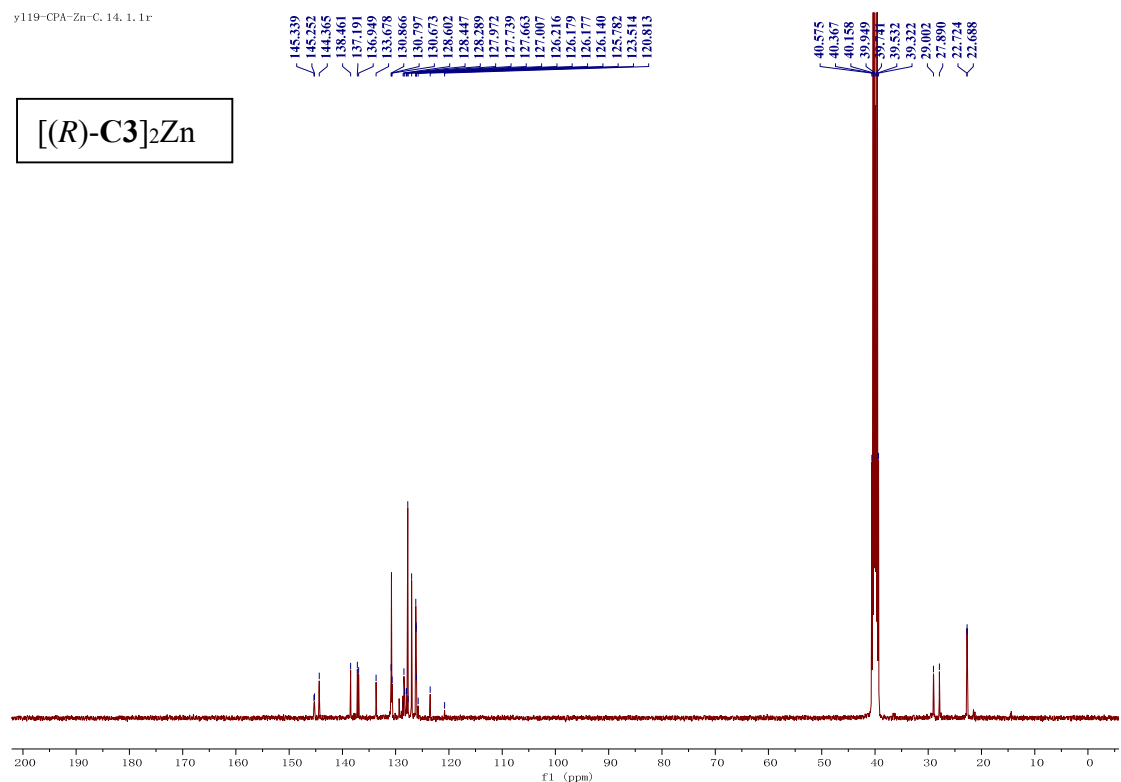




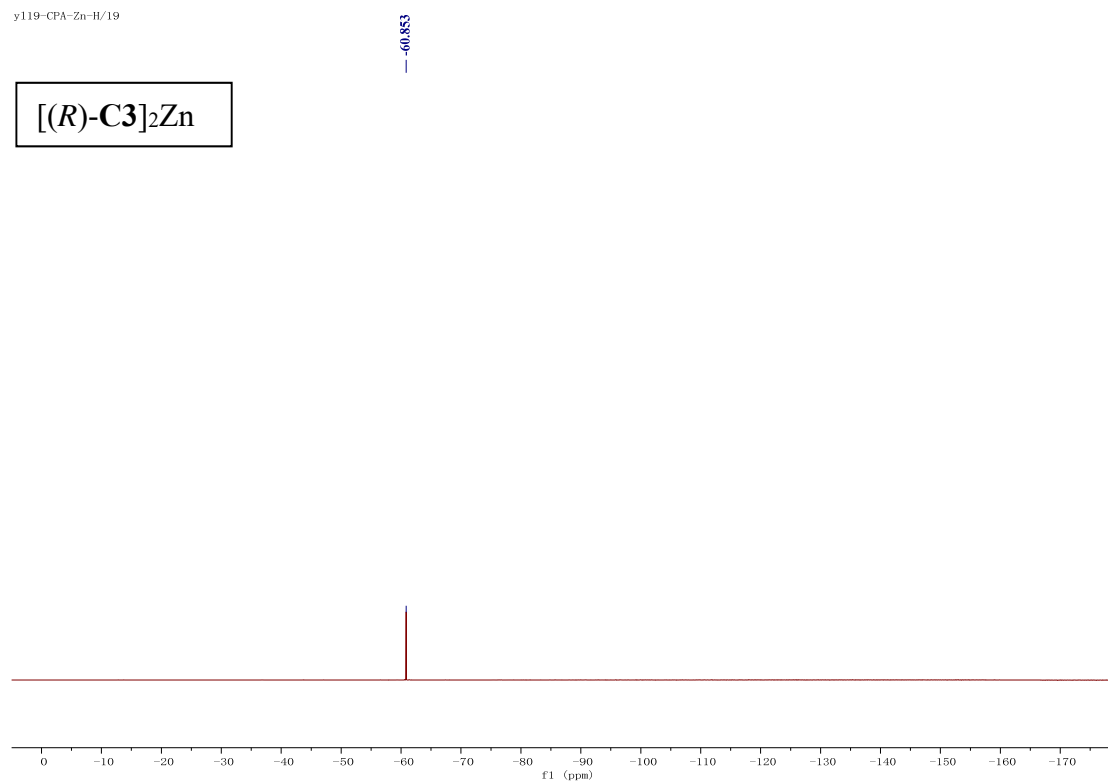
y119-CPA-Zn-H/1



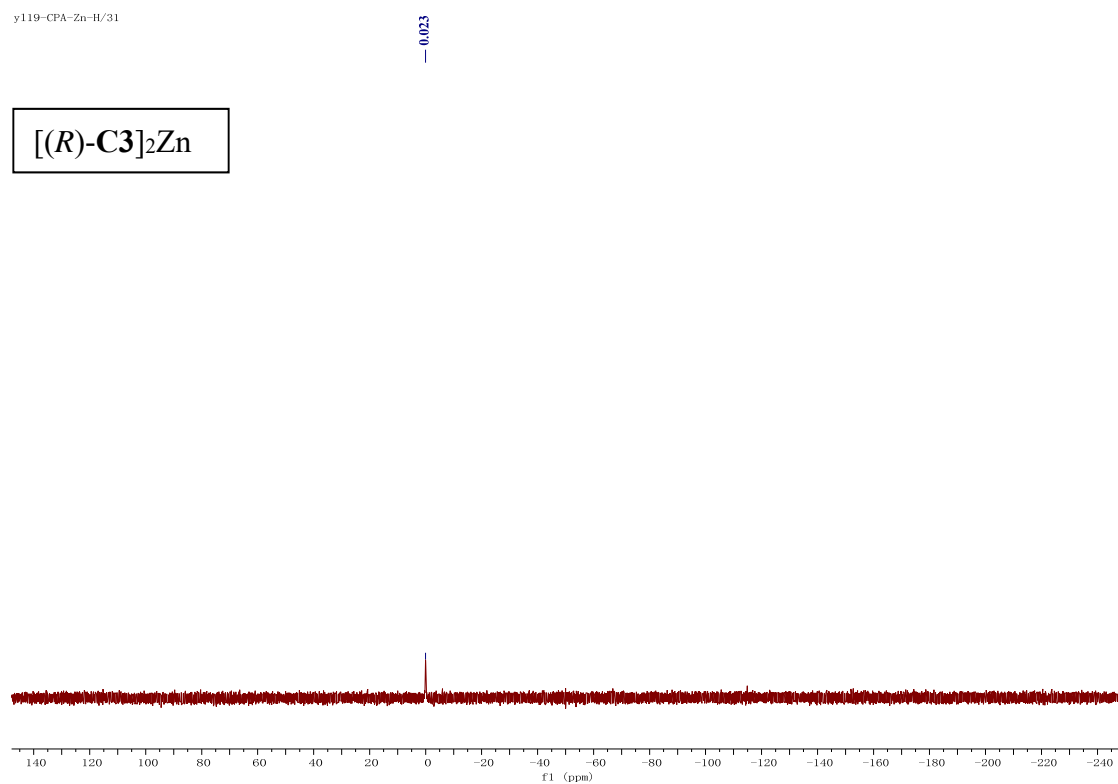
y119-CPA-Zn-C, 14, 1.1r



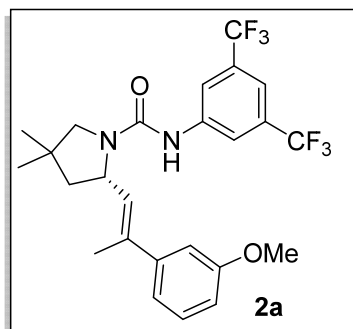
y119-CPA-Zn-H/19



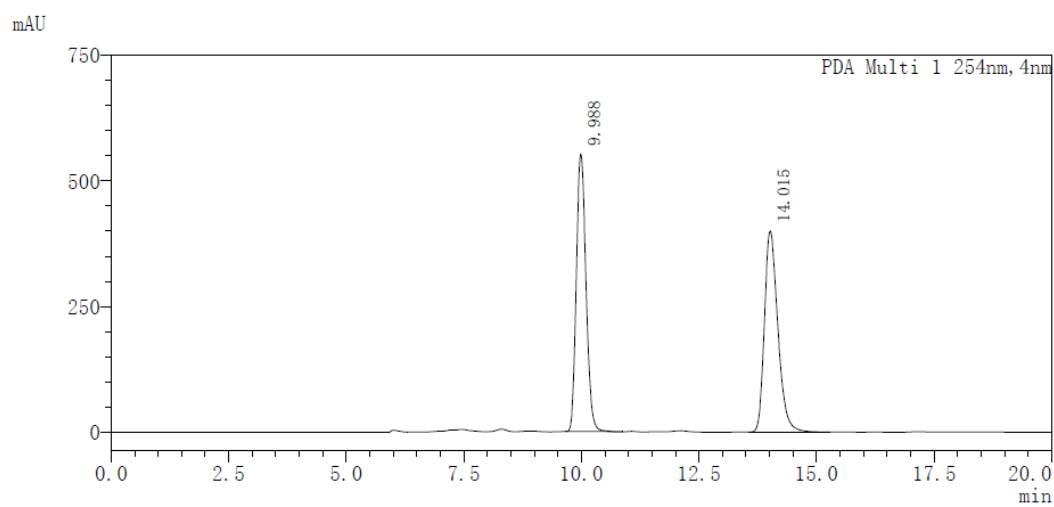
y119-CPA-Zn-H/31



HPLC Spectra

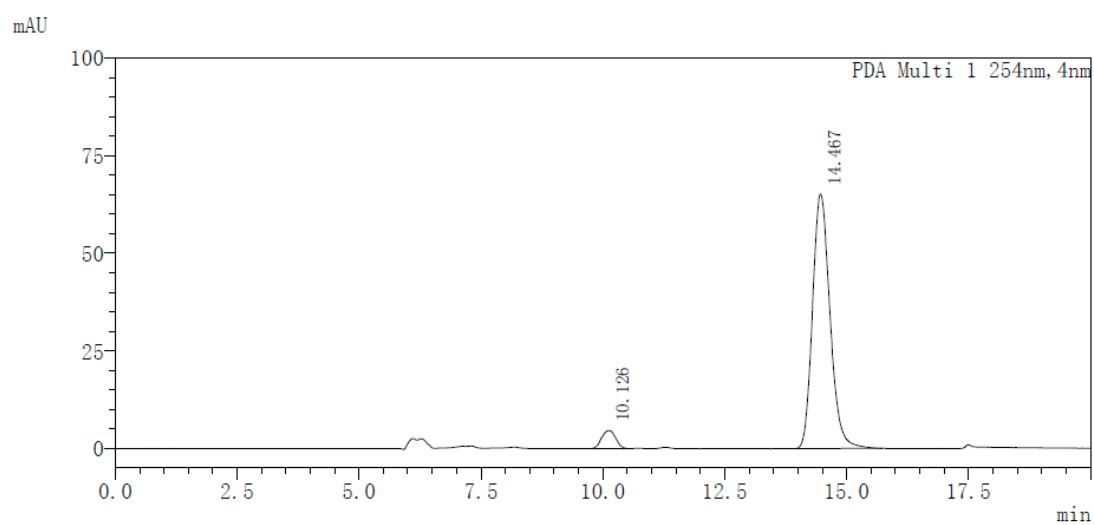


2a, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 0.5 mL/min. λ = 254 nm, t (minor) = 10.1 min, t (major) = 14.5 min, 95:5 er.



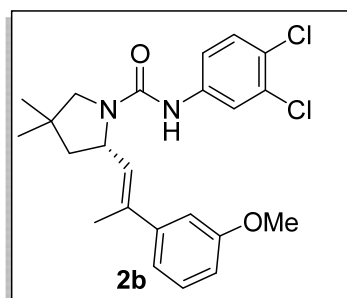
PDA Ch1 254nm

T	Hight	Area	Area%
9.988	551868	7912441	49.932
14.015	399549	7934126	50.068

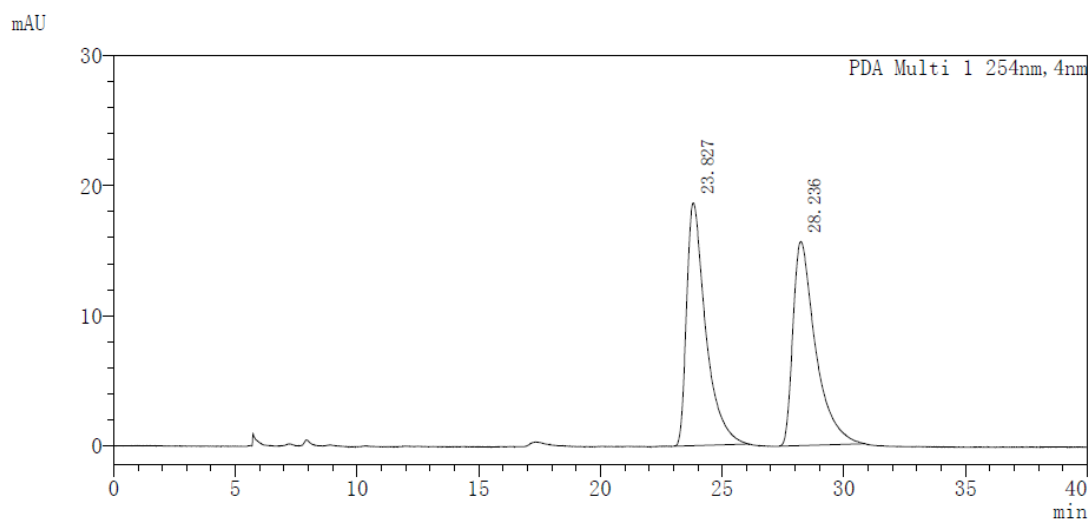


PDA Ch1 254nm

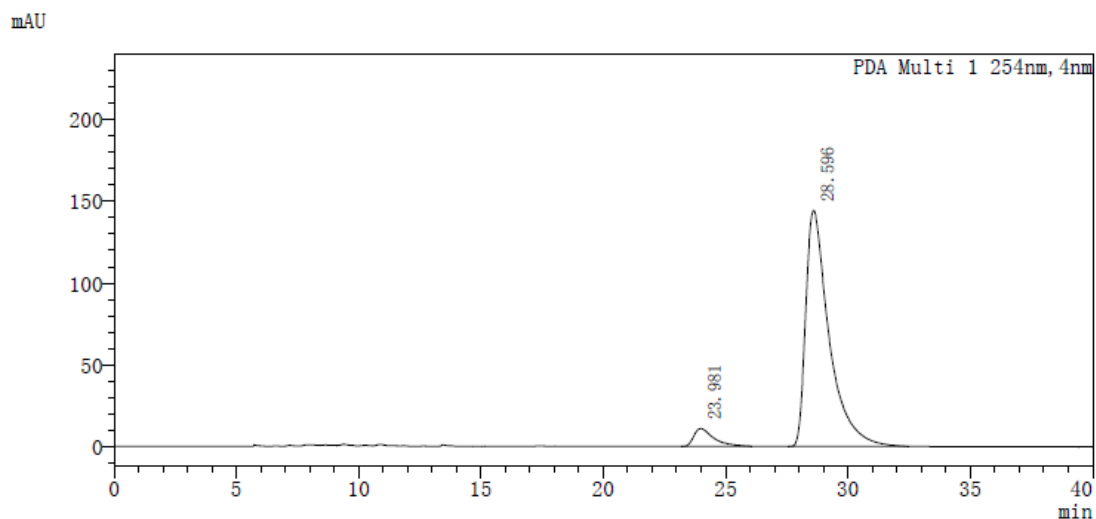
T	Hight	Area	Area%
10.126	4580	90626	5.277
14.467	65224	1626735	94.723



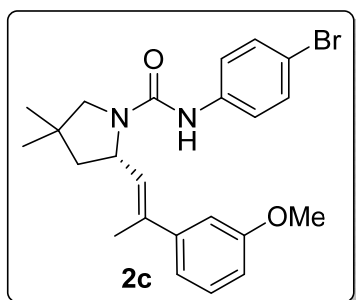
2b, HPLC condition: Chiralcel AD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 0.5 mL/min. λ = 254 nm, t (minor) = 24.0 min, t (major) = 28.6 min, 94:6 er.



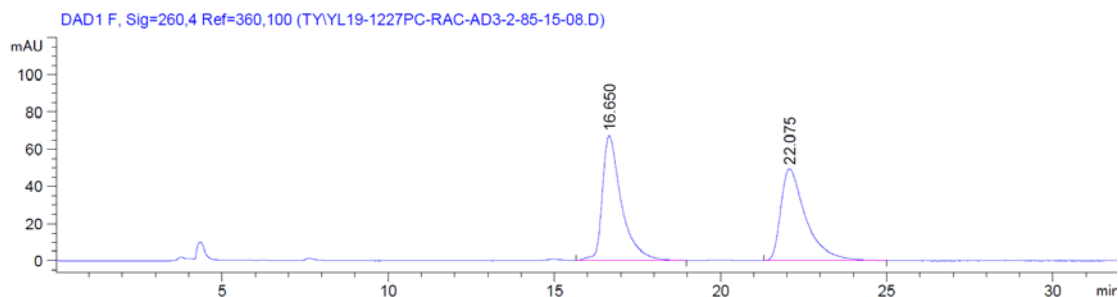
T	Hight	Area	Area%
23.827	18669	1020088	50.165
28.236	15684	1013372	49.835



T	Hight	Area	Area%
23.981	10910	606642	5.939
28.596	144406	9608206	94.061

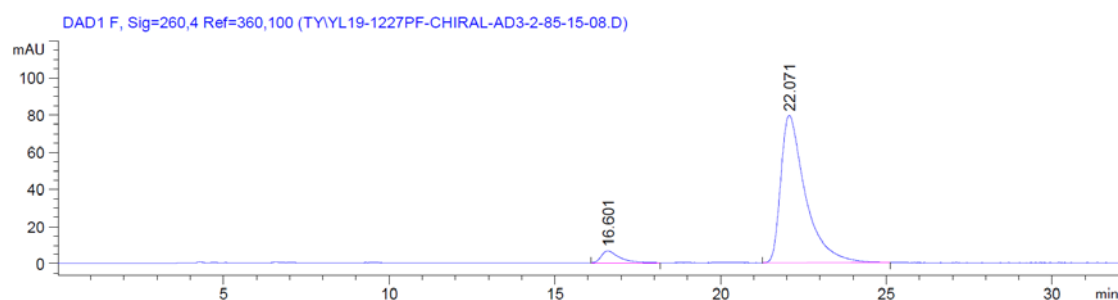


2c, HPLC condition: Chiralcel AD-3, *i*-PrOH/*n*-hexane = 85/15, flow rate 0.8 mL/min. λ = 260 nm, t (minor) = 16.6 min, t (major) = 22.0 min, 94:6 er.



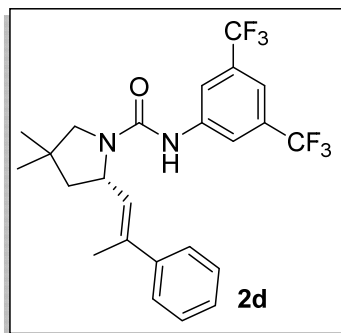
Signal 6: DAD1 F, Sig=260,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.650	BB	0.5759	2674.39111	66.97305	50.8674
2	22.075	BB	0.7511	2583.17969	49.32016	49.1326

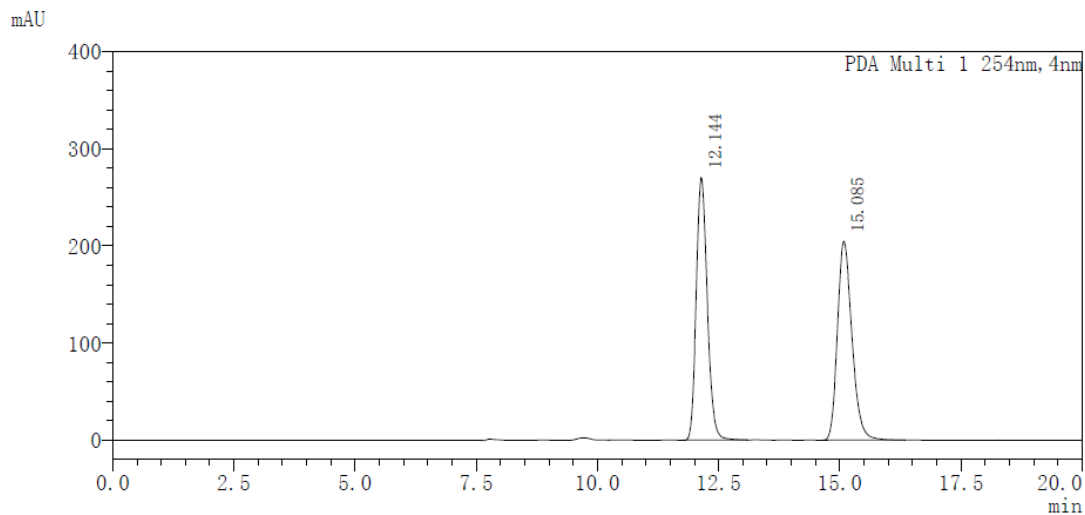


Signal 6: DAD1 F, Sig=260,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.601	BB	0.4759	244.58809	6.51709	5.6232
2	22.071	BB	0.7397	4105.05469	79.32757	94.3768

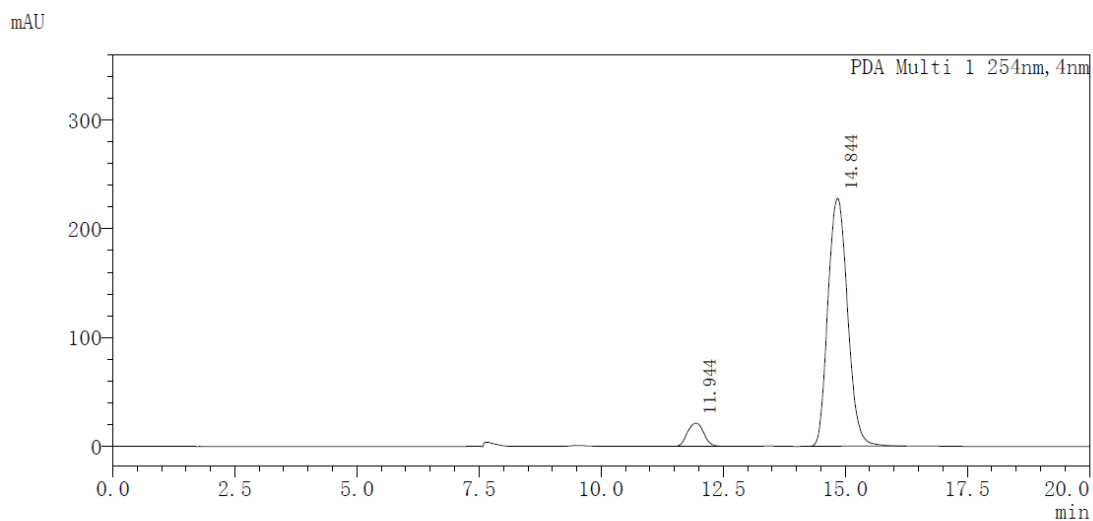


2d, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.4 mL/min. λ = 254 nm, t (minor) = 11.9 min, t (major) = 14.8 min, 93:7 er.



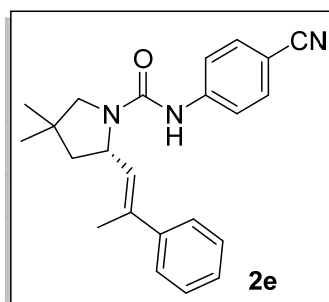
PDA Ch1 254nm

T	Hight	Area	Area%
12.144	270121	4283335	50.161
15.085	204525	4255854	49.839

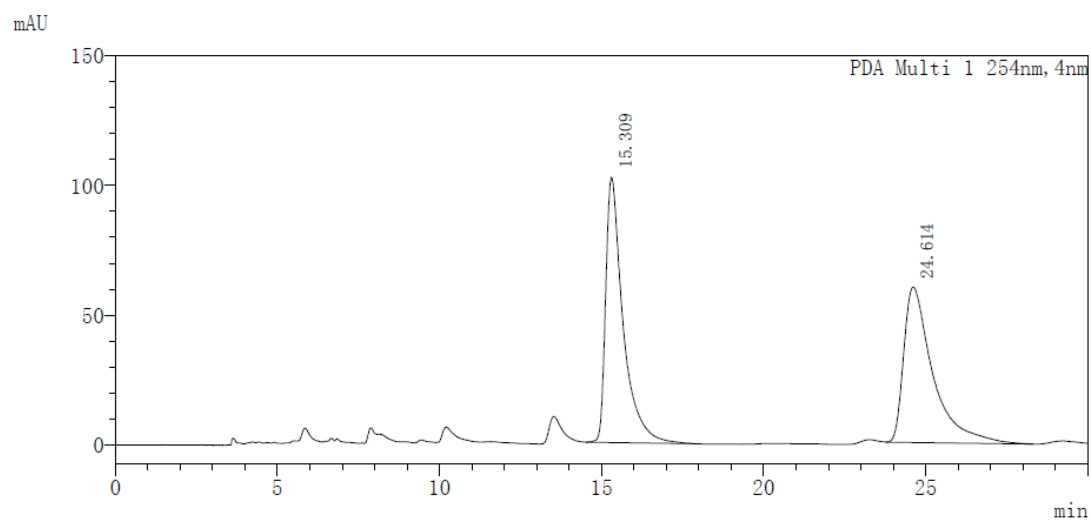


PDA Ch1 254nm

T	Hight	Area	Area%
11.944	21135	489675	7.123
14.844	227950	6385142	92.877

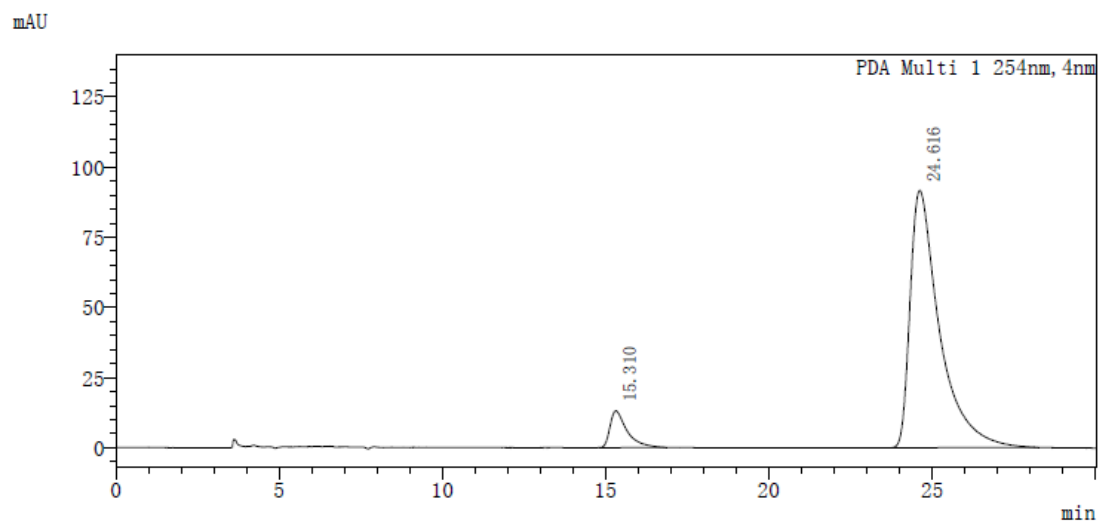


2e, HPLC condition: Chiralcel AD-3, *i*-PrOH/*n*-hexane = 85/15, flow rate 0.8 mL/min. λ = 254 nm, t (minor) = 15.3 min, t (major) = 24.6 min, 92.5:7.5 er.



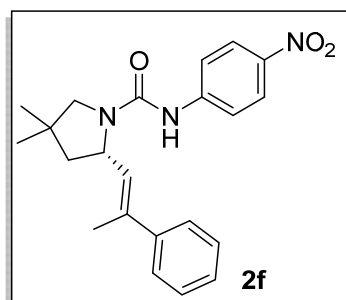
PDA Ch1 254nm

T	Hight	Area	Area%
15.309	102170	3809712	50.487
24.614	59873	3736153	49.513



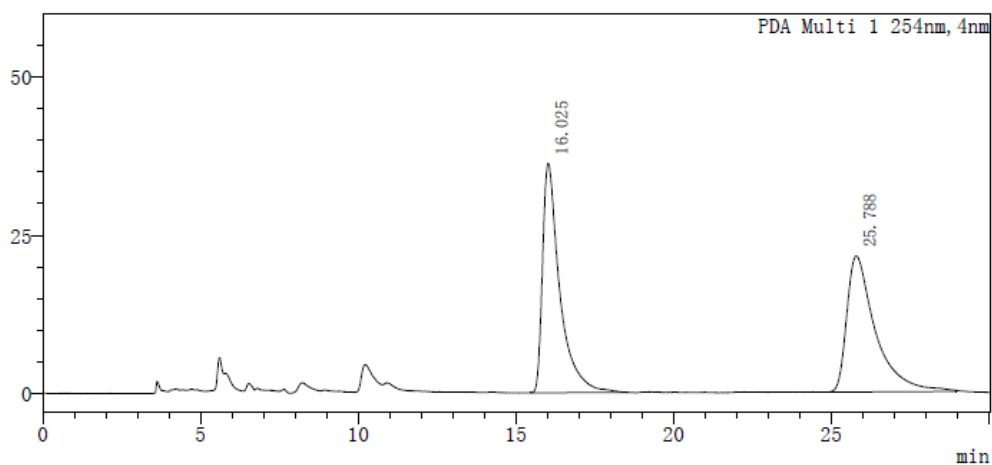
PDA Ch1 254nm

T	Hight	Area	Area%
15.310	13097	472478	7.600
24.616	91729	5744280	92.400



2f, HPLC condition: Chiralcel AD-3, *i*-PrOH/*n*-hexane = 85/15, flow rate 0.8 mL/min. λ = 254 nm, *t*(minor) = 16.0 min, *t*(major) = 25.8 min, 91:9 er.

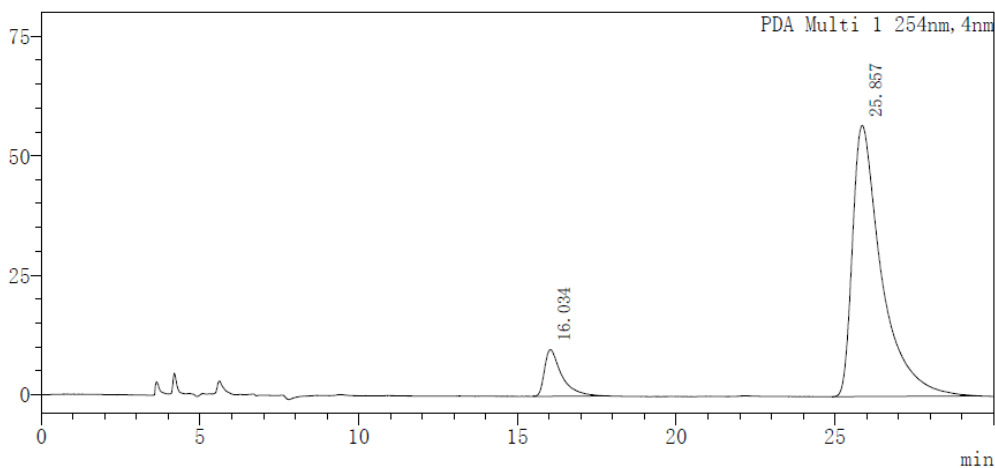
mAU



PDA Ch1 254nm

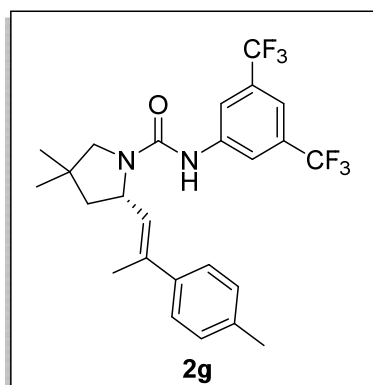
T	Hight	Area	Area%
16.025	36284	1378016	50.265
25.788	21530	1363481	49.735

mAU



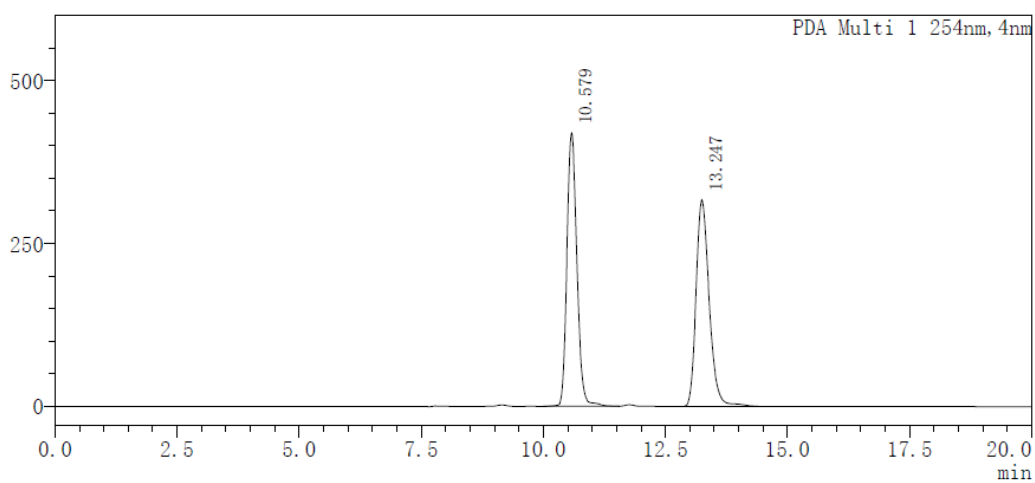
PDA Ch1 254nm

T	Hight	Area	Area%
16.034	9756	364084	9.178
25.857	56719	3602745	90.822



2g, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.4 mL/min. λ = 254 nm, t (minor) = 10.4 min, t (major) = 13.0 min, 92:8 er.

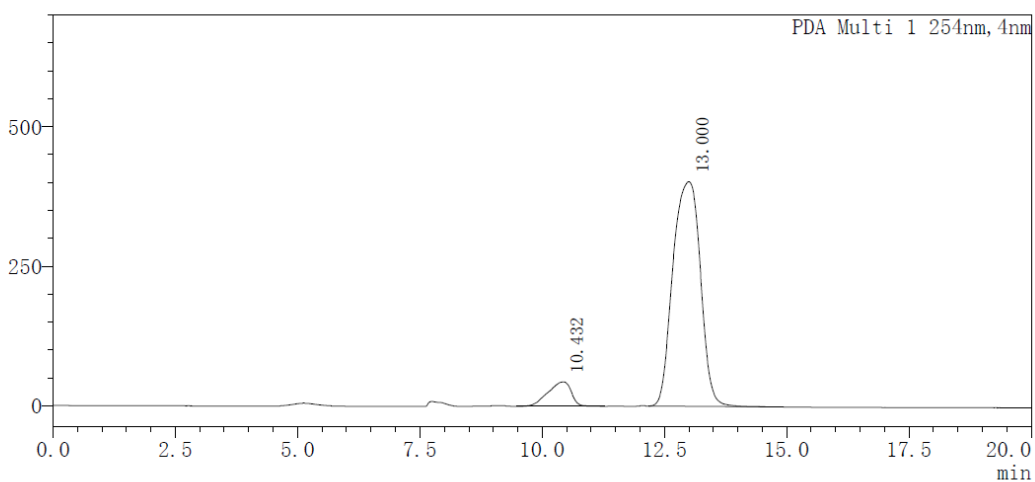
mAU



PDA Ch1 254nm

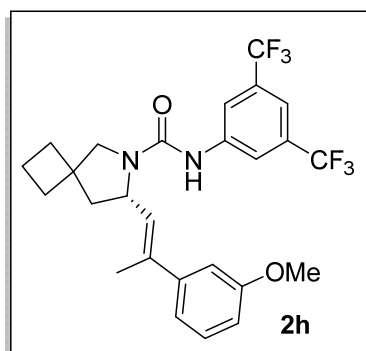
T	Hight	Area	Area%
10.579	419006	5964992	50.055
13.247	317383	5951905	49.945

mAU

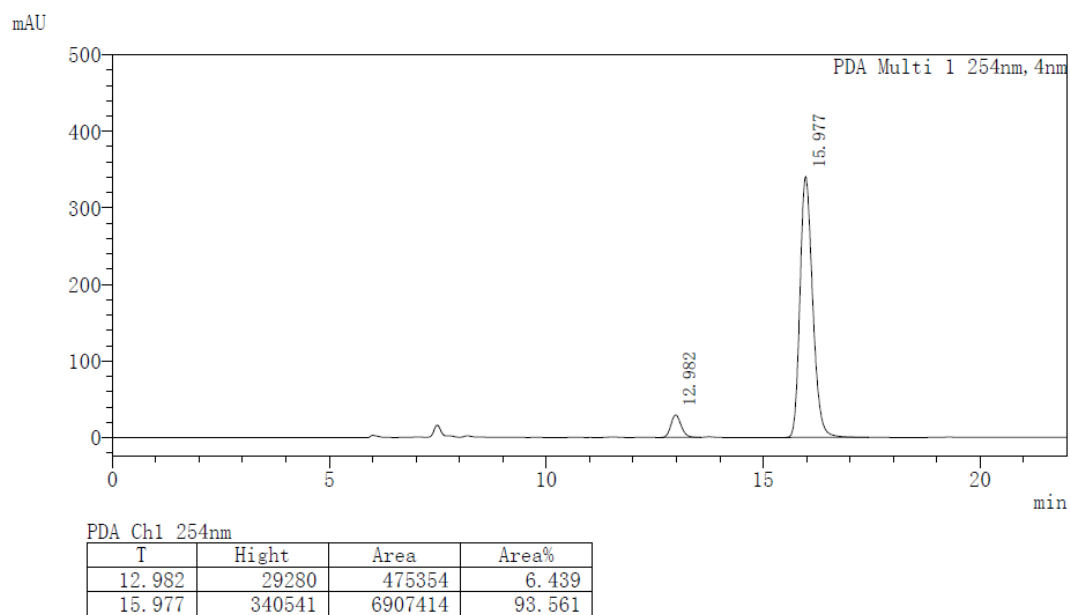
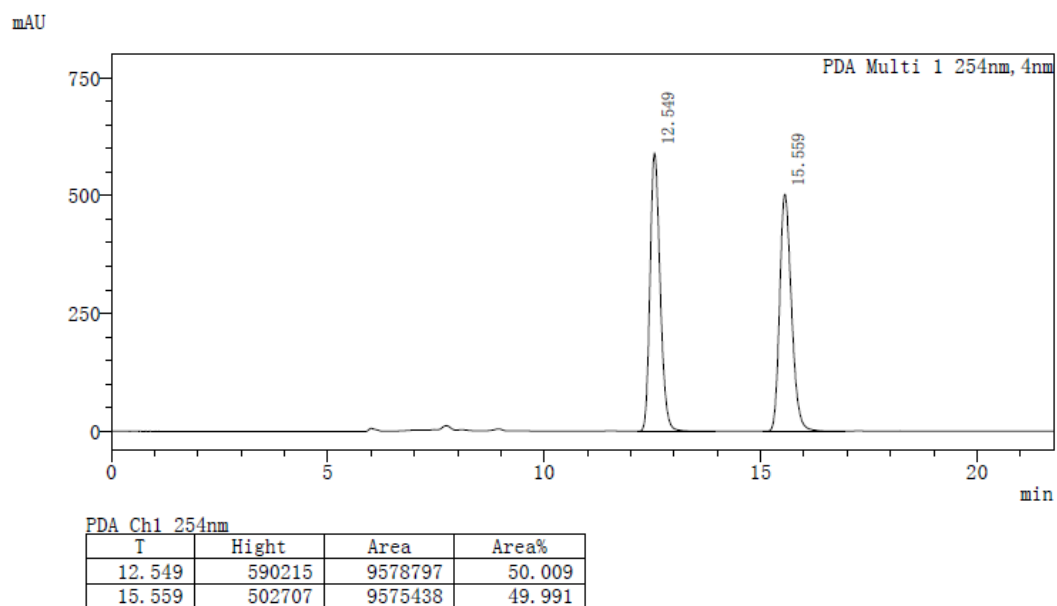


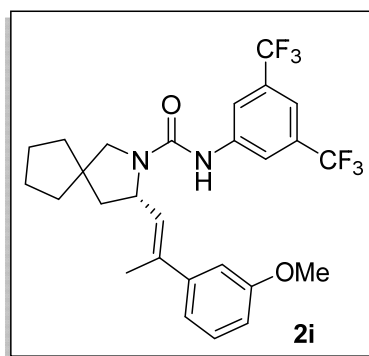
PDA Ch1 254nm

T	Hight	Area	Area%
10.432	43717	1445224	8.393
13.000	402421	15773169	91.607

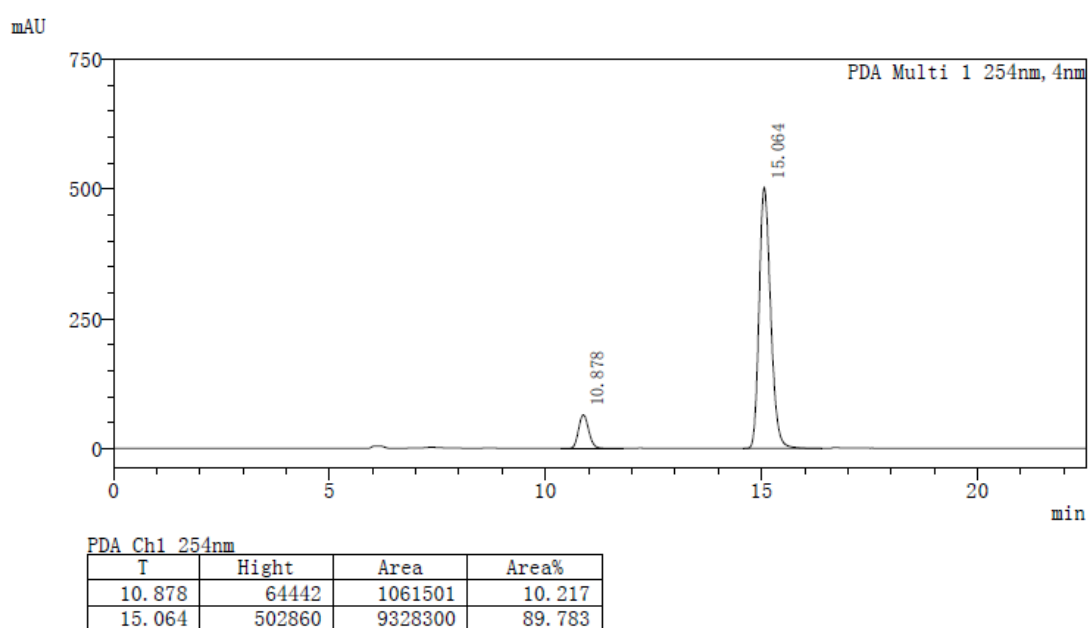
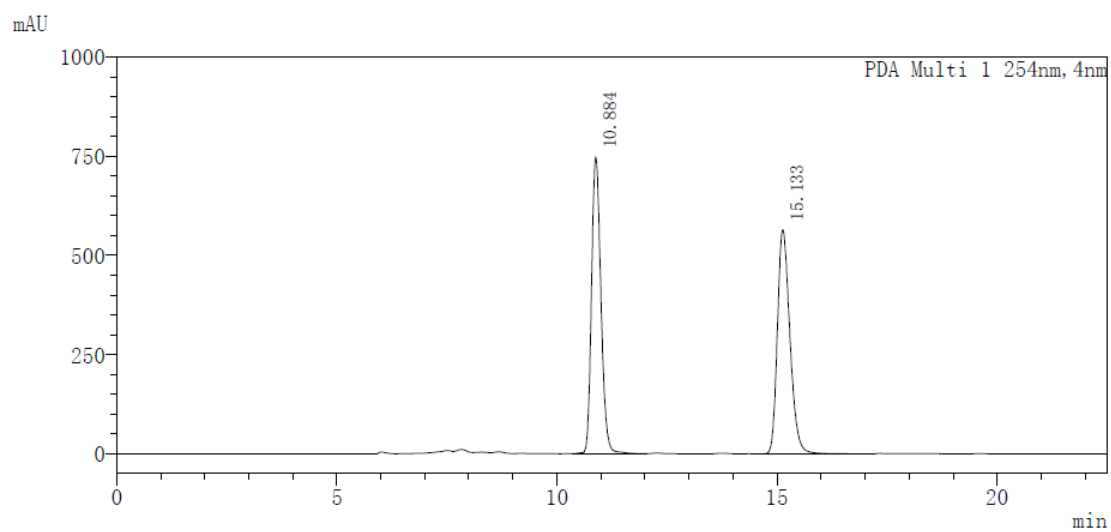


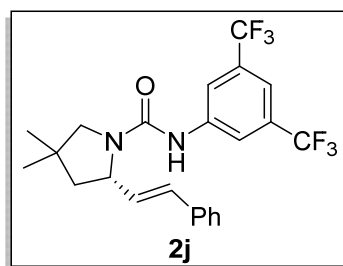
2h, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 0.5 mL/min. λ = 254 nm, t (minor) = 13.0 min, t (major) = 16.0 min, 94:6 er



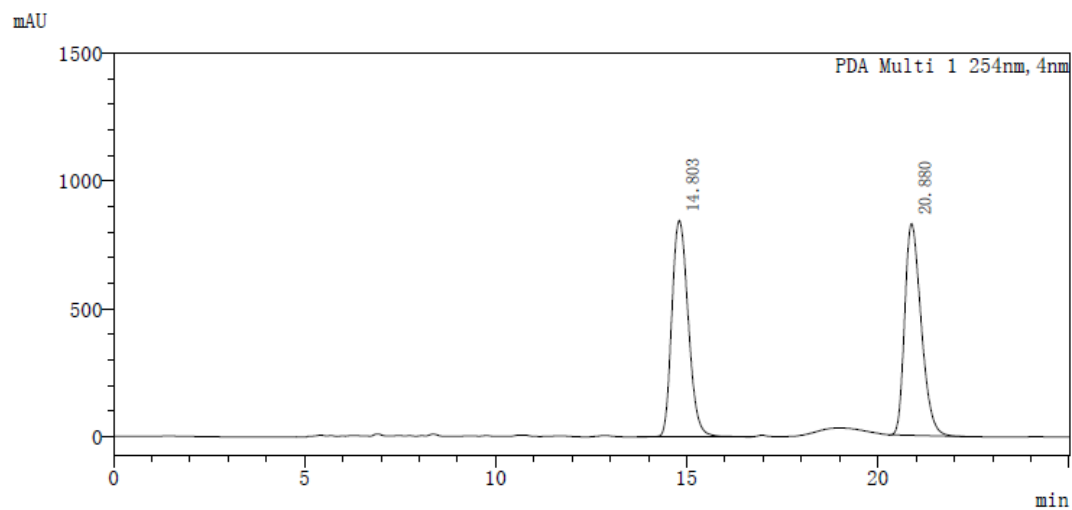


2i, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 0.5 mL/min. λ = 254 nm, t (minor) = 10.9 min, t (major) = 15.1 min, 90:10 er.

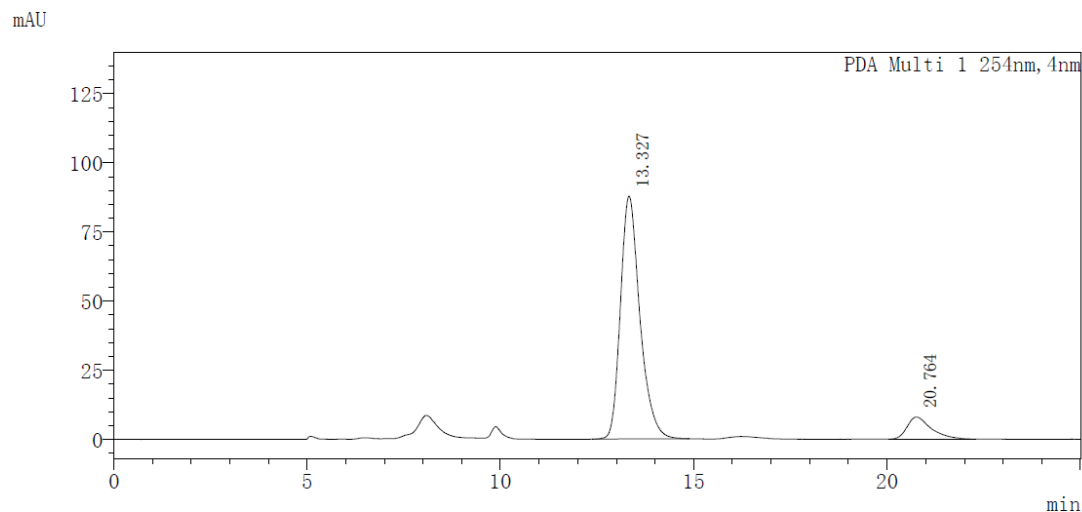




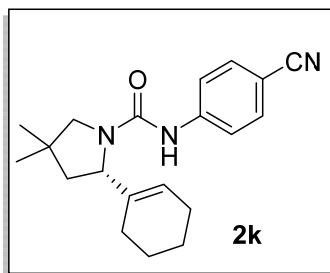
2j, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.6 mL/min. λ = 254 nm, t (major) = 13.3 min, t (minor) = 20.8 min, 90:10 er.



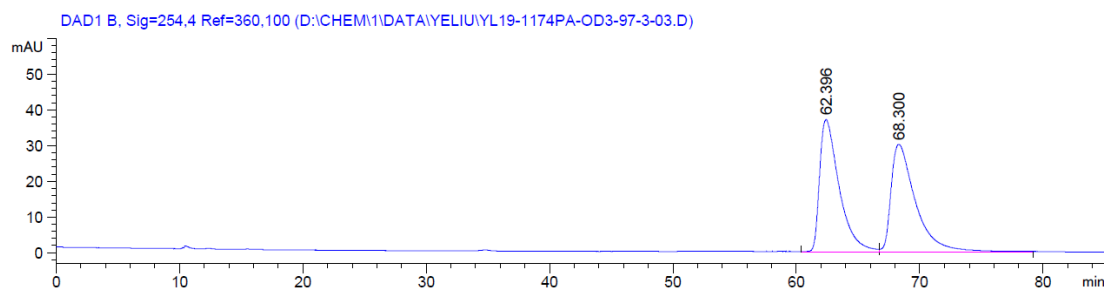
PDA Ch1 254nm			
T	Hight	Area	Area%
14.803	847099	25327353	50.445
20.880	828426	24880442	49.555



PDA Ch1 254nm			
T	Hight	Area	Area%
13.327	87825	3077339	89.993
20.764	7960	342186	10.007

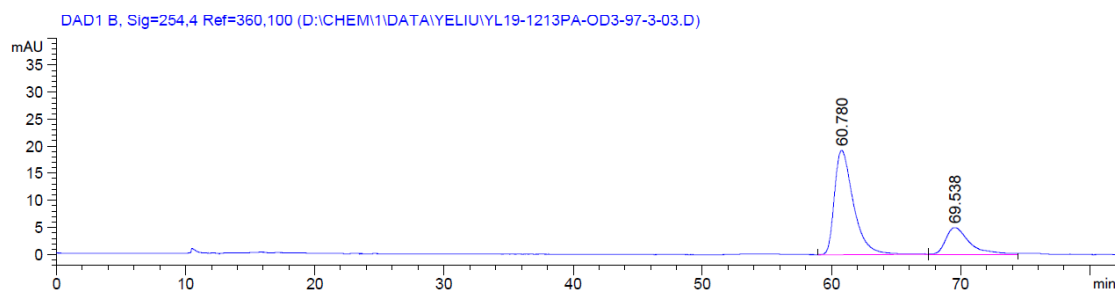


2k, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane =97/3, flow rate 0.3 mL/min. λ = 254 nm, *t*(major) =60.8 min, *t*(minor) = 69.5 min, 76:24 er.



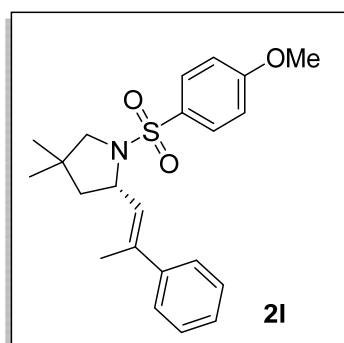
Signal 2: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	62.396	BV	1.6440	4048.45679	36.89947	49.6989
2	68.300	VB	2.0164	4097.51758	30.02031	50.3011



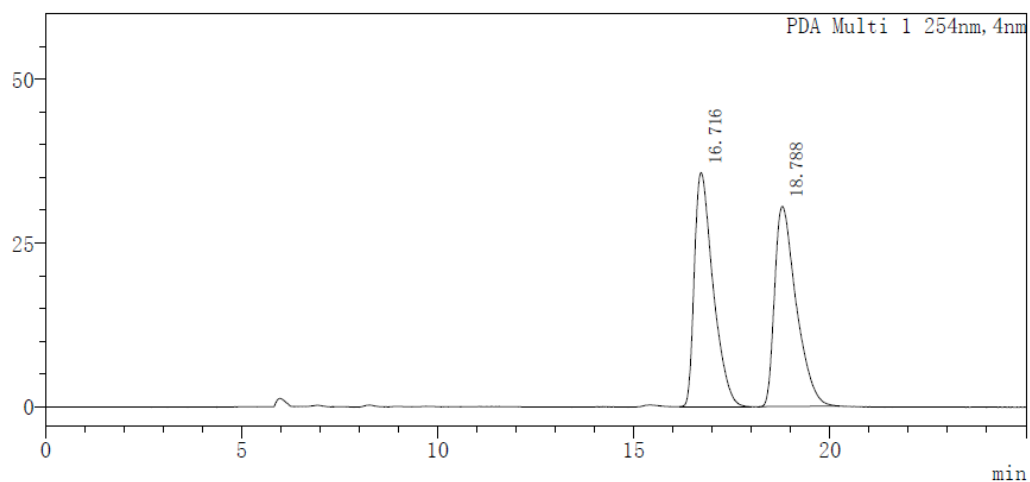
Signal 2: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	60.780	BV	1.5394	1974.55103	19.27051	75.5991
2	69.538	VV	1.8411	637.32129	4.96671	24.4009



2I, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 0.5 mL/min. λ = 254 nm, t(major) = 16.0 min, t(minor) = 18.4 min, 87:13 er.

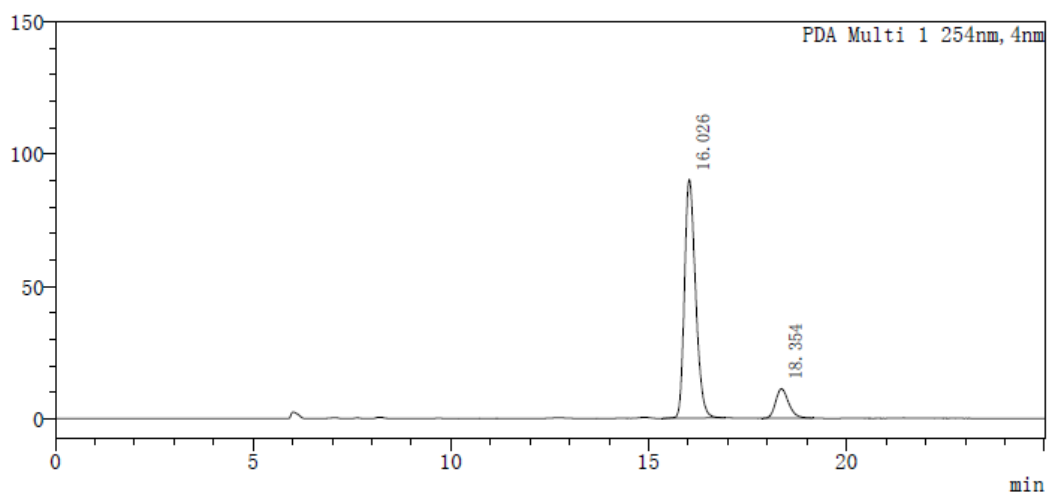
mAU



PDA Ch1 254nm

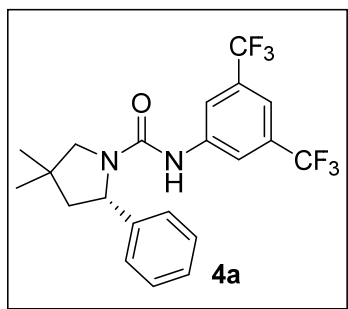
T	Hight	Area	Area%
16.716	35676	1186252	50.294
18.788	30549	1172386	49.706

mAU

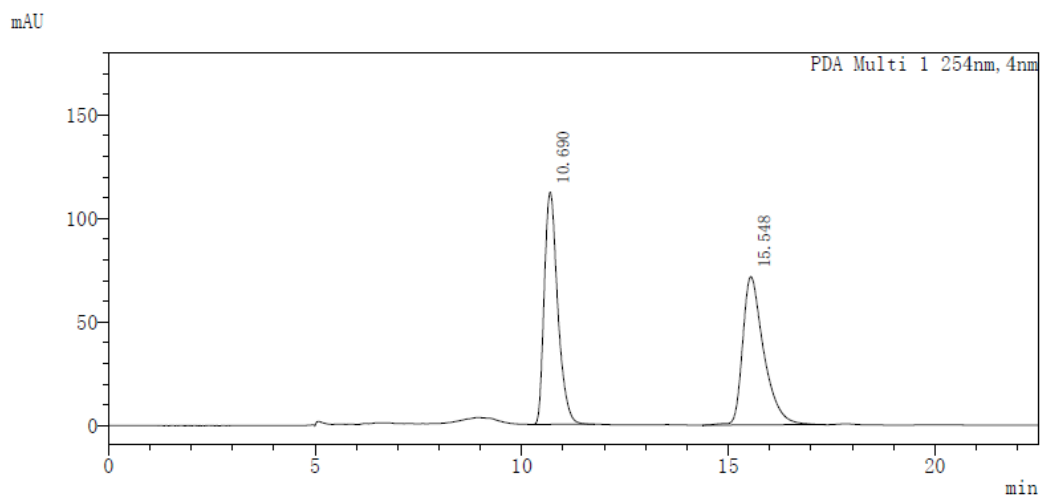


PDA Ch1 254nm

T	Hight	Area	Area%
16.026	90181	1761082	87.395
18.354	11106	253992	12.605

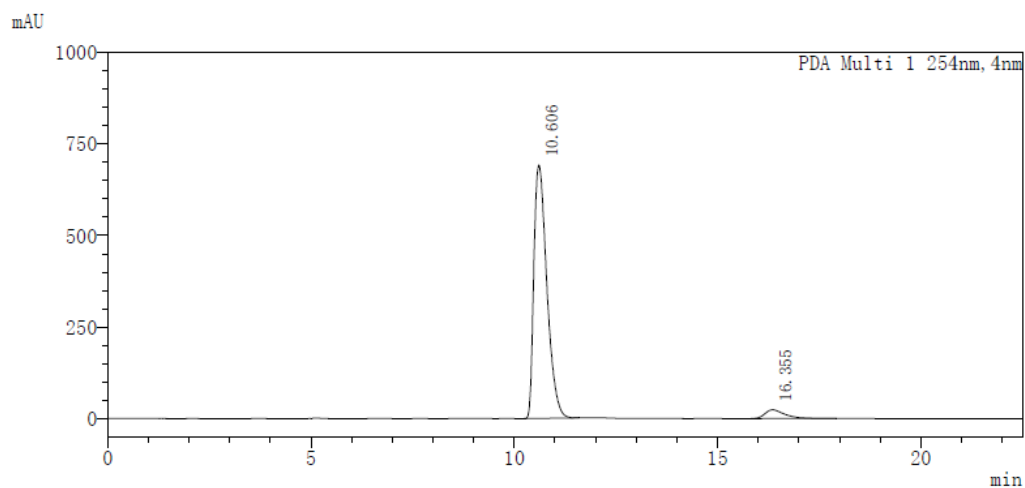


4a HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.6 mL/min. λ = 254 nm, t (major) = 10.6 min, t (minor) = 16.4 min, 95:5 er.



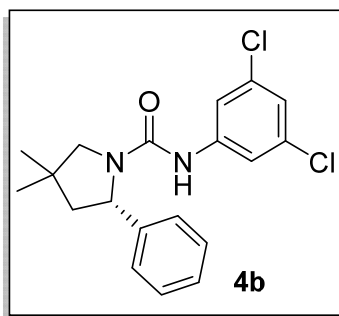
PDA Ch1 254nm

T	Hight	Area	Area%
10.690	112186	2541789	49.895
15.548	71624	2552516	50.105

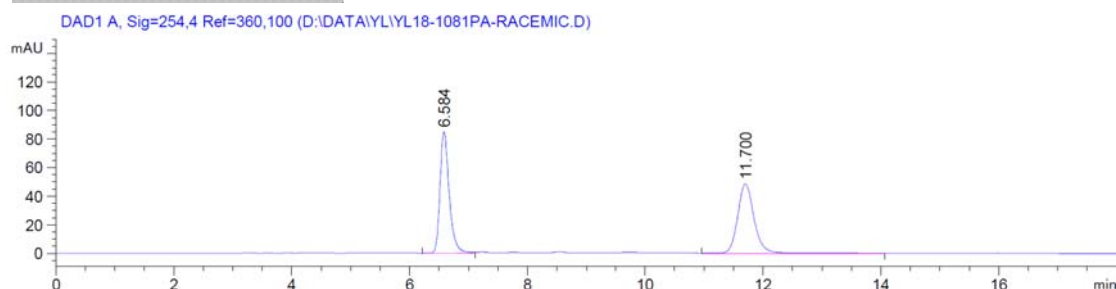


PDA Ch1 254nm

T	Hight	Area	Area%
10.606	691696	15377548	95.047
16.355	23674	801387	4.953

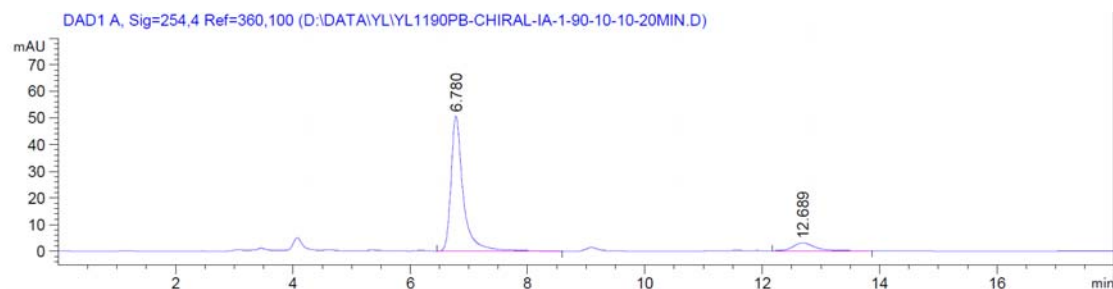


4b, HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 90/10, flow rate 1.0 mL/min. λ = 254 nm, t (major) = 6.8 min, t (minor) = 12.7 min, 91:9 er.



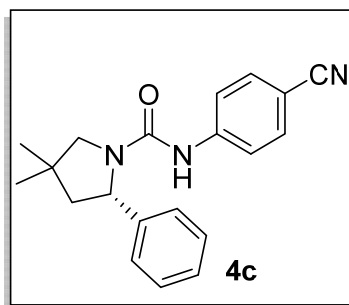
Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.584	BV	0.1622	921.12390	85.15656	49.8263
2	11.700	BV	0.2945	927.54657	48.63167	50.1737



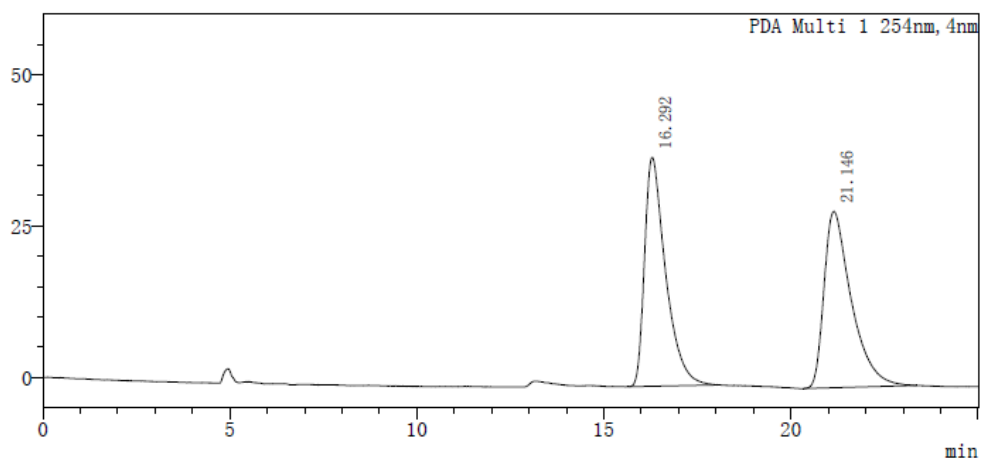
Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	6.780	BB	0.2119	736.43317	50.55672	90.8121
2	12.689	BB	0.3787	74.50823	2.93006	9.1879



4c, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 85/15, flow rate 0.6 mL/min. λ = 254 nm, t (major) = 16.1 min, t (minor) = 20.8 min, 92:8 er.

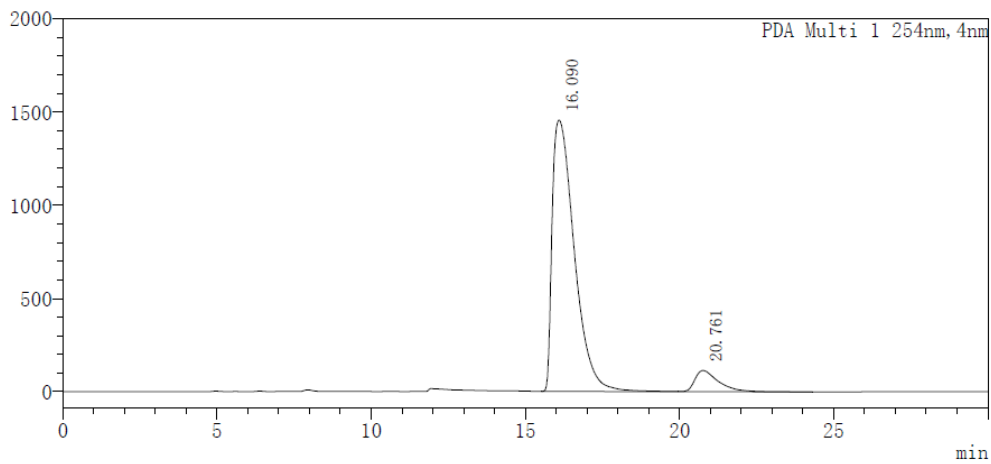
mAU



PDA Ch1 254nm

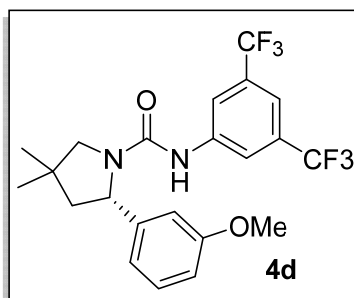
T	Hight	Area	Area%
16.292	37797	1483007	50.054
21.146	29106	1479782	49.946

mAU

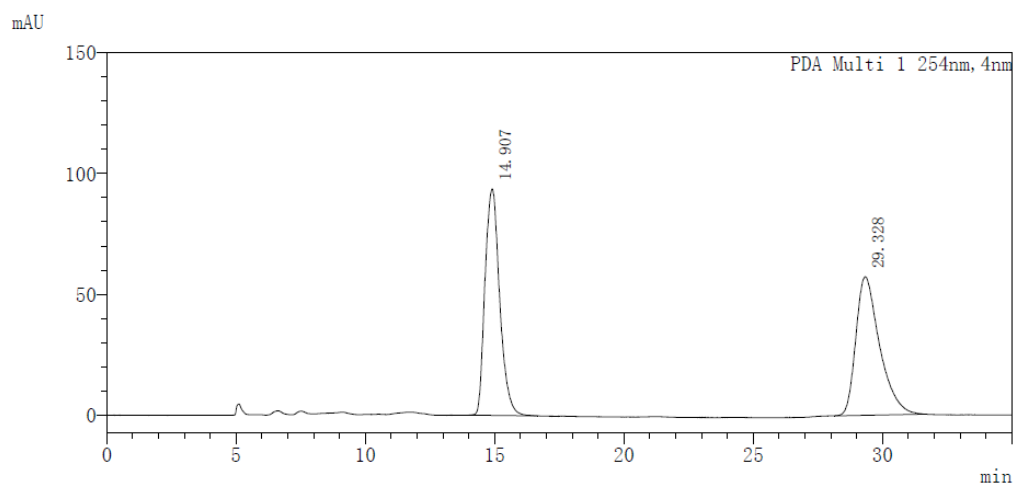


PDA Ch1 254nm

T	Hight	Area	Area%
16.090	1454650	72641019	92.326
20.761	113867	6037824	7.674

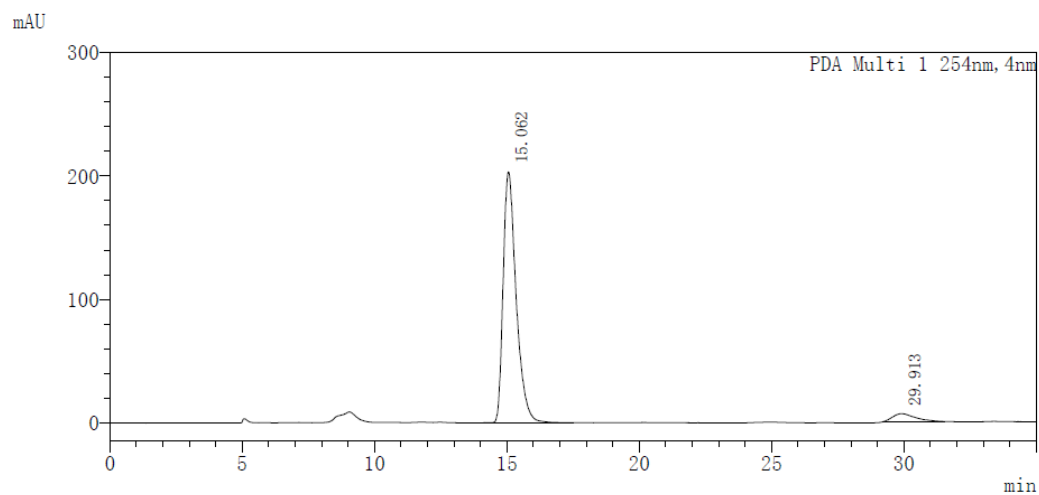


4d, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.6 mL/min. λ = 254 nm, t (major) = 15.1 min, t (minor) = 29.9 min, 95:5 er.



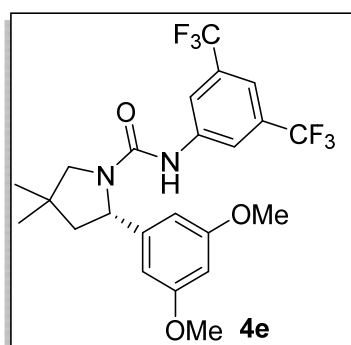
PDA Ch1 254nm

T	Hight	Area	Area%
14.907	93793	3702332	50.281
29.328	57269	3660976	49.719

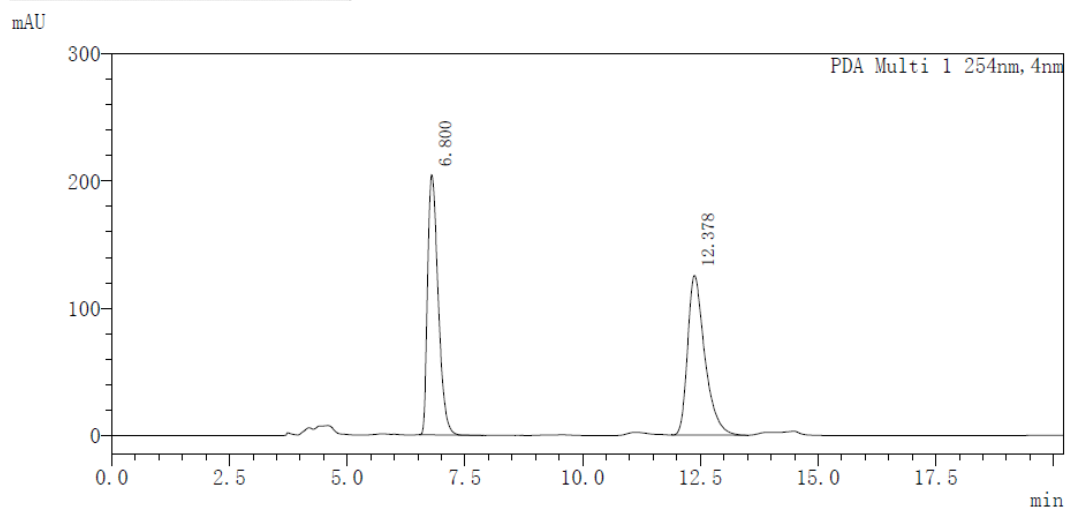


PDA Ch1 254nm

T	Hight	Area	Area%
15.062	203424	6978598	94.896
29.913	6254	375356	5.104

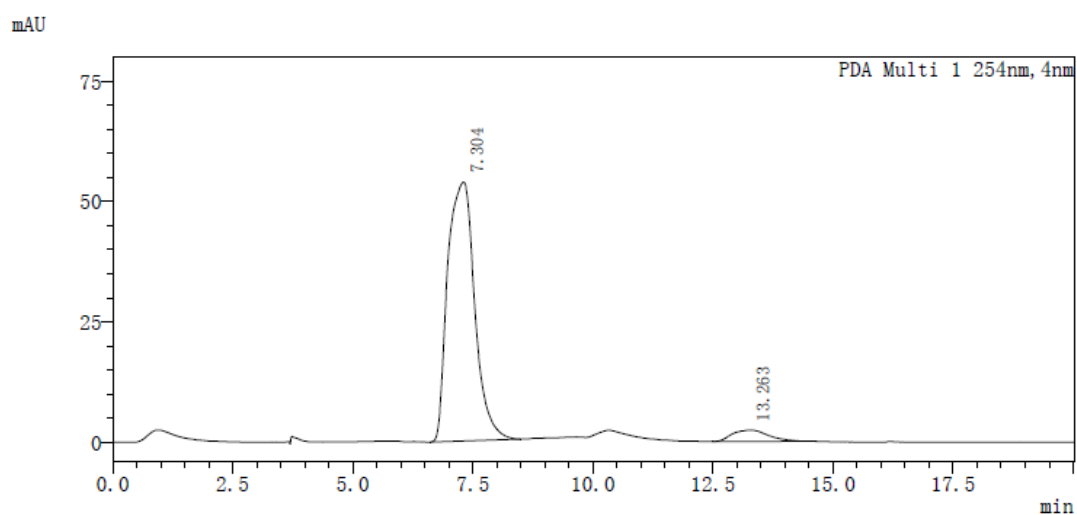


4e, HPLC condition: Chiralcel AD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.8 mL/min. λ = 254 nm, t (major) = 7.3 min, t (minor) = 13.3 min, 94.5:5.5 er.



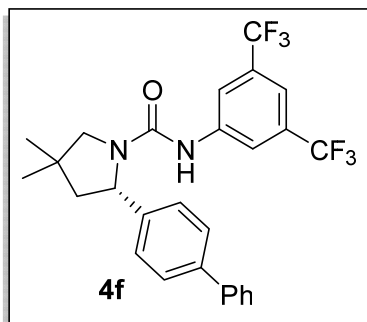
PDA Ch1 254nm

T	Hight	Area	Area%
6.800	204358	3206130	50.140
12.378	125410	3188207	49.860

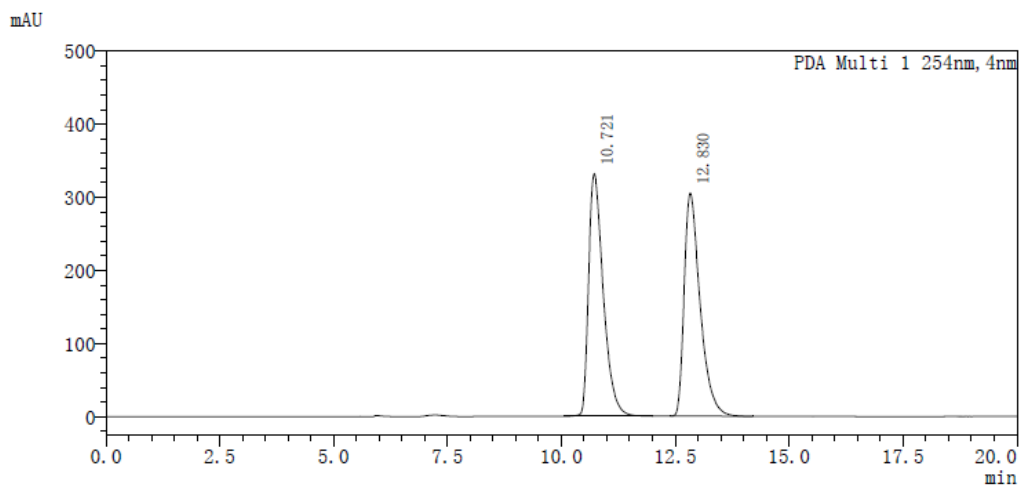


PDA Ch1 254nm

T	Hight	Area	Area%
7.304	53747	2099902	94.469
13.263	2339	122941	5.531

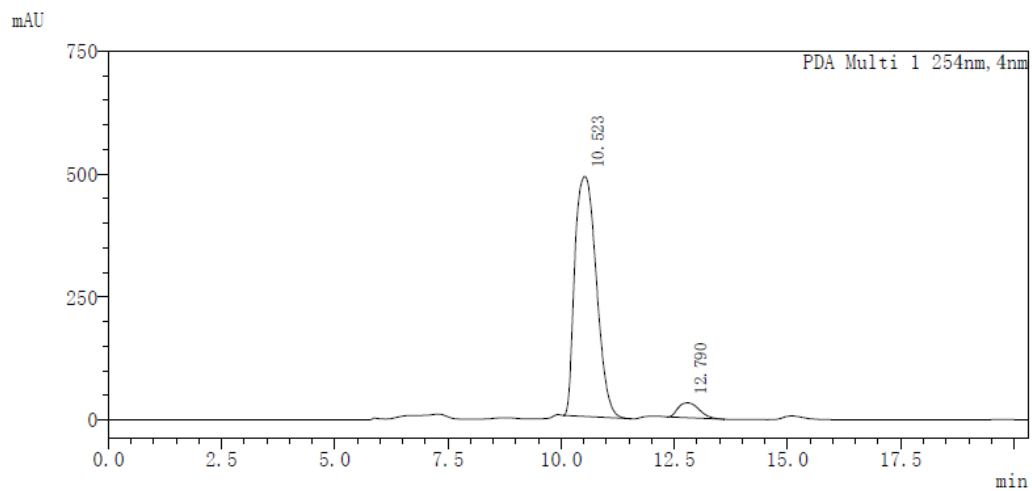


4f, HPLC condition: Chiralcel AD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.5 mL/min. λ = 254 nm, t (major) = 10.5 min, t (minor) = 12.8 min, 94:6 er.



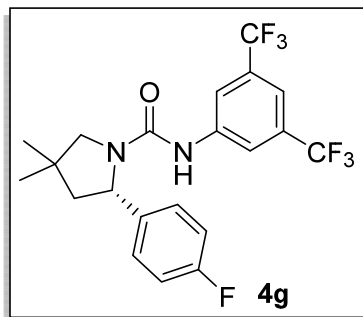
PDA Ch1 254nm

T	Hight	Area	Area%
10.721	331349	7427921	50.066
12.830	305030	7408255	49.934

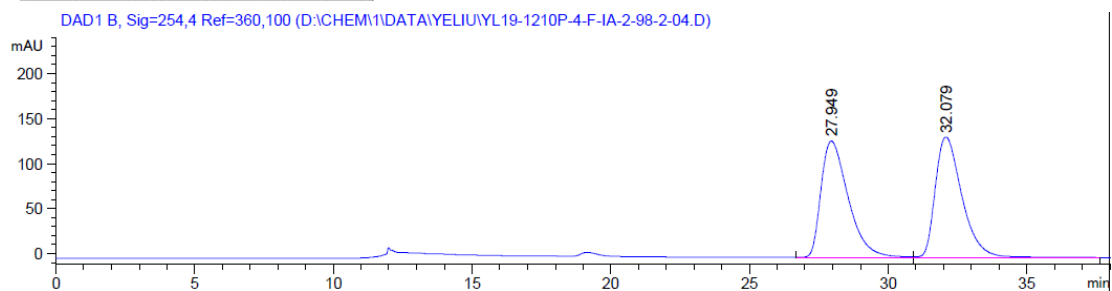


PDA Ch1 254nm

T	Hight	Area	Area%
10.523	488697	16144737	94.266
12.790	30295	982065	5.734

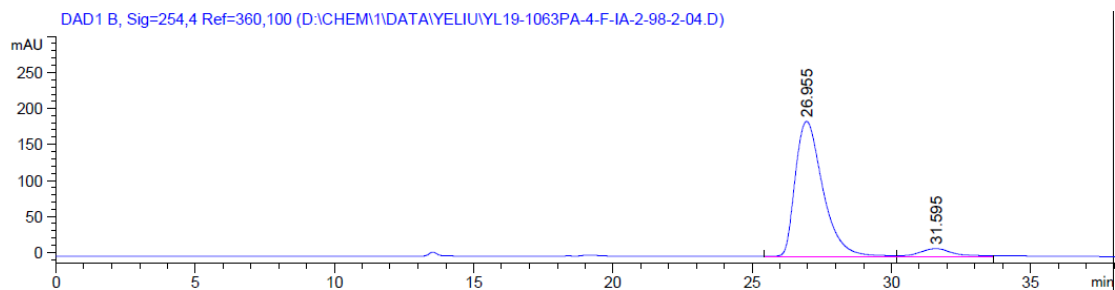


4g, HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 98/2, flow rate 0.4 mL/min. λ = 254 nm, t (major) = 26.9 min, t (minor) = 31.6 min, 94:6 er.



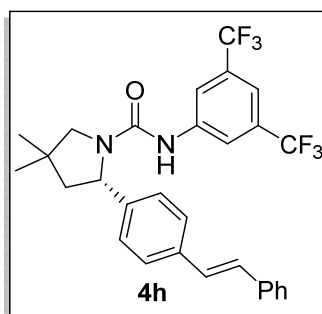
Signal 2: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	27.949	BV	1.0432	8822.51660	129.43439	49.6936
2	32.079	VB	1.0300	8931.31934	133.92444	50.3064



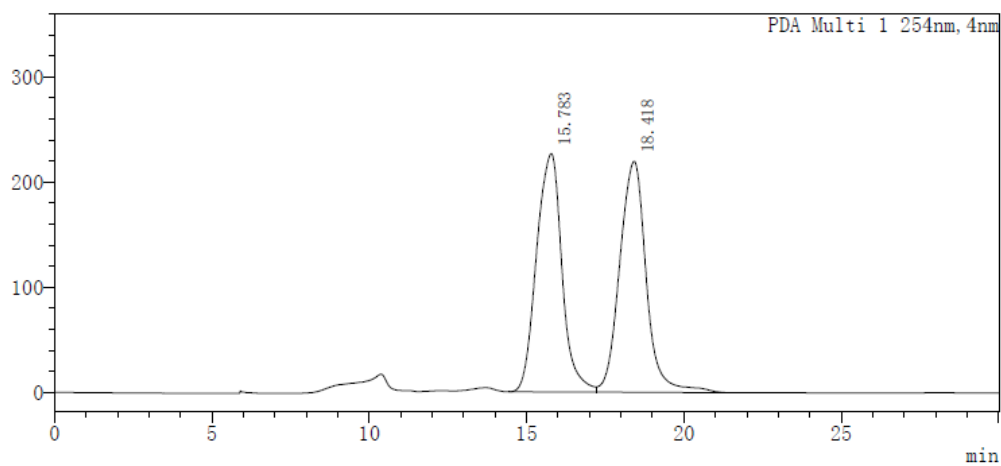
Signal 2: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	26.955	BV	1.0656	1.28801e4	187.45976	93.6796
2	31.595	VV	1.2307	868.99628	10.80079	6.3204



4h, HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.5 mL/min. λ = 254 nm, t (major) = 15.5 min, t (minor) = 18.7 min, 96:4 er.

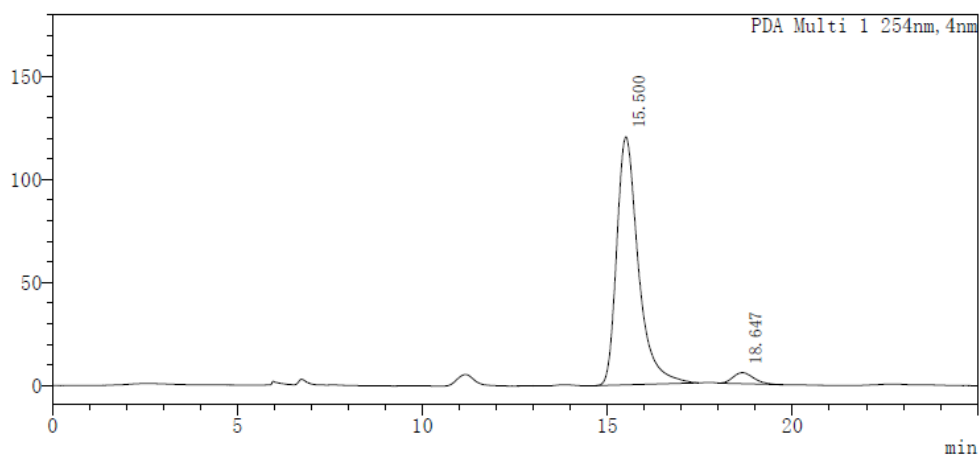
mAU



PDA Ch1 254nm

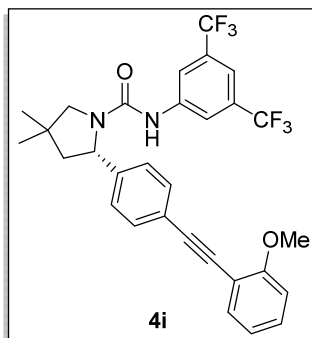
T	Hight	Area	Area%
15.783	226572	12749093	49.604
18.418	219687	12952506	50.396

mAU

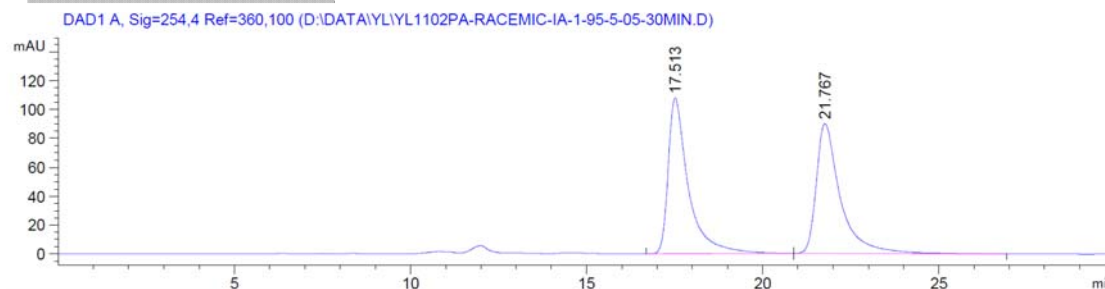


PDA Ch1 254nm

T	Hight	Area	Area%
15.500	120368	4900510	95.935
18.647	5398	207632	4.065

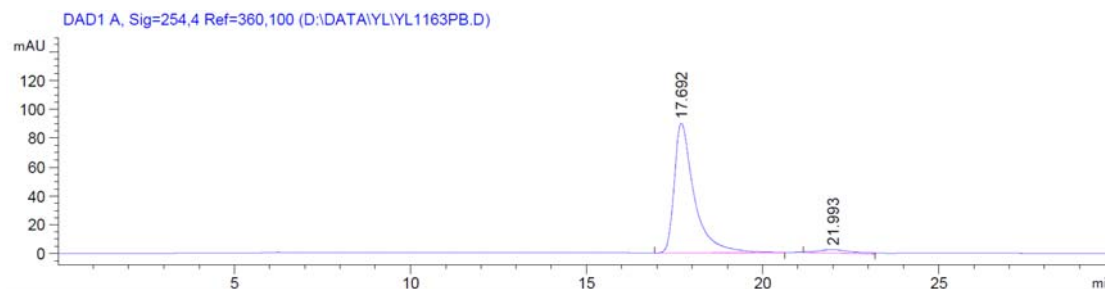


4i, HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.5 mL/min. λ = 254 nm, t (major) = 17.7 min, t (minor) = 22.0 min, 96:4 er.

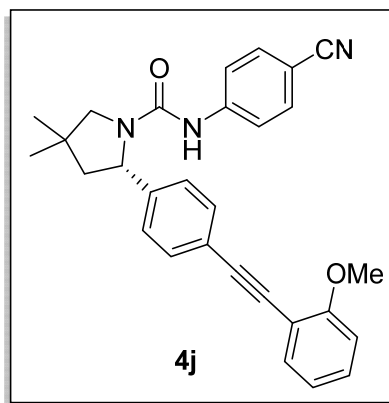


Signal 1: DAD1 A, Sig=254,4 Ref=360,100

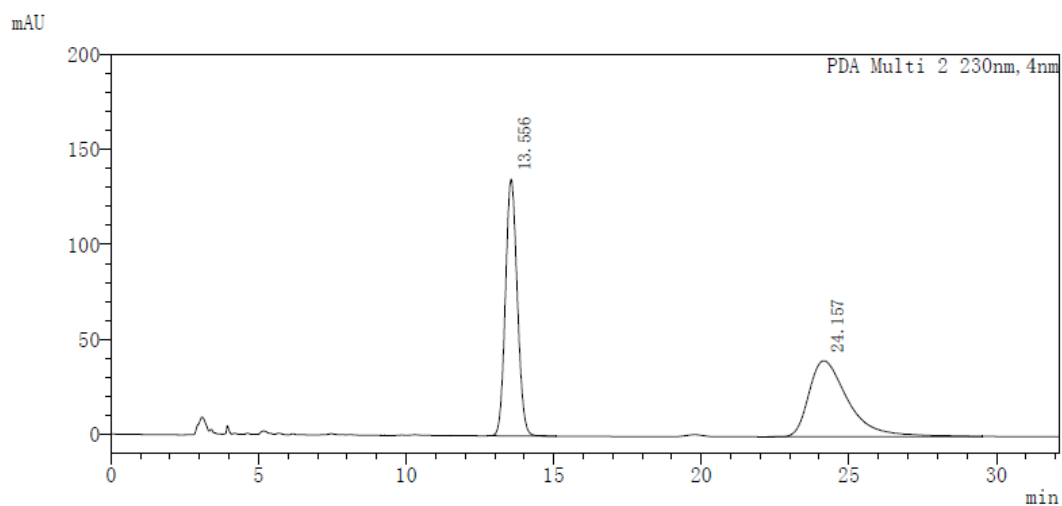
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.513	BB	0.5812	4318.60498	107.83671	50.0836
2	21.767	BB	0.6964	4304.19434	89.93429	49.9164



Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	17.692	BB	0.5762	3540.71045	89.76898	95.6155
2	21.993	MM R	1.0766	162.36003	2.51339	4.3845

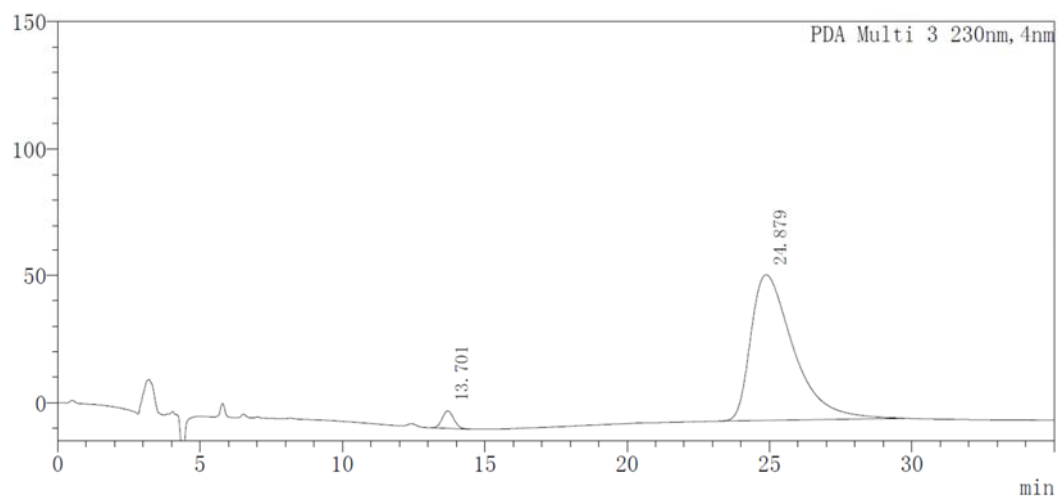


4j, HPLC condition: Chiralcel AD-H, *i*-PrOH/*n*-hexane = 75/25, flow rate 1.0 mL/min. λ = 230 nm, t (minor) = 13.7 min, t (major) = 24.9 min, 97:3 er.



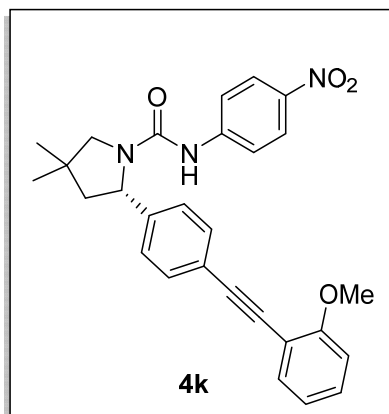
PDA Ch2 230nm

T	Hight	Area	Area%
13.556	135331	3785004	50.477
24.157	40084	3713483	49.523

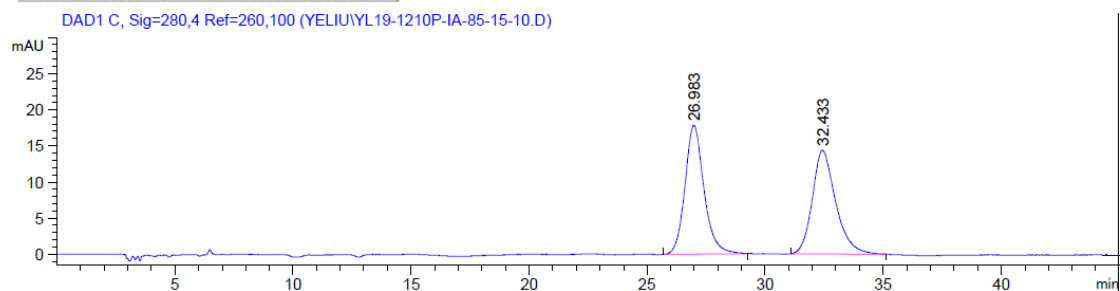


PDA Ch3 230nm

T	Hight	Area	Area%
13.701	6884	195130	3.171
24.879	57149	5958344	96.829

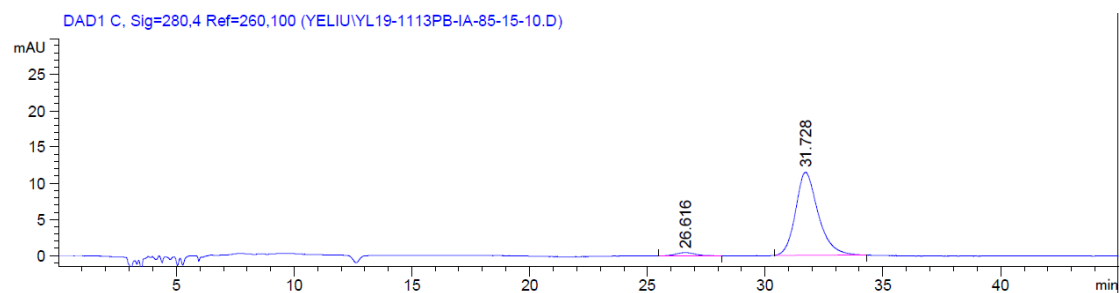


4k, HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 85/15, flow rate 1.0 mL/min. λ = 280 nm, *t*(minor) = 26.6 min, *t*(major) = 31.7 min, 96.5:3.5 er.



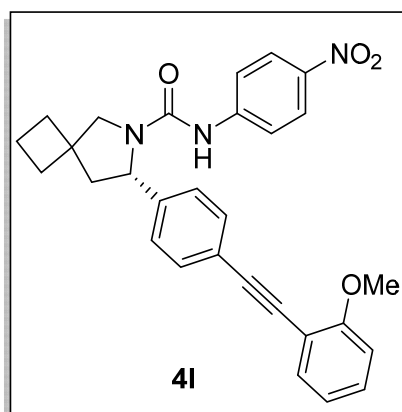
Signal 3: DAD1 C, Sig=280,4 Ref=260,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	26.983	BB	0.7927	997.63281	17.83261	50.2570
2	32.433	BB	0.9233	987.43036	14.34117	49.7430



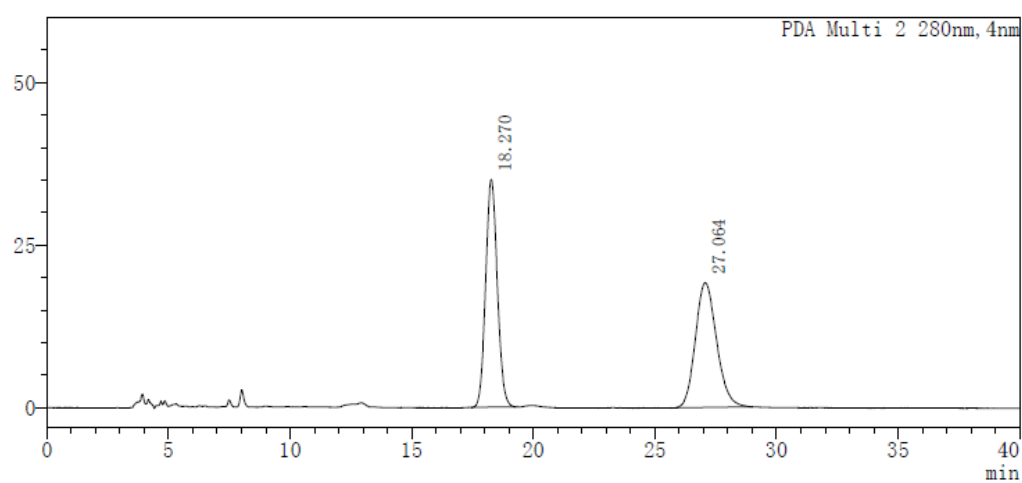
Signal 3: DAD1 C, Sig=280,4 Ref=260,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	26.616	MM R	0.9846	28.43053	4.81239e-1	3.5692
2	31.728	BB	0.9004	768.11865	11.48307	96.4308



4I, HPLC condition: Chiralcel AD-H, *i*-PrOH/*n*-hexane = 70/30, flow rate 1.0 mL/min. λ = 280 nm, t (minor) = 18.3 min, t (major) = 27.0 min, 96:4 er.

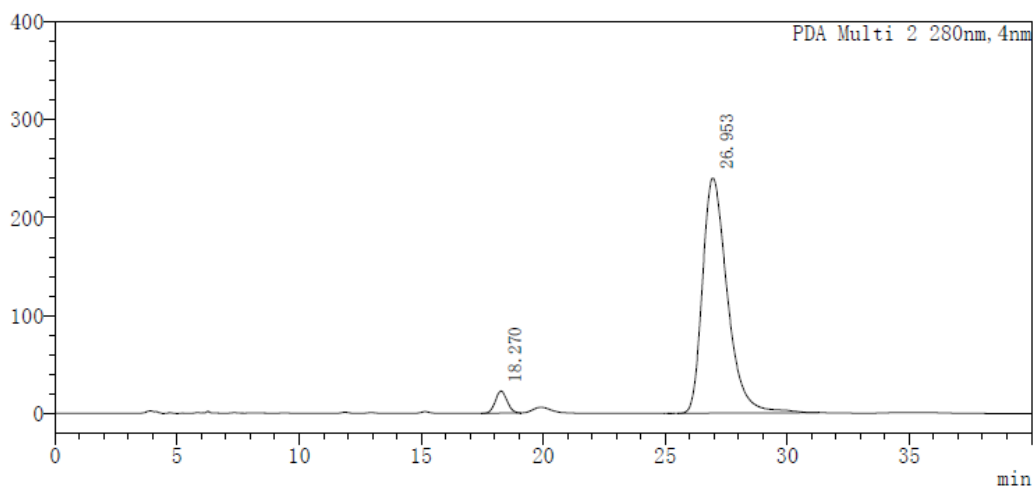
mAU



PDA Ch2 280nm

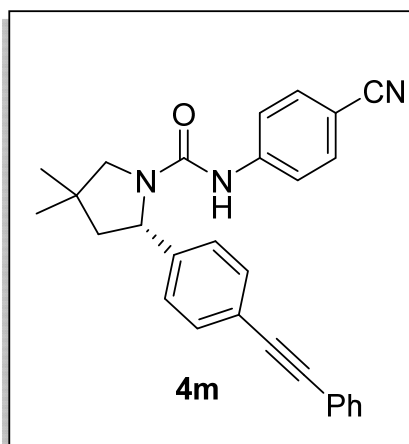
T	Hight	Area	Area%
18.270	35020	1165926	50.036
27.064	19208	1164237	49.964

mAU

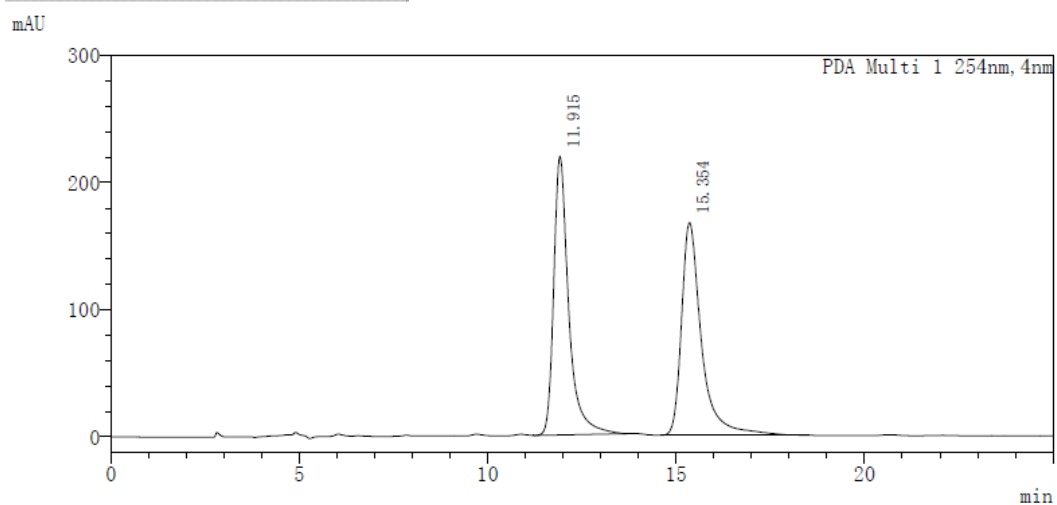


PDA Ch2 280nm

T	Hight	Area	Area%
18.270	22468	750124	4.211
26.953	240264	17062663	95.789

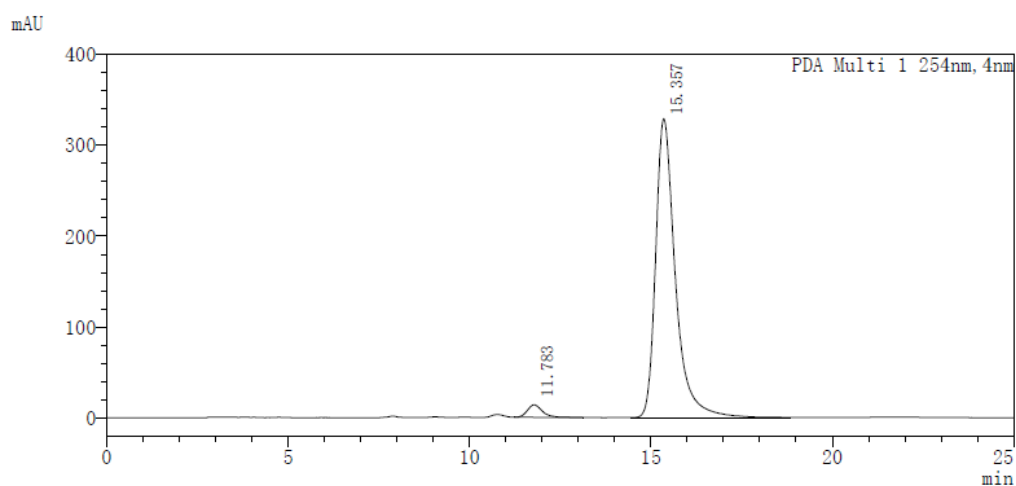


4m, HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 80/20, flow rate 1.0 mL/min. λ = 254 nm, t (minor) = 11.8 min, t (major) = 15.4 min, 97:3 er.



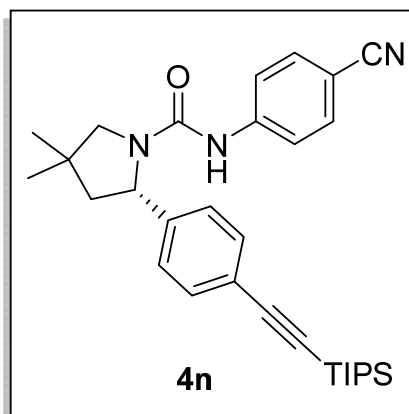
PDA Ch1 254nm

T	Hight	Area	Area%
11.915	219037	6048885	49.485
15.354	167353	6174746	50.515

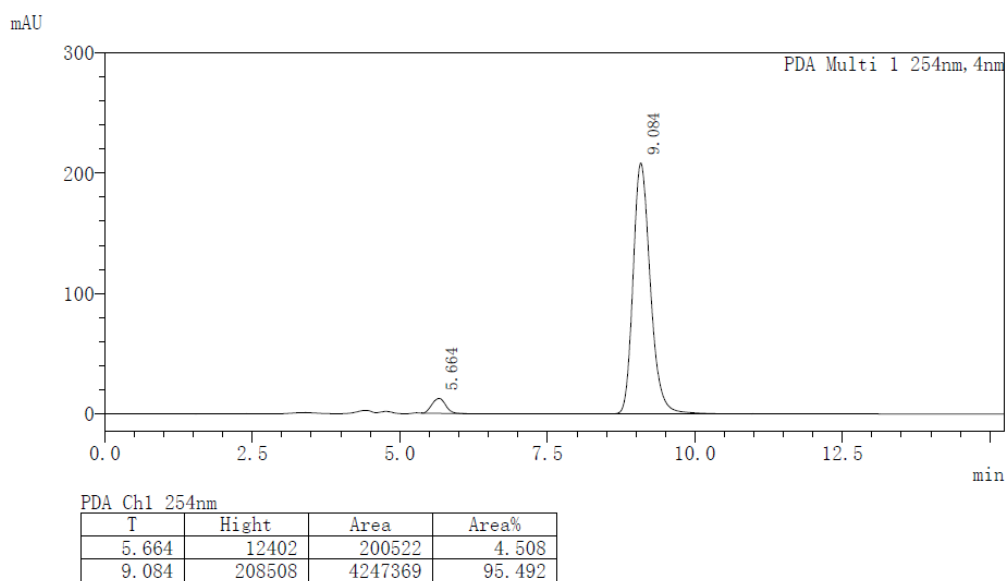
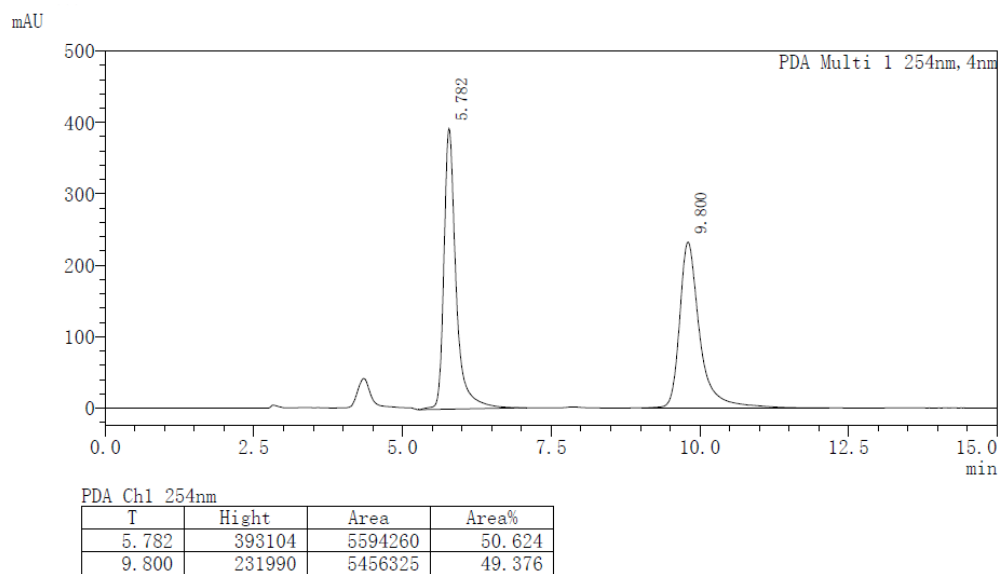


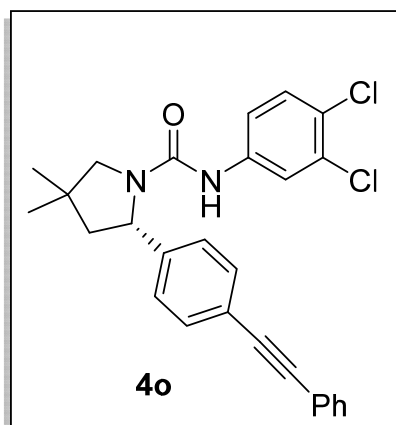
PDA Ch1 254nm

T	Hight	Area	Area%
11.783	13672	393224	3.069
15.357	328811	12418723	96.931



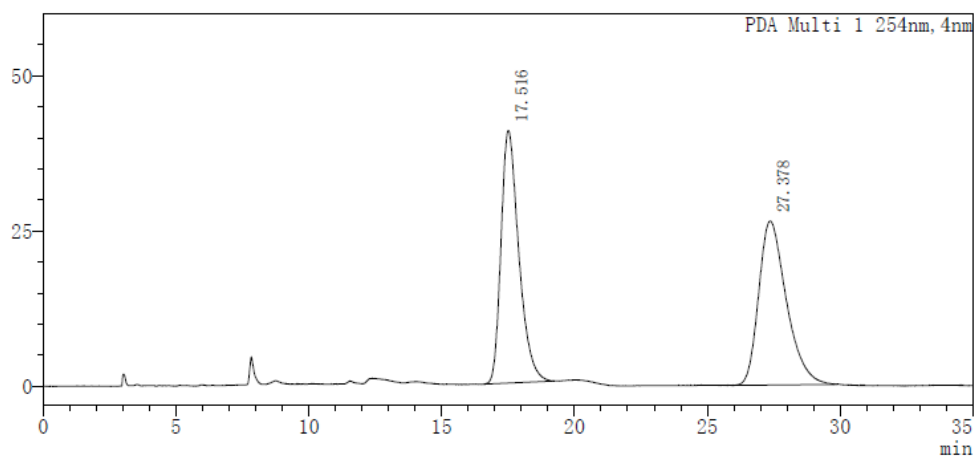
4n, HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 80/20, flow rate 1.0 mL/min. λ = 254 nm, t (minor) = 5.7 min, t (major) = 9.1 min, 95.5:4.5 er.





4o, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 1.0 mL/min. λ = 254 nm, t (major) = 17.9 min, t (minor) = 28.4 min, 95:5 er.

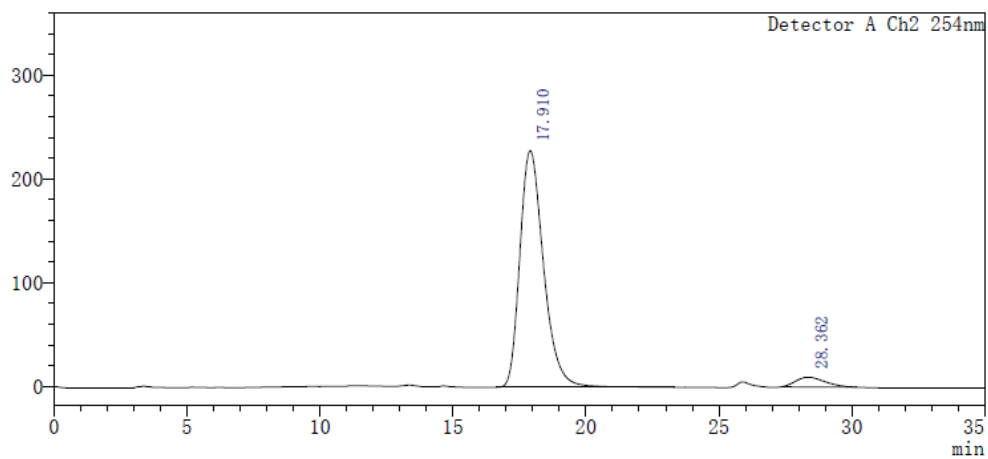
mAU



PDA Ch1 254nm

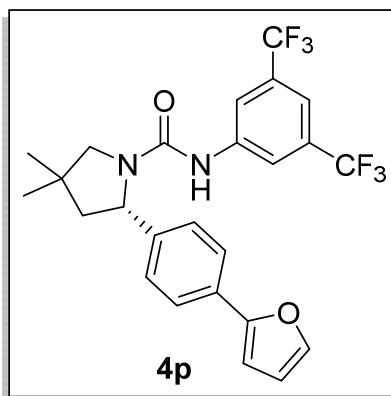
T	Hight	Area	Area%
17.516	40688	1901659	49.805
27.378	26408	1916512	50.195

mV

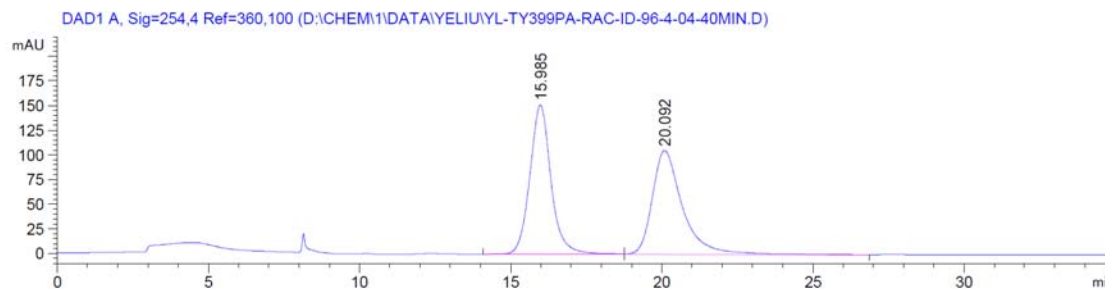


Detector A Ch2 254nm

T	Hight	Area	Area%
17.910	228130	14163127	95.182
28.362	9371	716900	4.818

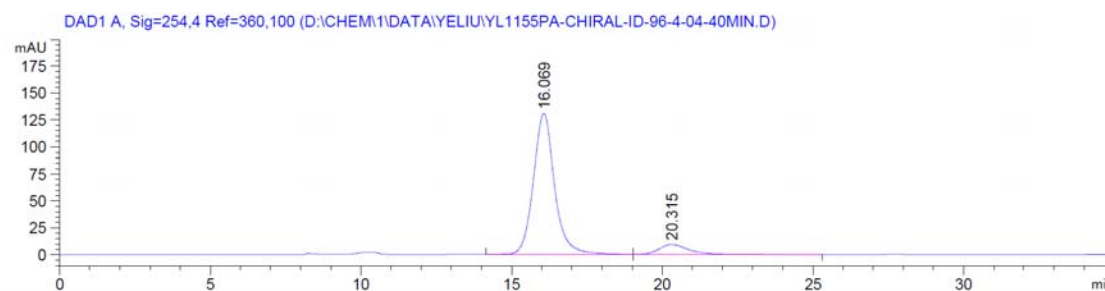


4p, HPLC condition: Chiralcel ID, *i*-PrOH/*n*-hexane = 96/4, flow rate 0.4 mL/min. λ = 254 nm, t (major) = 16.1 min, t (minor) = 20.3 min, 91:9 er.



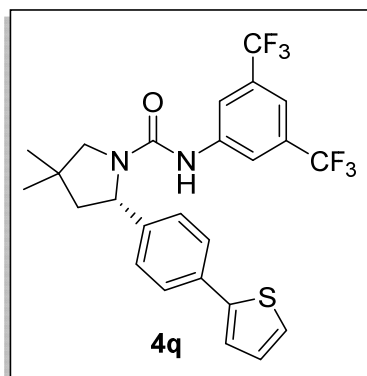
Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.985	BV	0.7424	7464.87451	151.61041	50.2034
2	20.092	VV	1.0427	7404.38623	105.50870	49.7966

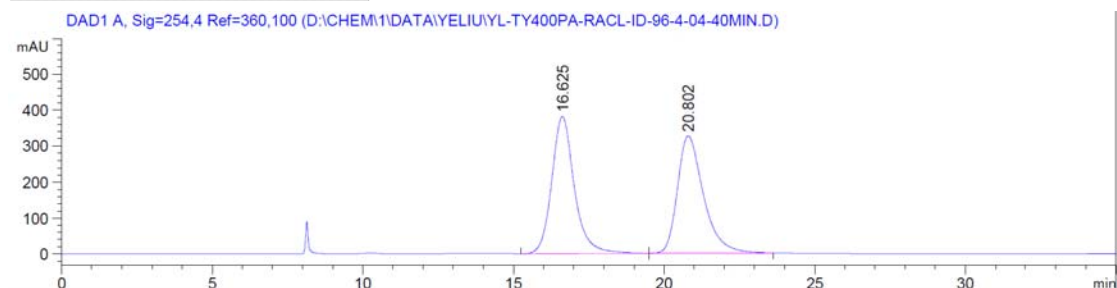


Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.069	VV	0.7419	6442.18652	130.95869	90.6113
2	20.315	VB	1.0597	667.50903	9.31883	9.3887

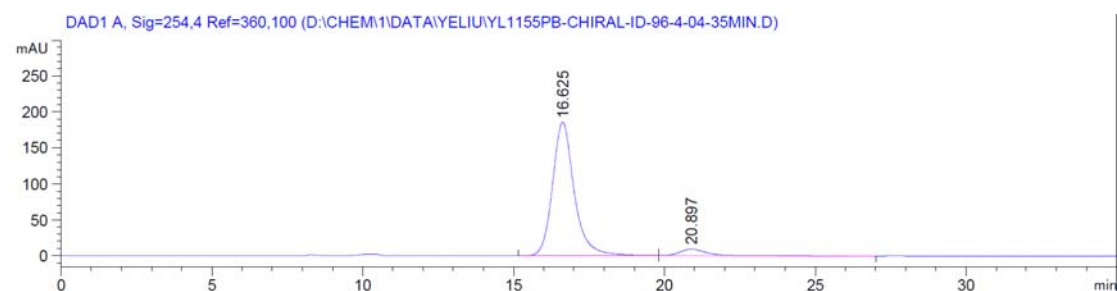


4q, HPLC condition: Chiralcel ID, *i*-PrOH/*n*-hexane = 96/4, flow rate 0.4 mL/min. λ = 254 nm, t (major) = 16.6 min, t (minor) = 20.9 min, 94:6 er.



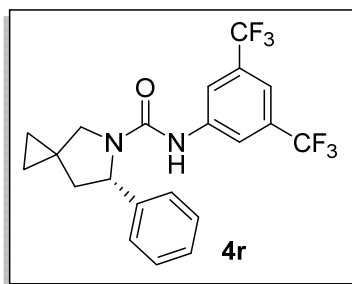
Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.625	BB	0.7900	1.99101e4	380.80334	50.0914
2	20.802	BV	0.9230	1.98374e4	325.52408	49.9086

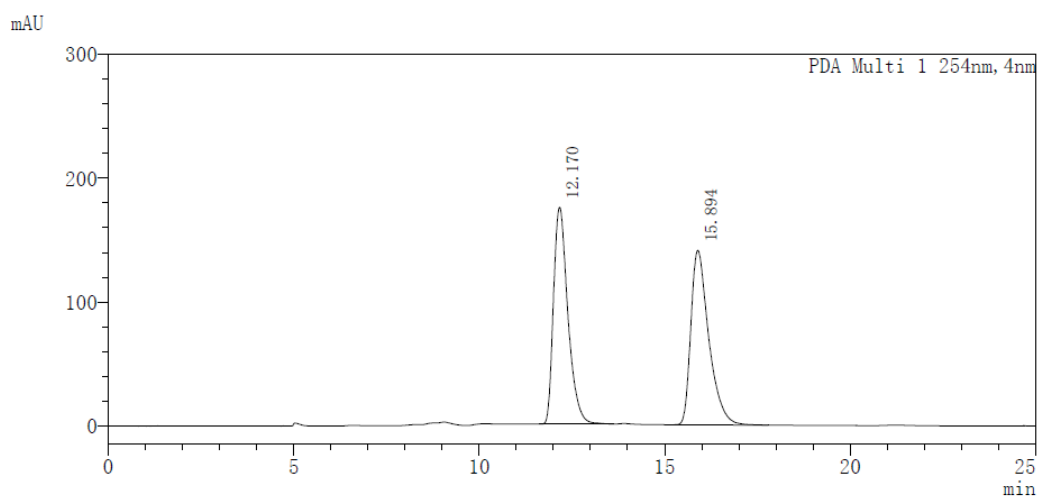


Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.625	VV	0.7628	9390.16211	185.41672	93.5521
2	20.897	VV	1.0119	647.19409	9.43900	6.4479

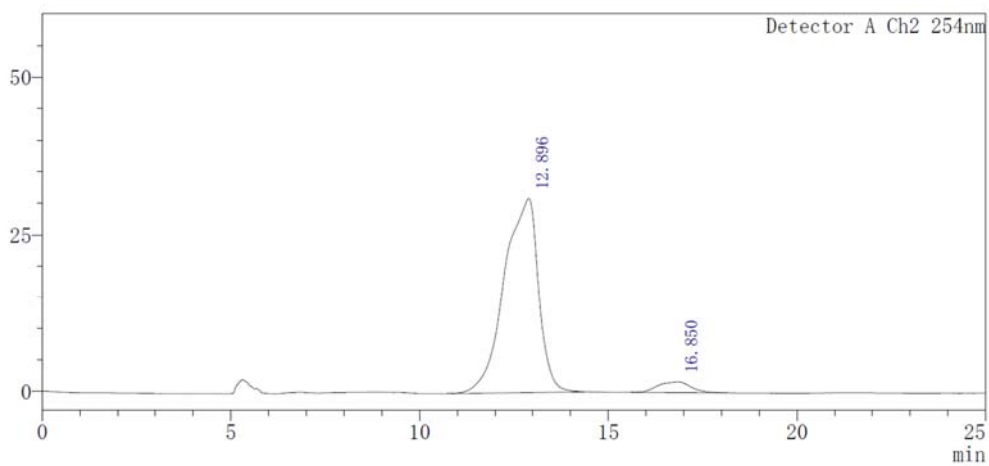


4r, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.6 mL/min. λ = 254 nm, *t*(major) = 12.3 min, *t*(minor) = 16.0 min, 95:5 er.



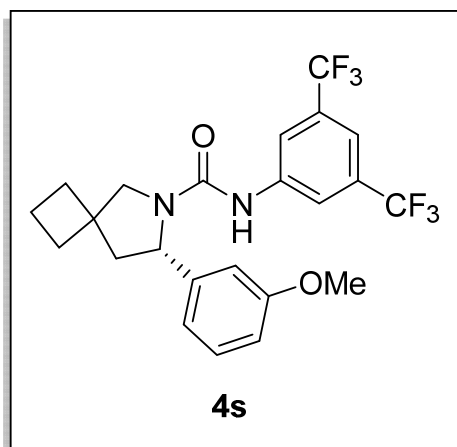
PDA Ch1 254nm

T	Hight	Area	Area%
12.170	175127	4732898	50.005
15.894	140909	4732023	49.995

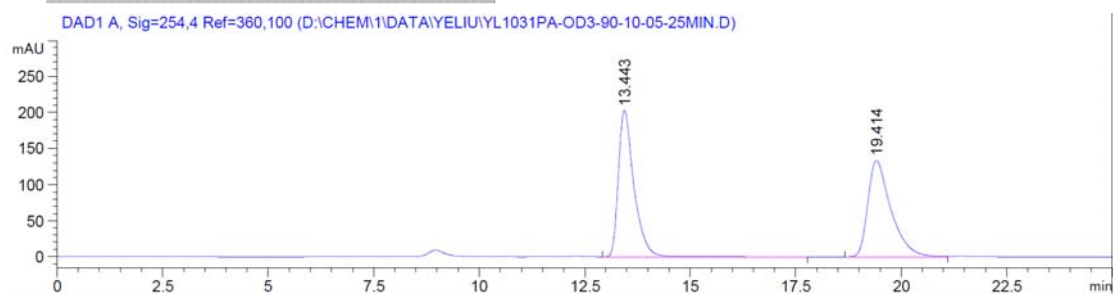


Detector A Ch2 254nm

T	Hight	Area	Area%
12.896	30962	1959794	94.906
16.850	1705	105187	5.094
			100.000

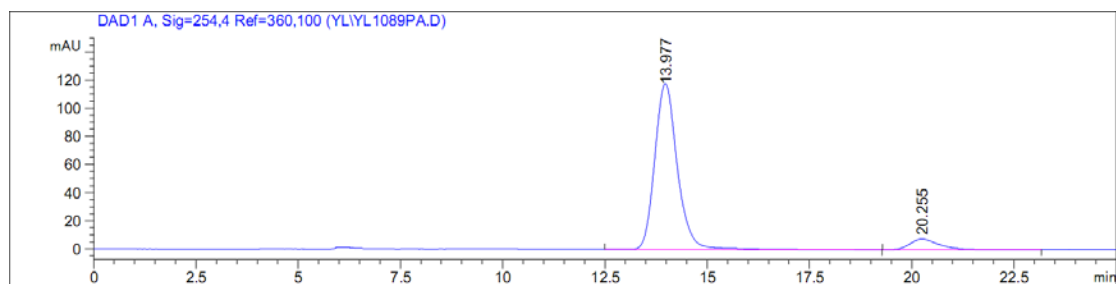


4s, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 0.5 mL/min. λ = 254 nm, t (major) = 14.0 min, t (minor) = 20.3 min, 92:8 er.



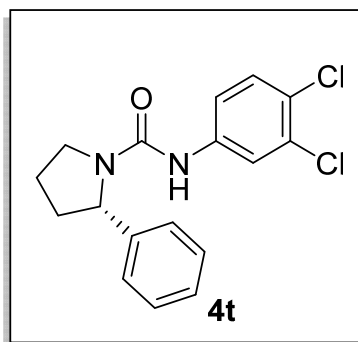
Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.443	BB	0.3836	5213.37646	202.99559	50.5273
2	19.414	BV	0.5641	5104.55713	134.09773	49.4727

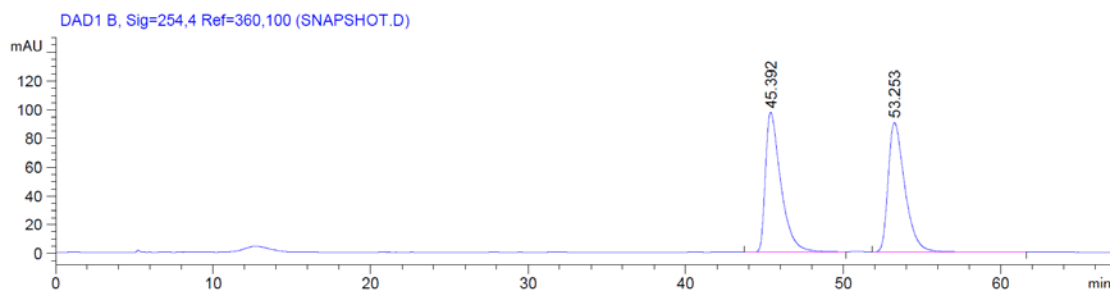


Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	13.977	BB	0.5836	4529.45410	118.05036	92.2677
2	20.255	BB	0.7440	379.58371	7.68792	7.7323

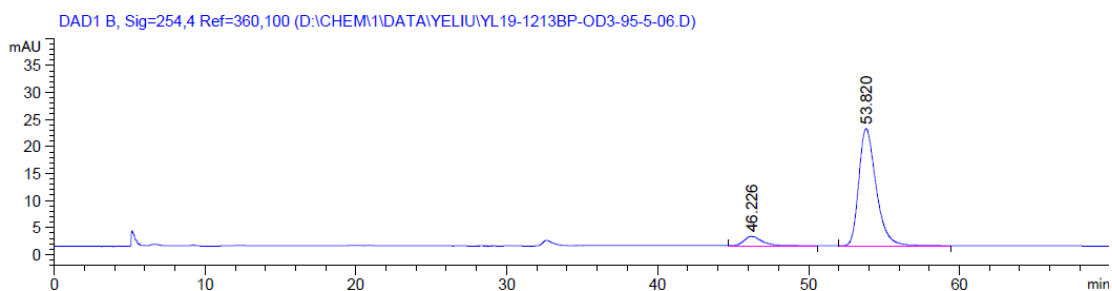


4t, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.6 mL/min. λ = 254 nm, t_r (minor) = 46.2 min, t_r (minor) = 53.8 min, 92.5:7.5 er.



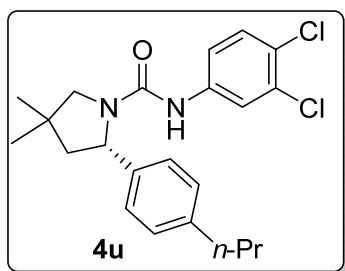
Signal 2: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	45.392	BV	1.0204	6656.81152	97.03388	49.9147
2	53.253	VB	1.1202	6679.56201	90.17438	50.0853

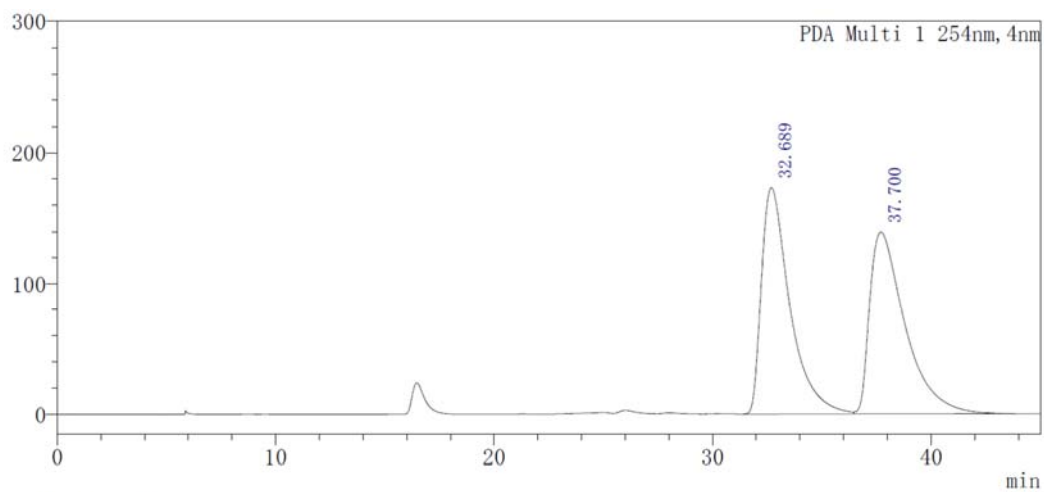


Signal 2: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	46.226	VB	1.2391	142.26128	1.71622	7.5970
2	53.820	BV	1.2158	1730.33655	21.66531	92.4030

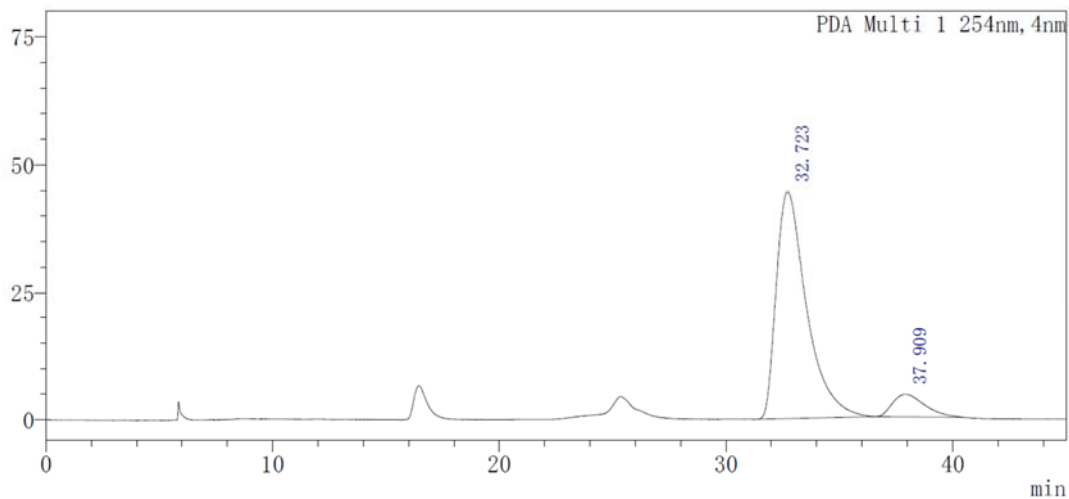


4u, Chiralcel AD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.5 mL/min. λ = 254 nm, $t(\text{major})$ = 32.7 min, $t(\text{minor})$ = 37.9 min, 90.5:9.5 er.



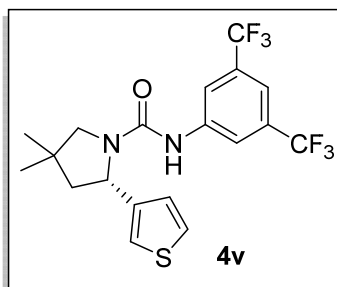
PDA Ch1 254nm

T	Hight	Area	Area%
32.689	173127	15669415	50.005
37.700	139152	15666086	49.995

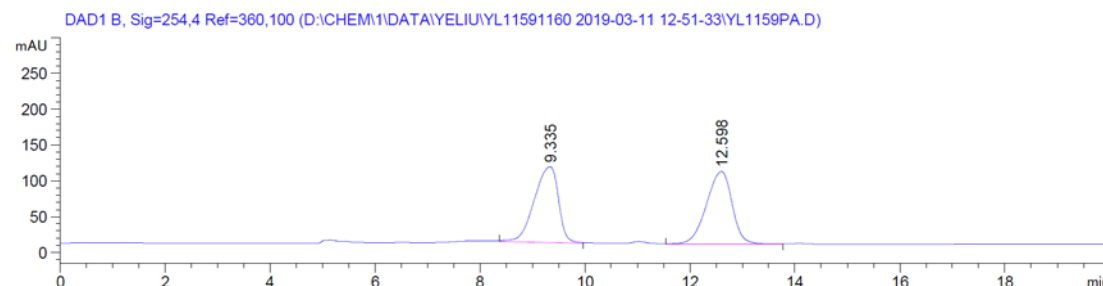


PDA Ch1 254nm

T	Hight	Area	Area%
32.723	44533	4131630	90.585
37.909	4403	429427	9.415

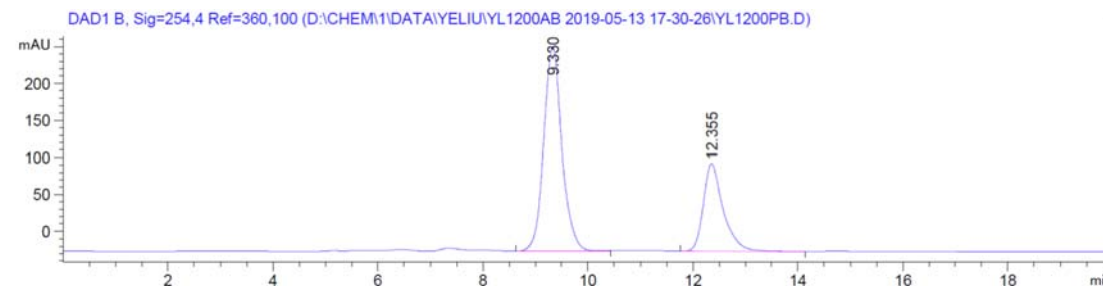


4v, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 90/10, flow rate 0.6 mL/min. λ = 254 nm, t (major) = 9.3 min, t (minor) = 12.4 min, 68:32 er.



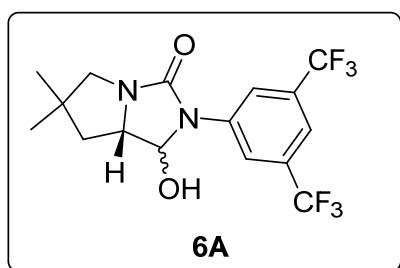
Signal 2: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.335	MM R	0.5372	3416.96558	106.00591	50.0680
2	12.598	BB	0.5299	3407.68457	100.92472	49.9320

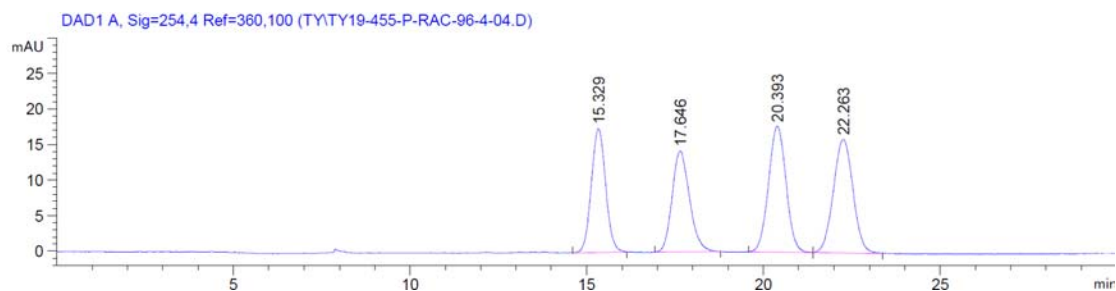


Signal 2: DAD1 B, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.330	BV	0.3698	6680.89697	276.60239	68.1099
2	12.355	BB	0.3880	3128.09131	118.48897	31.8901

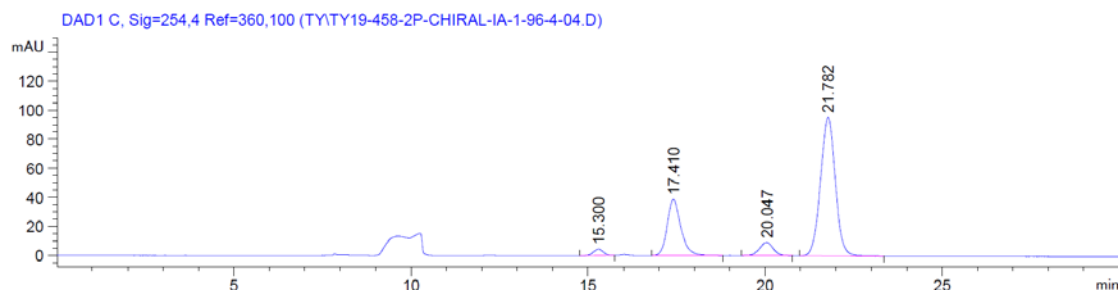


6A, HPLC condition: Chiralcel IA, *i*-PrOH/*n*-hexane = 96/4, flow rate 0.4 mL/min. λ = 254 nm, minor diastereomer: $t(\text{minor})$ = 15.3 min, $t(\text{major})$ = 17.4 min, 92:8 er; major diastereomer: $t(\text{minor})$ = 20.0 min, $t(\text{major})$ = 21.8 min, 92:8 er.



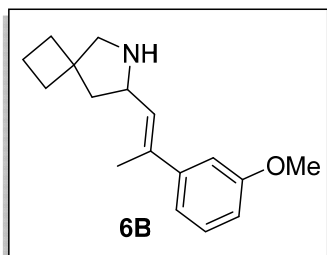
Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.329	BB	0.4334	491.75681	17.41818	22.2807
2	17.646	BB	0.5091	488.25284	14.20116	22.1220
3	20.393	BB	0.5078	611.56238	17.75667	27.7089
4	22.263	BB	0.5565	615.52234	16.01059	27.8884

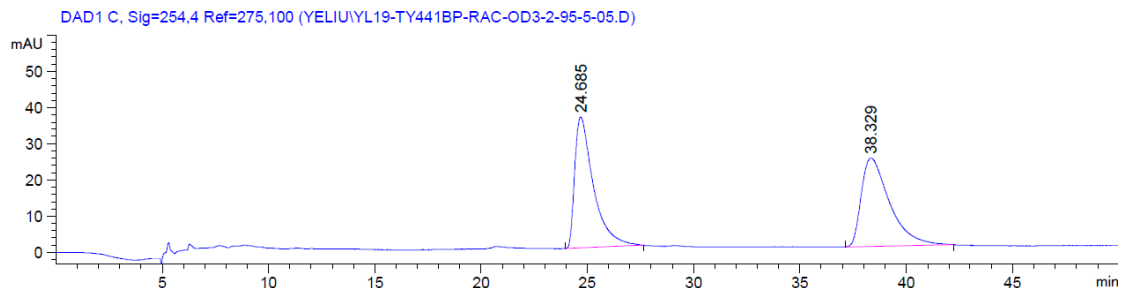


Signal 3: DAD1 C, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	15.300	BB	0.2831	89.68925	4.56707	2.1531
2	17.410	BB	0.4009	1044.49670	38.69160	25.0743
3	20.047	BB	0.3910	244.10229	9.15821	5.8600
4	21.782	BB	0.4484	2787.31519	95.53551	66.9126

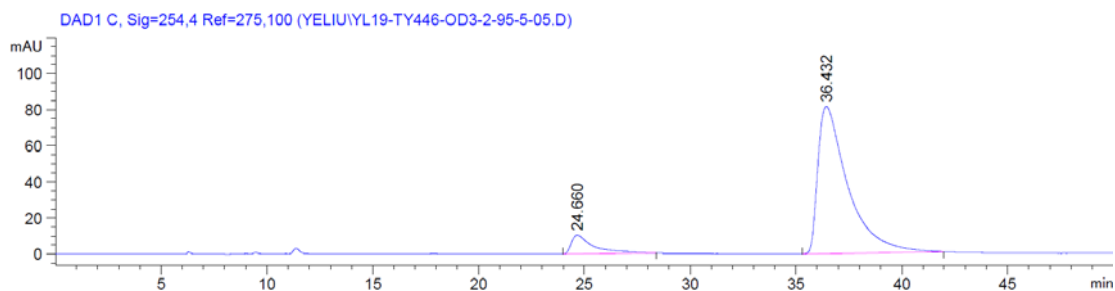


6B, HPLC condition: Chiralcel OD-3, *i*-PrOH/*n*-hexane = 95/5, flow rate 0.5 mL/min. λ = 254 nm, *t*(minor) = 24.7 min, *t*(major) = 36.4 min, 91.5:8.5 ee.



Signal 3: DAD1 C, Sig=254,4 Ref=275,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	24.685	BB	0.8860	2212.98999	36.16180	49.4496
2	38.329	BB	1.1661	2262.25659	24.39159	50.5504



Signal 3: DAD1 C, Sig=254,4 Ref=275,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	24.660	BB	0.9421	747.91583	10.31248	8.5299
2	36.432	BB	1.3472	8020.20410	81.56428	91.4701